



Mechanism of action of celastrol against rheumatoid arthritis: A network pharmacology analysis



Song Xinqiang^{a,b,*}, Zhang Yu^a, Dai Erqin^a, Du Hongtao^a, Wang Lei^{a,**}

^a Department of Biological Sciences, Xinyang Normal University, Xinyang 464000, China

^b Institute for Conservation and Utilization of Agro-bioresources in Dabie Mountains, Xinyang 464000, China

ARTICLE INFO

Keywords:

Celastrol
Rheumatoid arthritis
Ingenuity pathway analysis
Network pharmacology
Docking

ABSTRACT

Network pharmacology uses bioinformatics to broaden our understanding of drug actions and thereby to advance drug discovery. Here we apply network pharmacology to generate testable hypotheses about the multi-target mechanism of celastrol against rheumatoid arthritis. We reconstructed drug–target pathways and networks to predict the likely protein targets of celastrol and the main interactions between those targets and the drug. Then we validated our predictions of four candidate targets (IKK- β , JNK, COX-2, MEK1) by performing docking studies with celastrol. The results suggest that celastrol acts against rheumatoid arthritis by regulating the function of several signaling proteins, including MMP-9, COX-2, c-Myc, TGF- β , c-JUN, JAK-1, JAK-3, IKK- β , SYK, MMP-3, JNK and MEK1, which regulate the functions of Th1 and Th2 cells, macrophages, fibroblasts and endothelial cells in rheumatoid arthritis. Celastrol is predicted to affect networks involved mainly in cancer, connective tissue disorders, organismal injury and abnormalities, tissue development, cell death and survival. This network pharmacology strategy may be useful for discovery of multi-target drugs against complex diseases.

1. Introduction

Rheumatoid arthritis is an immune-related disease that generally gives rise to continuous joint destruction, decreased life expectancy and working ability, disability, and even elevated mortality [1–3]. The disease is most commonly treated using synthetic and biological disease-modifying anti-rheumatic drugs (DMARDs) [4,5]. These drugs cannot cure the disease, however, and they often cause severe side effects, such as reduced immunity and cancer. Moreover, biological DMARDs place an extreme financial burden on patients without showing correspondingly high efficacy [5–7].

Traditional Chinese medicine may be able to offer more cost-effective alternative treatments against rheumatoid arthritis [8]. For centuries, the principal active ingredient of *Tripterygium wilfordii* Hook.f., called celastrol, has been used in traditional Chinese medicine to treat inflammation and autoimmune diseases including rheumatoid arthritis, systemic lupus erythematosus, nephritis, and asthma [9,10]. Celastrol shows anti-inflammatory and immunomodulatory activities, as well as pro-apoptotic effects in many types of cancer, such as ovarian and pancreatic cancers, myeloma, myeloid leukemia, and thyroid carcinoma [11–22]. However, how celastrol exerts therapeutic effects on patients with rheumatoid arthritis is unclear.

Herbal medicines such as celastrol feature multiple components and multiple targets, and analyzing this complexity can be achieved through systems pharmacology as applied in network pharmacology analysis. Systems pharmacology is an emerging field of pharmacology which utilizes network analysis of drug action as one of its approaches and network pharmacology is a method by integrating systems approaches, computational and experimental methods to illuminate the molecular mechanisms of drug [23]. Because network pharmacology can provide a good understanding of the principles of network theory and systems biology, it has been considered to be the next paradigm in drug discovery [24].

In the present study, we explored potential mechanisms of action of celastrol against rheumatoid arthritis using an integrated systems pharmacology approach. First, we identified potential molecular targets of celastrol and the interaction pathways in which those targets play roles. Then we examined overlap in those pathways and the networks they form. We also performed docking studies to predict the interactions that allow celastrol to bind to its predicted targets. Our results may help understanding the mechanisms celastrol treats rheumatoid arthritis and, more generally, to discover natural products against complex diseases.

* Correspondence to: X. Song, Institute for Conservation and Utilization of Agro-bioresources in Dabie Mountains, Xinyang 464000, China.

** Corresponding author.

E-mail addresses: xqsong2012@126.com (X. Song), wangleibio@126.com (L. Wang).

