



Review

Treatment of chronic hepatitis due to hepatitis B and hepatitis delta virus coinfection

Giuseppina Brancaccio^a, Giovanni B. Gaeta^{b,*}^a Infectious Diseases, Department of Molecular Medicine, University of Padua, Padua, Italy^b Infectious Diseases, Department of Mental and Physical Health, Campania University, Naples, Italy

ARTICLE INFO

Article history:

Received 5 March 2019

Accepted 15 September 2019

Editor: Prof. Philippe Colson

Keywords:

Hepatitis delta virus

Peg-IFN

Prenylation inhibitor

Entry inhibitor

HBsAg release blocker

IFN-lambda

ABSTRACT

An estimated 20–40 million individuals worldwide are infected with hepatitis delta virus (HDV), mostly with rapidly evolving liver disease. Therapy of chronic HDV infection remains an unmet need. To date, only interferon (IFN)-based therapy is recommended for HDV infection and response rates are unsatisfactory; in addition, many patients are intolerant to or ineligible for IFN treatment. In recent years, innovative approaches have been in development, including the following: targeting virus entry into hepatocytes; inhibition of the host enzyme farnesyltransferase by prenylation inhibitors, leading to inhibition of complete virion formation and release; blockade of hepatitis B surface antigen (HBsAg) secretion, inhibiting virus release; and IFN-lambda, which causes fewer adverse effects than IFN-alfa. Clinical trials are ongoing with encouraging preliminary results.

© 2019 Elsevier B.V. and International Society of Chemotherapy. All rights reserved.

1. Introduction

Hepatitis delta virus (HDV) has an estimated global burden of 20–40 million chronically infected individuals; however, estimates are largely inaccurate due to the lack of systematic screening for HDV in hepatitis B surface antigen (HBsAg)-positive individuals [1,2]. Clusters of infection are still present in Eastern Europe, sub-Saharan Africa, South America and Asia [3,4]. In Western Europe, following a rapid decrease in prevalence over the period 1980–2000 [5], a re-emergence of chronic HDV infection has been noted, mostly due to immigration from endemic areas [6–9]. Eight genotypes of HDV are known [10]; genotype 1 is ubiquitous and largely predominates in Europe, whilst the other genotypes have a typical geographical distribution. Superinfection by HDV in a chronic carrier of hepatitis B virus (HBV) generates a chronic infection in >80% of cases, which may evolve rapidly towards cirrhosis and hepatocellular carcinoma; simultaneous infection by the two viruses causes severe acute hepatitis that progresses towards chronic infection in <5% of cases [11]. HBV replication is suppressed in most chronically coinfecting patients; in approximately 25% HBV may dominate or both viruses may fluctuate over time [12,13].

HDV is a unique human pathogen. Its small RNA (1.7 kb) encodes for only one protein, the hepatitis delta antigen (HDAG),

in two forms, namely small (S-HDAG) and large (L-HDAG), whereas it lacks functional proteins that usually drive viral replication [14,15]. As such, HDV utilises human polymerase II and to some extent polymerase I to replicate and requires HBsAg to generate the complete virion, which uses the HBV cell receptor to enter hepatocytes. Further characteristics of HDV are: (i) the presence of ribozyme, a non-coding RNA typically present in plant virusoids and some viroids, that cleaves multimeric synthesised RNA into monomeric forms; and (ii) the use of host adenosine deaminase to convert adenosine to inosine on the antigenomic RNA; this causes a change of the UAG amber codon in the S-HDAG open reading frame (ORF) to a UGG tryptophan codon for the L-HDAG ORF [16]. Finally, prenylation at the CXXX box motif (where C represents cysteine and X any other amino acid) of L-HDAG by human farnesyltransferase is critical to promote its binding to HBsAg to generate the complete HDV virion (Fig. 1).

Due to its simple structure, HDV lacks the usual targets for most antiviral drugs, e.g. viral polymerase or other functional proteins. To date, the only drug registered for HDV infection is interferon (IFN) or pegylated interferon (peg-IFN). Innovative therapeutic approaches target non-replicative steps of the virus cycle, such as entry into hepatocytes or the prenylation process. In addition, some anti-HBV drugs that block HBsAg release have the potential to prevent the production of HDV virions. This brief review focuses on the results of IFN-based therapies and examines current data on new therapies.

* Corresponding author.

E-mail address: giovannibattista.gaeta@unicampania.it (G.B. Gaeta).