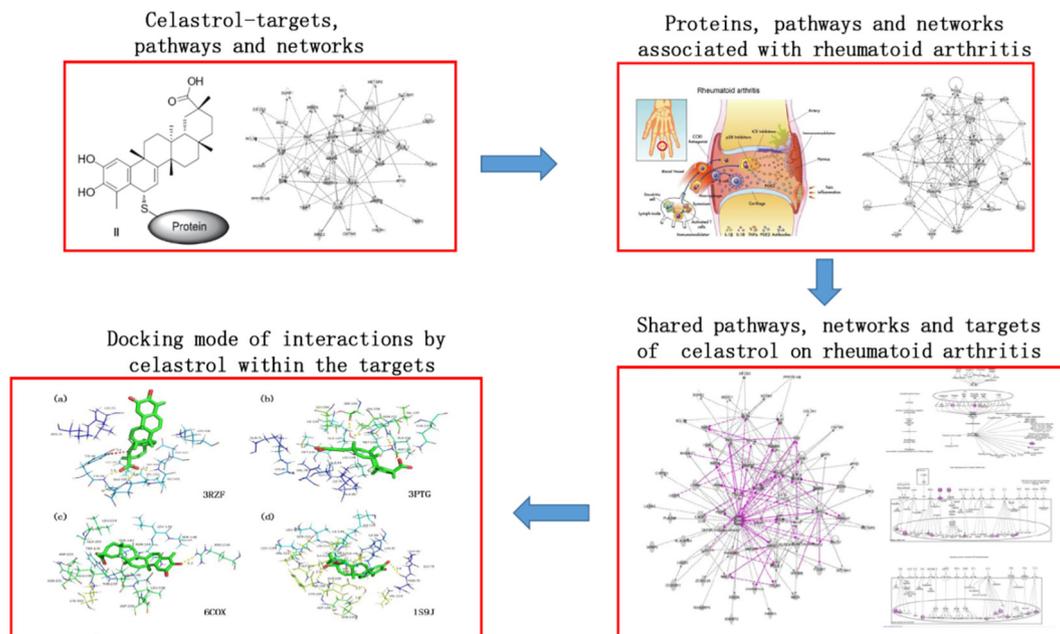


Fig. 1. Workflow in the network pharmacology approach.

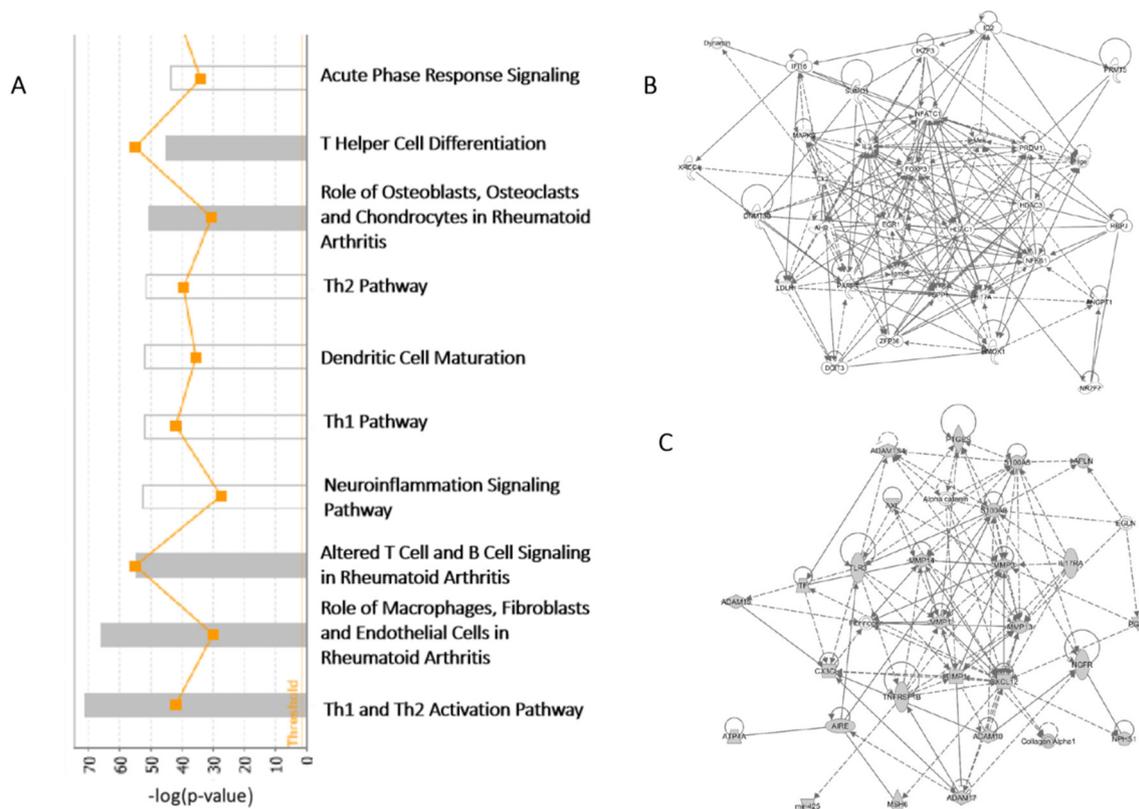


Fig. 2. Pathways and networks associated with rheumatoid arthritis. (A) Pathways formed by proteins associated with rheumatoid arthritis. (B–C) Representative networks formed by proteins associated with rheumatoid arthritis.

2. Materials and methods

2.1. Data preparation

Rheumatoid arthritis-related genes were obtained from the National Center for Biotechnology Information (<https://www.ncbi.nlm.nih.gov/>) using the term “rheumatoid arthritis”, followed by filtering with the

term “*Homo sapiens*”. Potential targets of celastrol were obtained in online resources in systems pharmacology (<http://lsp.nwu.edu.cn/index.php>) and PubChem (<https://pubchem.ncbi.nlm.nih.gov/>).

A total of 25 human proteins likely targeted by celastrol were obtained from TCMSP (Traditional Chinese Medicine Systems Pharmacology Database and Analysis Platform, <http://lsp.nwu.edu.cn/tcmsp.php>) and the PubChem database. Their symbols were uploaded

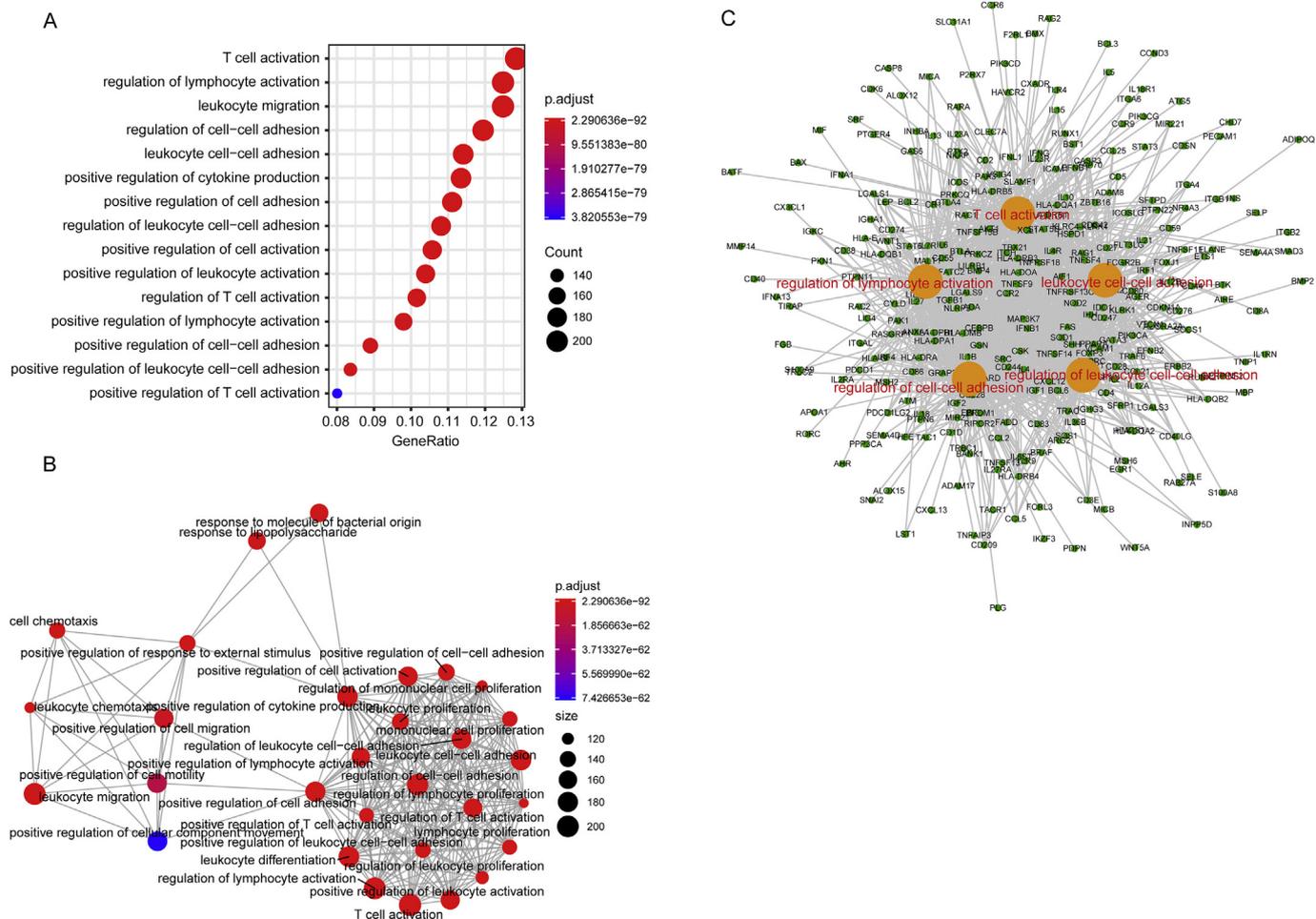


Fig. 3. Gene Ontology (GO) enrichment and network analysis of RA target genes. (A) Top 15 functionally enriched biological processes with corresponding adjusted *p*-values analyzed by clusterProfiler, which are displayed in a dot plot. The color scales indicated the different thresholds of adjusted *p*-values, and the sizes of the dots represented the gene count of each term. (B) Interaction networks between enriched biological processes analyzed by enrichMap in the clusterProfiler package. The color scales indicated different thresholds of adjusted *p*-values, and the sizes of the dots represented the gene count of each term. (C) Sub-network showing important genes in top 5 GO term and RA target genes. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

into IPA, and 111 pathways and 3 networks were assembled.

2.2. Prediction of pathways and networks affected by celestrol

Pathways and networks were built by Ingenuity Pathway Analysis (IPA, www.ingenuity.com) based on the functions of the human genes related to rheumatoid arthritis and the potential celestrol targets. Enrichment Analysis of RA Target Gene Ontology (GO) Enrichment and Network were performed by R (R 3.6.0 for Windows) and Cytoscape 3.6.1 (<http://www.cytoscape.org>).

Pathways and networks were ranked according to the amounts of the molecules participating in pathways and networks, respectively. Pathways and networks shared by targets related to rheumatoid arthritis and the potential celestrol targets were identified using the “Compare” module within IPA.

2.3. Binding of celestrol to predicted targets

Docking studies were performed with selected targets using Autodock4.2 based on the crystal structures of the targets as deposited in the RCSB Protein Data Bank (<http://www.pdb.org/pdb/home/home.do>). A CHARMM force field was employed, and hydrogen atoms were added to the proteins. The binding site was defined as a sphere encompassing protein residues within 12 Å of the original ligand. Default

values were used for other parameters, and Genetic Algorithm runs were performed for each ligand. The protocol of the integrated systems pharmacology approach is described in Fig. 1.