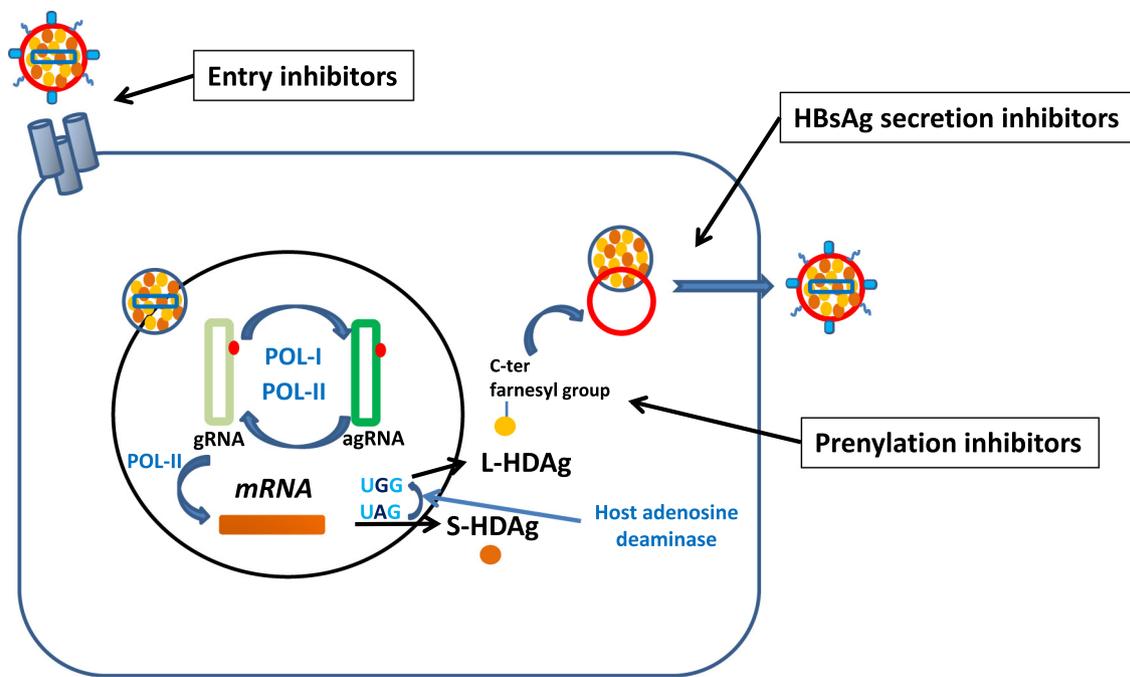


Fig. 1. Main steps in the hepatitis delta virus (HDV) replication cycle. Black arrows indicate the targets of novel therapies. agRNA, antigenomic RNA; gRNA, genomic RNA; HBsAg, hepatitis B surface antigen; L-HDAg, large hepatitis delta antigen; POL, polymerase; S-HDAg, small hepatitis delta antigen.

2. Endpoints of treatment

Antiviral treatment in chronic HDV infection, as for other chronic viral hepatitis, is aimed at preventing the progression of liver disease and death. In clinical practice and in trials evaluating antiviral drugs, surrogate efficacy endpoints are measured. The ideal endpoint is sustained undetectability of HDV-RNA in plasma, which can be achieved following HBsAg clearance; a transient presence of HDV-RNA in plasma has been reported in a few cases following HBsAg clearance [17,18]. Unfortunately, clearance of HBsAg remains a rare event with current treatments against HBV infection. In practice, a decline of HDV-RNA of ≥ 2 log with alanine aminotransferase normalisation at the end of therapy is considered a valuable efficacy endpoint [19]. Maintenance of response or reaching HDV-RNA undetectability at Week 24 or Week 48 after the end of therapy are additional endpoints; however, late relapses following IFN-based treatments may occur. An extensive evaluation of the endpoints of treatment in chronic HDV is available from Yurdaydin et al. [19].

In evaluating early literature studies, it must be noted that there was large variability in the sensitivity and reproducibility of HDV-RNA detection methods [20]; however, a reference standard is now available [WHO N.7657/12, Paul-Ehrlich-Institut].

3. Interferon-alfa (IFN- α)-based therapies

Recombinant IFN- α was introduced for the therapy of chronic HDV infection in the mid-1980s. It was soon clear that the standard dose of 3 million units (MU) thrice weekly had poor efficacy and that 9 MU thrice weekly or 5 MU daily were required to reach some response in terms of viral suppression. Farci et al. showed that at higher doses, IFN- α resulted in long-term improvement of liver fibrosis in some patients [21]. Another cohort study demonstrated a long-term benefit in the rate of liver-related complications and in the survival in 35 patients who responded to IFN- α given for a median of 24 months compared with those who remained viraemic [22]. In general, maintenance of viral suppression favourably influenced the clinical course of the disease, as

measured by the occurrence of liver-related events, death or liver transplant. The presence of cirrhosis at baseline predicted an adverse outcome as did persistence of HDV replication or lack of IFN-based therapy [23–26]. Nevertheless, tolerability to IFN- α is poor and patients with advanced liver disease or major contraindications to IFN- α are not eligible for treatment. According to a Cochrane review, an overall a virological response was achieved in 12% of treated patients [27].