3. Results

3.1. Rheumatoid arthritis-related gene pathways and networks

A total of 1175 human genes associated with rheumatoid arthritis were identified in the GenBank database, and the encoded proteins were assembled into a set of 339 pathways and 25 networks using IPA. These pathways involve primarily activation of Th1 and Th2 cells, macrophages and fibroblasts in rheumatoid arthritis; altered T cell and B cell signaling in the disease and signaling pathways during the acute phase of the disease. The networks involve mainly cell movement, immune cell trafficking, hematological system development and function, inflammatory response, connective tissue disorders, organismal injury and abnormalities, as well as cell-to-cell signaling and interactions (Fig. 2). Gene Ontology (GO) Enrichment and Network Analysis showed that T cell activation, regulation of lymphocyte activation, leukocyte migration, regulation of cell-cell adhesion and leukocyte cell-cell adhesion covered the top 3 functions of RA target proteins (Fig. 3).

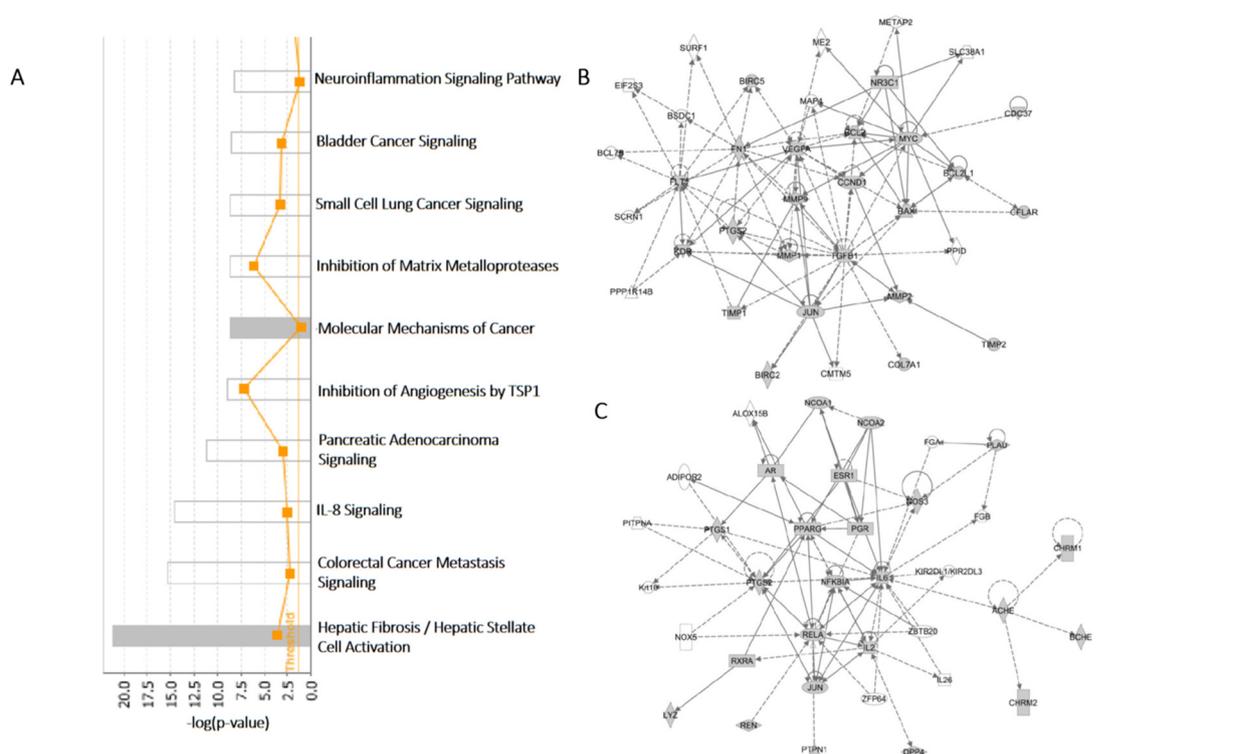


Fig. 4. Pathways and networks by predicted celestrol targets. (A) Pathways involving predicted celestrol targets. (B–C) Representative networks assembled by predicted celestrol targets.

3.2. Networks involving celestrol targets and their functions

The predicted celestrol targets participate primarily in such pathways as hepatic fibrosis, hepatic stellate cell activation, IL-8 signaling, and colorectal cancer metastasis signaling. These networks are involved mainly in cancer, connective tissue disorders, organismal injury and abnormalities, tissue development, cell death and survival, as well as endocrine system development and function (Fig. 4).

3.3. Overlapping networks and special celestrol targeted proteins

228 pathways were identified by “Canonical Pathway” and “Networks” modules of IPA, and 3 shared networks were obtained between the set of predicted celestrol targets and the set of proteins encoded by rheumatoid arthritis-associated genes. These overlapping pathways involve primarily activation of Th1 and Th2 cells, macrophages, fibroblasts and endothelial cells in rheumatoid arthritis. The networks involve primarily cancer, connective tissue disorders, organismal injury and abnormalities, tissue development, cell death and survival.

Based on the canonical pathways and networks, we predicted the following proteins to be the direct targets of celestrol in rheumatoid arthritis: MMP-9, COX-2, c-Myc, TGF- β , c-JUN, JAK-1, JAK-3, IKK- β , SYK, MMP-3, JNK, and MEK1 (Fig. 5).

3.4. Binding mode

Docking studies were performed between celestrol and the following selected potential targets (Fig. 6): IKK- β (PDB: 3RZF), JNK (PDB: 3PTG), COX-2 (PDB: 6COX) and MEK1 (PDB: 1S9J). These potential targets were chosen because they are high-degree nodes in rheumatoid arthritis-associated networks potentially affected by celestrol. High-degree nodes often play more important network roles than low-degree nodes.

The analysis in Fig. 6a predicts that celestrol binds to 3RZF by

forming a stable hydrophobic interaction with a binding pocket consisting of GLY-102, GLU-100, CYS-99, TYR-98, LYS-106, ASP-103, GLY-101, VAL-152, ILE-165, ARG-31 and LEU-21. A $\pi - \pi$ interaction is predicted to form between the benzene ring of celestrol and TYR-98, as well as three H-bonds between the oxygen atoms on celestrol and GLY-102 (length: 1.9 Å), GLU-100 (2.2 Å) and CYS-99 (1.8 Å). These interactions anchor celestrol to the binding site in 3RZF.

The analysis in Fig. 6b predicts that celestrol binds tightly to a highly hydrophobic pocket of 3PTG through stable hydrophobic interactions with 19 residues, including LEU-206, ILE-124, SER-193, VAL-196, VAL-197, ASN-152, and ALA-151. The analysis in Fig. 5c predicts that celestrol binds compactly to a pocket in 6COX consisting of 16 residues, including ASP-239, THR-237, ASP-229, TRP-139, ASN-231, GLU-236, LEU-145, SER-146, and ARG-26. Celestrol is also predicted to form an H-bond with ARG-216 (2.2 Å). In Fig. 5d, celestrol is predicted to interact with 1S9J via PHE-209, ASN-78, ILE-139, ILE-99, LYS-97, GLY79, VAL-224, MET-143, and ILE-141. Celestrol is also predicted to form H-bonds with PHE-209 (length: 2.8 Å) and ASN-78 (3.4 Å).

These docking studies provide evidence how celestrol binds to its targets, which may be useful for basic understanding the mechanism of drug action.

4. Discussion

Traditional drug discovery is largely based upon the paradigm of ‘one molecule, one target, one disease’, but there is a growing recognition that drugs work by targeting multiple proteins [25–31]. In addition, biological pathways and networks are abundant and robust, so affecting only a single target can easily fail to produce the desired therapeutic effects [17,32–40]. Therefore, the development of models that can predict multiple drug-target interactions may hold the key to future success in drug discovery against complex diseases such as rheumatoid arthritis.

In the present report, we integrated information from publicly available databases to predict interactions between celestrol and its