The advent of peg-IFN α simplified administration schedules but did not significantly improve treatment outcomes (Table 1) [28–40]. Two trials with peg-IFN α 2b reported a sustained virological response (SVR), defined as HDV-RNA undetectable at Month 6 post-therapy, of 43% and 25%, respectively [28,29]; co-administration of ribavirin did not improve the response rate. Overall response rates obtained in studies using peg-IFN α for ≥ 12 months were in the range 17–47%, with the exception of one study performed in South America that reported a 95% response in patients infected by HDV genotype 3 [38]. Only three studies were randomised [29,33,36] and showed response rates from 25–31% in the treatment arms receiving peg-IFN alone for 48–96 weeks. Combination with adefovir or tenofovir was of no benefit even when treatment was prolonged to 96 weeks [33,36]. Interestingly, a 5-year post-therapy follow-up study showed that late HDV-RNA relapse, i.e. later than a SVR assessed 6 months after the end of therapy using sensitive HDV-RNA detection methods, occurred in approximately 50% of patients, suggesting that the paradigm of SVR at Month 6 post-therapy may be inappropriate for HDV [41]. These unsatisfactory results stimulated some studies aimed at identifying on therapy those patients with a higher chance of response. Niro et al. examined the kinetics of HBsAg and HDV-RNA decline and found that a decrease of both markers within Month 6 of treatment predicted the definitive clearance of HDV-RNA [42]. Keskin et al. found that patients who achieved undetectable HDV-RNA within 6 months of therapy had a high probability of maintaining virological response at Week 24 after the end of therapy [43]. Lutterkort et al. found that a pre-therapy virological pattern of dominance of HBV replication (found in 7% of 109 patients) or of non-dominance (in 17%; i.e. the viral load of one virus did not exceed that of the

Table 1
Summary of studies using pegylated interferon (peg-IFN) for chronic hepatitis delta virus (HDV) infection.

References	Type/dose	No. of patients	Duration of therapy	Duration of follow-up	Overall virological response at EOT (%)	SVR (%)
[28]	Peg-IFN α 2b 1.5 μ g/kg	14	12 months	16 months	57	43
[29]	Peg-IFN α 2b 1.5 μ g/kg	16	18 months	6 months	19	25
	Peg-IFN α 2b 1.5 μ g/kg + RBV	22	12 months	6 months	9	18
[30]	Peg-IFN α 2b 1.5 μ g/kg	12	12 months	12 months	–	17
[31]	Peg-IFN α 2b 1.5 μ g/kg	49	13 months	26 months	33	25
[32]	Peg-IFN α 2b 1.5 μ g/kg	11	24 months	6 months	56	–
		7	12 months	6 months	57	
[33]	Peg-IFN α 2a 180 μ g	29	12 months	6 months	24	31
	Peg-IFN α 2a 180 μ g + adefovir 10 mg/day	31	12 months	6 months	23	24
	Adefovir 10 mg/day	30	12 months	6 months	0	0
[34]	Peg-IFN α 2b	277 enrolled (238 evaluated)	48 weeks	24 weeks	29.8	29.4
[35]	Peg-IFN α 2a 180 μ g or peg-IFN α 2b 1.5 μ g/kg	32	24 months	6 months	50	47
[36]	Peg-IFN α 2a + tenofovir	59	96 weeks	24 weeks	48	29
	Peg-IFN α 2a + placebo	61	96 weeks	24 weeks	33	21
[37]	Peg-IFN α 2a 90–270 μ g/week	13	6–240 weeks (median, 140 weeks)	–	–	39 (3 lost HBsAg)
[38] ^a	Peg-IFN α 2a 180 μ g + entecavir 0.5 mg/day	22	48 weeks	48 weeks	95	95
[39]	Peg-IFN α 2a	41	12 months		39	37
	Peg-IFN α 2b	15	12 months		13	13
[40]	IFN or peg-IFN	99	6–126 months (median, 24 months)	24–225 months (median, 55 months)	–	35.3

EOT, end of treatment; SVR, sustained virological response; RBV, ribavirin; HBsAg, hepatitis B surface antigen.

^a South American study enrolling patients with HDV genotype 3.

other virus) was associated with a favourable cytokine/chemokine network and a higher response rate to peg-IFN α compared with patients with HDV dominance [44]. It remains unclear whether therapy should be prolonged on the basis of individual kinetics of HBsAg decrease in patients who achieve negative HDV-RNA. The recently introduced HBV marker, hepatitis B core-related antigen (HBcrAg), might have a role in defining the progression of liver disease in combination with other virological markers [45] but has not been assessed as a predictor of response to therapy.