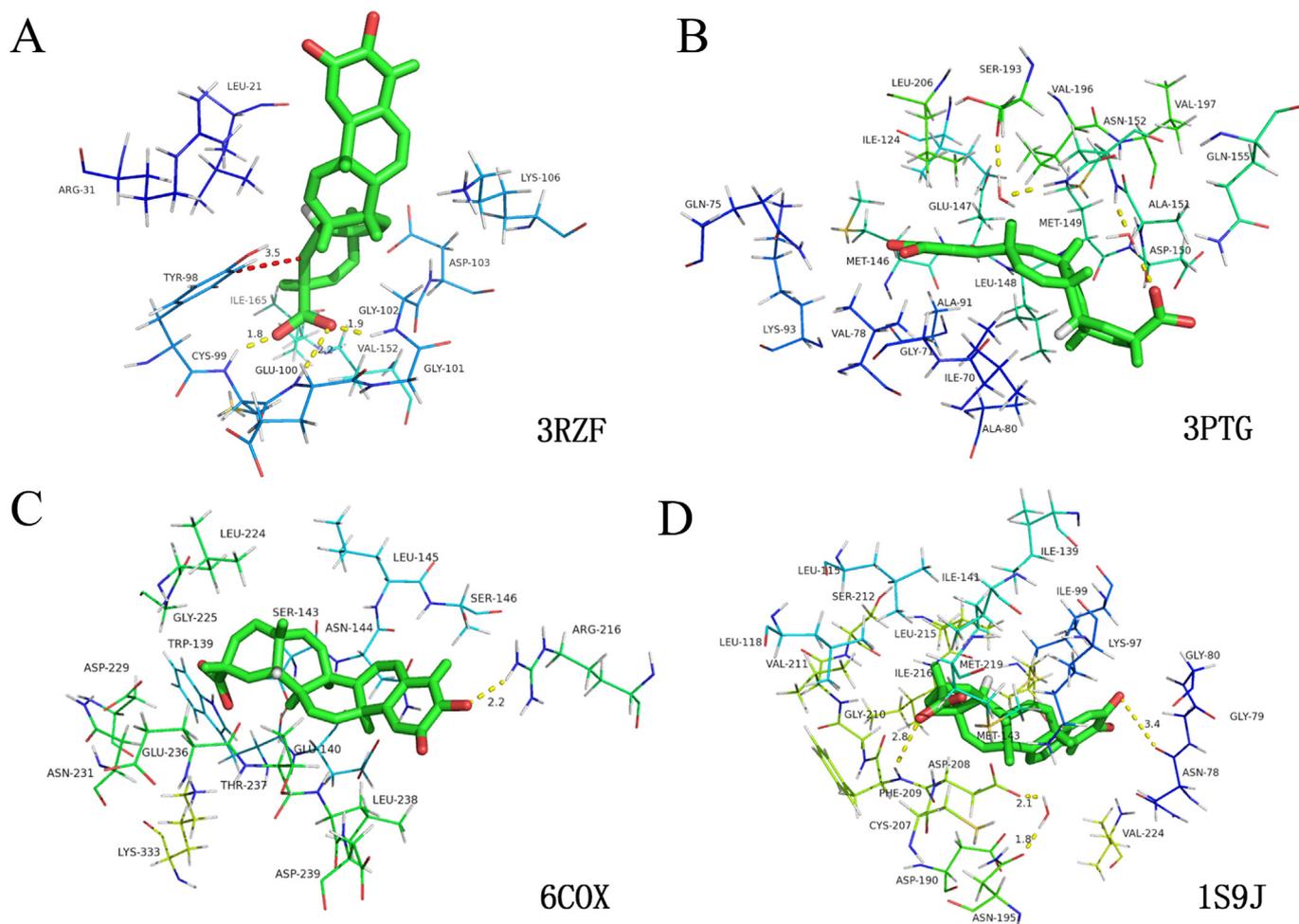


Fig. 6. Molecular models of celastrol binding to the predicted targets (A) 3RZF, (B) 3PTG, (C) 6COX and (D) 1S9J. The yellow dashed lines show H-bonds, and the red dashed lines show π - π interactions, with interaction distances indicated above the lines. The sticks represent the celastrol molecule, and the lines represent residues in the binding site. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

networks that we identified as associated with rheumatoid arthritis and affected by celastrol involve primarily cancer, connective tissue disorders, organismal injury and abnormalities, tissue development, cell death and survival. Consistent with our findings, celastrol has been shown to inhibit the proliferation of various cancer cell lines, including C6 glioma, human monocytic leukemia, melanoma, pancreatic cancer, RPMI 8266 myeloma, and K-562 human chronic myelogenous leukemia [42–46].

Though this network pharmacology strategy may be a quick method to predict the targets by drugs on complex diseases, the targets sometimes can be obtained too much. It is necessary to verify the targets by cell experiments.

In particular, our results predict that celastrol exerts therapeutic effects against rheumatoid arthritis at least in part by modulating the function of MMP-9, COX-2, c-Myc, TGF- β , c-JUN, JAK-1, JAK-3, IKK- β , SYK, MMP-3, JNK and MEK1. Our study may inspire and guide further work to establish the molecular targets of celastrol in rheumatoid arthritis and to apply the network pharmacology approach to drug discovery against other inflammatory and autoimmune diseases.

Declaration of Competing Interest

The authors declare no conflicts of interest.

Acknowledgments

Supported by National Natural Science Foundation of China (grant No. U1804179), Henan Science and Technology Innovation Team, Investigation on Plant Resources in Dabie Mountains and the study and utilization of active components of special plants (grant no. 2017083), Nanhu Scholars Program for Young Scholars of Xinyang Normal University (grant no. 2018001).

References

- [1] J.S. Smolen, D. Aletaha, I.B. McInnes, Rheumatoid arthritis, *Lancet* 388 (10055) (2016) 2023–2038.
- [2] M. Abbasi, M.J. Mousavi, S. Jamalzebi, R. Alimohammadi, M.H. Bezan, H. Mohammadi, S. Aslani, Strategies toward rheumatoid arthritis therapy; the old and the new, *J. Cell. Physiol.* 234 (7) (2019) 10018–10031.
- [3] P. Wehr, H. Purvis, S.C. Law, R. Thomas, Dendritic cells, T cells and their interaction in rheumatoid arthritis, *Clin. Exp. Immunol.* 196 (1) (2019) 12–27.
- [4] G. Boleto, L. Kanagaratnam, M. Drame, J.H. Salmon, Safety of combination therapy with two bDMARDs in patients with rheumatoid arthritis: a systematic review and meta-analysis, *Semin. Arthritis Rheum.* 58 (18) (2018) 30653–30659.
- [5] S.J. Chen, G.J. Lin, J.W. Chen, K.C. Wang, C.H. Tien, C.F. Hu, C.N. Chang, W.F. Hsu, H.C. Fan, H.K. Sytwu, Immunopathogenic mechanisms and novel immune-modulated therapies in rheumatoid arthritis, *Int. J. Mol. Sci.* 20 (6) (2019).
- [6] J. Ding, G. Orozco, Identification of rheumatoid arthritis causal genes using functional genomics, *Scand. J. Immunol.* 89 (5) (2019) 1–12 e12753.
- [7] J. Karami, S. Aslani, A. Jamshidi, M. Garshasbi, M. Mahmoudi, Genetic implications in the pathogenesis of rheumatoid arthritis; an updated review, *Gene* 702 (2019) 8–16.
- [8] S. Tasneem, B. Liu, B. Li, M.I. Choudhary, W. Wang, Molecular pharmacology of inflammation: medicinal plants as anti-inflammatory agents, *Pharmacol. Res.* 139

- (2019) 126–140.
- [9] D. Luo, Z. Zuo, H. Zhao, Y. Tan, C. Xiao, Immunoregulatory effects of *Tripterygium wilfordii* Hook F and its extracts in clinical practice, *Front Med* (2019), <https://doi.org/10.1007/s11684-018-0649-5>.
- [10] S.W. Ng, Y. Chan, D.K. Chellappan, T. Madheswaran, F. Zeeshan, Y.L. Chan, T. Collet, G. Gupta, B.G. Oliver, P. Wark, et al., Molecular modulators of celastrol as the keystones for its diverse pharmacological activities, *Biomed. Pharmacother.* 109 (2019) 1785–1792.
- [11] T. Bufu, X. Di, Z. Yilin, L. Gege, C. Xi, W. Ling, Celastrol inhibits colorectal cancer cell proliferation and migration through suppression of MMP3 and MMP7 by the PI3K/AKT signaling pathway, *Anti-Cancer Drugs* 29 (6) (2018) 530–538.
- [12] L. Guo, S. Luo, Z. Du, M. Zhou, P. Li, Y. Fu, X. Sun, Y. Huang, Z. Zhang, Targeted delivery of celastrol to mesangial cells is effective against mesangioproliferative glomerulonephritis, *Nat. Commun.* 8 (1) (2017) 878.
- [13] Z. Jiang, Q. Cao, G. Dai, J. Wang, C. Liu, L. Lv, J. Pan, Celastrol inhibits colorectal cancer through TGF-beta1/Smad signaling, *Oncotargets Ther* 12 (2019) 509–518.
- [14] X. Li, H. Wang, J. Ding, S. Nie, L. Wang, L. Zhang, S. Ren, Celastrol strongly inhibits proliferation, migration and cancer stem cell properties through suppression of Pin1 in ovarian cancer cells, *Eur. J. Pharmacol.* 842 (2019) 146–156.
- [15] X. Li, G. Zhu, X. Yao, N. Wang, R. Hu, Q. Kong, D. Zhou, L. Long, J. Cai, W. Zhou, Celastrol induces ubiquitin-dependent degradation of mTOR in breast cancer cells, *Oncotargets Ther* 11 (2018) 8977–8985.
- [16] Y. Qi, R. Wang, L. Zhao, L. Lv, F. Zhou, T. Zhang, F. Lu, H. Yan, G. Duan, Celastrol suppresses tryptophan catabolism in human colon cancer cells as revealed by metabolic profiling and targeted metabolite analysis, *Biol. Pharm. Bull.* 41 (8) (2018) 1243–1250.
- [17] H. Xiao-Pei, C. Ji-Kuai, W. Xue, Y.F. Dong, L. Yan, Z. Xiao-Fang, P. Ya-Min, C. Wen-Jun, Z. Jiang-Bo, Systematic identification of celastrol-binding proteins reveals that Shoc2 is inhibited by celastrol, *Biosci. Rep.* 38 (6) (2018).
- [18] L.N. Xu, N. Zhao, J.Y. Chen, P.P. Ye, X.W. Nan, H.H. Zhou, Q.W. Jiang, Y. Yang, J.R. Huang, M.L. Yuan, et al., Celastrol inhibits the growth of ovarian cancer cells in vitro and in vivo, *Front. Oncol.* 9 (2019) 2.
- [19] C. Zhang, R. Wang, Z. Liu, E. Bunker, S. Lee, M. Giuntini, D. Chapnick, X. Liu, The plant triterpenoid celastrol blocks PINK1-dependent mitophagy by disrupting PINK1's association with the mitochondrial protein TOM20, *J. Biol. Chem.* 294 (18) (2019) 7472–7487.
- [20] D. Zhang, Z. Chen, C. Hu, S. Yan, Z. Li, B. Lian, Y. Xu, R. Ding, Z. Zeng, X.K. Zhang, et al., Celastrol binds to its target protein via specific noncovalent interactions and reversible covalent bonds, *Chem Commun (Camb)* 54 (91) (2018) 12871–12874.
- [21] Q. Zhao, F. Liu, Y. Cheng, X.R. Xiao, D.D. Hu, Y.M. Tang, W.M. Bao, J.H. Yang, T. Jiang, J.P. Hu, et al., Celastrol protects from cholestatic liver injury through modulation of SIRT1-FXR signaling, *Mol. Cell. Proteomics* 18 (3) (2019) 520–533.
- [22] Y.L. Zhong, G.J. Xu, S. Huang, L. Zhao, Y. Zeng, X.F. Xiao, J.L. An, J. Liu, T. Yang, Celastrol induce apoptosis of human multiple myeloma cells involving inhibition of proteasome activity, *Eur. J. Pharmacol.* 853 (2019) 184–192.
- [23] S. Li, T.P. Fan, W. Jia, A. Lu, W. Zhang, Network pharmacology in traditional Chinese medicine, *Evid. Based Complement. Alternat. Med.* 2014 (2014) 138460.
- [24] X.M. Wu, C.F. Wu, Network pharmacology: a new approach to unveiling traditional Chinese medicine, *Chin. J. Nat. Med.* 13 (1) (2015) 1–2.
- [25] B. Boezio, K. Audouze, P. Ducrot, O. Taboureau, Network-based approaches in pharmacology, *Mol Inform* 36 (10) (2017).
- [26] W. Chen, W. Da, C. Li, H. Fan, R. Liang, J. Yuan, X. Huang, R. Yang, J. Zhang, J. Zhu, Network pharmacology-based identification of the protective mechanisms of taraxasterol in experimental colitis, *Int. Immunopharmacol.* 71 (2019) 259–266.
- [27] F. Cheng, I.A. Kovacs, A.L. Barabasi, Network-based prediction of drug combinations, *Nat. Commun.* 10 (1) (2019) 1197.
- [28] Y. Dong, P. Qiu, R. Zhu, L. Zhao, P. Zhang, Y. Wang, C. Li, K. Chai, D. Shou, H. Zhao, A combined phytochemistry and network pharmacology approach to reveal the potential antitumor effective substances and mechanism of *Phellinus igniarius*, *Front. Pharmacol.* 10 (2019) 266.
- [29] X. Feng, H. Shi, X. Chao, F. Zhao, L. Song, M. Wei, H. Zhang, Deciphering the pharmacological mechanism of the herb *Radix Ophiopogonis* in the treatment of nasopharyngeal carcinoma by integrating iTRAQ-coupled 2-D LC-MS/MS analysis and network investigation, *Front. Pharmacol.* 10 (2019) 253.
- [30] B. Fleisher, A.N. Brown, S. Ait-Oudhia, Application of pharmacometrics and quantitative systems pharmacology to cancer therapy: the example of luminal a breast cancer, *Pharmacol. Res.* 124 (2017) 20–33.
- [31] A. Huang, G. Fang, Y. Pang, Z. Pang, A network pharmacology approach to explore mechanism of action of Longzuan Tongbi formula on rheumatoid arthritis, *Evid. Based Complement. Alternat. Med.* 2019 (2019) 5191362.
- [32] H. Lim, L. Xie, Omics data integration and analysis for systems pharmacology, *Methods Mol. Biol.* 2019 (1939) 199–214.
- [33] Y.J. Lin, W.M. Liang, C.J. Chen, H. Tsang, J.S. Chiou, X. Liu, C.F. Cheng, T.H. Lin, C.C. Liao, S.M. Huang, et al., Network analysis and mechanisms of action of Chinese herb-related natural compounds in lung cancer cells, *Phytomedicine* 58 (2019) 152893.
- [34] J.F. Liu, A.N. Hu, J.F. Zan, P. Wang, Q.Y. You, A.H. Tan, Network pharmacology deciphering mechanisms of volatiles of Wendan granule for the treatment of Alzheimer's disease, *Evid. Based Complement. Alternat. Med.* 2019 (2019) 7826769.
- [35] T.T. Luo, Y. Lu, S.K. Yan, X. Xiao, X.L. Rong, J. Guo, Network pharmacology in research of Chinese medicine formula: methodology, application and prospective, *Chin J Integr Med* (2019), <https://doi.org/10.1007/s11655-019-3064-0>.
- [36] C. Ma, T. Xu, X. Sun, S. Zhang, S. Liu, S. Fan, C. Lei, F. Tang, C. Zhai, C. Li, et al., Network pharmacology and bioinformatics approach reveals the therapeutic mechanism of action of baicalein in hepatocellular carcinoma, *Evid. Based Complement. Alternat. Med.* 2019 (2019) 7518374.
- [37] E. Shawk, Prediction of potential cancer-related molecular targets of North African plants constituents using network pharmacology-based analysis, *J. Ethnopharmacol.* 28 (238) (2019) 111826–111834 111826.
- [38] M. Wei, Y. Liu, Z. Pi, S. Li, M. Hu, Y. He, K. Yue, T. Liu, Z. Liu, F. Song, et al., Systematically characterize the anti-Alzheimer's disease mechanism of lignans from *S. chinensis* based on in-vivo ingredient analysis and target-network pharmacology strategy by UHPLC(-)Q-TOF-MS, *Molecules* 24 (7) (2019).
- [39] Y. Xiong, Y. Yang, W. Xiong, Y. Yao, H. Wu, M. Zhang, Network pharmacology-based research on the active component and mechanism of the antihepatoma effect of *Rubia cordifolia* L., *J. Cell. Biochem.* 120 (8) (2019) 12461–12472, <https://doi.org/10.1515/hsz-2018-0469>.
- [40] R. Zhang, X. Zhu, H. Bai, K. Ning, Network pharmacology databases for traditional Chinese medicine: review and assessment, *Front. Pharmacol.* 10 (2019) 123.
- [41] J. Xi, Q. Li, X. Luo, Y. Wang, J. Li, L. Guo, G. Wu, Celastrol inhibits glucocorticoid-induced osteoporosis in rat via the PI3K/AKT and Wnt signaling pathways, *Mol. Med. Rep.* 18 (5) (2018) 4753–4759.
- [42] S.R. Chen, Y. Dai, J. Zhao, L. Lin, Y. Wang, Y. Wang, A mechanistic overview of triptolide and celastrol, natural products from *Tripterygium wilfordii* Hook F, *Front. Pharmacol.* 9 (2018) 104.
- [43] Z. Wang, Z. Zhai, X. Du, Celastrol inhibits migration and invasion through blocking the NF-kappaB pathway in ovarian cancer cells, *Exp Ther Med* 14 (1) (2017) 819–824.
- [44] D. Kashyap, A. Sharma, H.S. Tuli, K. Sak, T. Mukherjee, A. Bishayee, Molecular targets of celastrol in cancer: recent trends and advancements, *Crit Rev Oncol Hematol* 128 (2018) 70–81.
- [45] Y. Gao, S. Zhou, L. Pang, J. Yang, H.J. Li, X. Huo, S.Y. Qian, Celastrol suppresses nitric oxide synthases and the angiogenesis pathway in colorectal cancer, *Free Radic. Res.* (2019) 1–11.
- [46] A. Zuo, P. Zhao, Y. Zheng, H. Hua, X. Wang, Tripterine inhibits proliferation, migration and invasion of breast cancer MDA-MB-231 cells by up-regulating microRNA-15a, *Biol. Chem.* (2018).