4. Nucleos(t)ide analogues

Lamivudine, administered alone or in combination with IFN, was ineffective in inducing a decrease in HDV-RNA [46–48]. The same results were obtained with adefovir [33]. Current-generation analogues (entecavir, tenofovir) given alone for 24–96 weeks were also ineffective. In a well-designed randomised trial, tenofovir plus peg-IFN α for 96 weeks resulted in a better virological response at the end of treatment compared with peg-IFN α alone; however, the advantage was lost during the post-therapy follow-up [36,49,50]. In patients with advanced liver disease replicating both HBV and HDV and treated with entecavir or tenofovir for a median of 50 months, liver-related event rates were 3–10-fold higher than in pair-matched HBV mono-infected patients receiving the same therapy [51]. In all, therapeutic attempts to decrease the background of HBV infection failed to decrease HDV production since analogues did not affect HBsAg production.

5. Entry inhibitors

HBV enters hepatocytes through binding of the pre-S1 domain (S-HBsAg) of HBsAg with its receptor on the hepatocyte membrane, the sodium-taurocholate cotransporter peptide (NTCP), which drives the internalisation process [52]. HDV, being coated with HBsAg, utilises the same receptor (Fig. 1). Myrcludex B (MyrB; bulevirtide) is a synthetic *N*-acylated peptide derived from the pre-S1 domain that inhibits the HBV receptor on the

hepatocyte surface and prevents the infection of healthy cells and viral spread within the liver [53]. Other classes of entry inhibitors, not targeting NTCP, have been tested in vitro. Bogolomov et al. provided the proof of concept that MyrB could decrease HDV viraemia [54]. They randomised 24 patients to three groups that received, respectively: MyrB 2 mg daily as subcutaneous (s.c.) injection for 24 weeks, followed by weekly peg-IFN α 2a alone for 48 weeks (Myr cohort); MyrB 2 mg daily plus peg-IFN α 2a for 24 weeks, followed by peg-IFN α 2a alone for 24 weeks (Myr-IFN cohort); and peg-IFN α 2a alone for 48 weeks (IFN cohort). At an interim analysis at Week 24 of treatment, they demonstrated a decline of ≥ 1 log in HDV-RNA in 6/7 evaluable patients in the Myr group, with disappearance in 2; in 7/7 patients in the Myr-IFN group, with disappearance in 5; and in 6/7 patients treated with peg-IFN α 2a alone. In a phase 2 study, 120 patients, all receiving tenofovir for ≥ 12 weeks, were assigned to add 2, 5 or 10 mg of MyrB subcutaneously for 24 weeks or no MyrB [55]. An HDV-RNA decline of ≥ 2 log was achieved at Week 24 in 46.4% of the 2 mg group, 46.8% of the 5 mg group and 76.6% of the 10 mg group [55]. This was an interesting proof of concept that IFN-free therapy is feasible in chronic HDV infection; surprisingly, no patient achieved the primary endpoint of an HBsAg decline of ≥ 0.5 log IU/mL. MyrB was generally safe; it caused an elevation in the concentration of serum bile acids in all dosed patients that remained clinically silent and promptly returned to normal after the end of therapy. Phase 3 studies are in progress exploring longer treatment durations or various combinations with peg-IFN. Recently, the safety and efficacy of MyrB monotherapy at a daily dose of 10 mg for 48 weeks was reported in three patients with cirrhosis treated on a compassionate basis [56].

6. Prenylation inhibitors

These drugs target the host enzyme farnesyltransferase, which catalyses the farnesylation process at the carboxy edge of L-HDAG, thus making it able to bind HBsAg. Lonafarnib (LNF) was first introduced as an anti-leukaemia drug. Its use in chronic HDV infection

was first assessed in vitro and in vivo in a mouse model [57,58]. In a phase 2A, randomised, proof-of-concept study, Koh et al. assigned 14 HDV patients to receive oral LNF at 100 mg (group 1) or 200 mg (group 2) twice a day or placebo for 28 days, followed by a 6-month follow-up; the primary outcome was a decrease in serum HDV-RNA [59]. By the end of therapy, the mean HDV-RNA decline in serum from baseline was 0.73 log IU/mL [95% confidence interval (CI) 0.17–1.31] in group 1 and 1.54 log IU/mL (95% CI 1.21–1.93) in group 2 versus 0.13 log IU/mL (95% CI –0.14 to 0.37) in the placebo group. At post-therapy follow-up, HDV-RNA returned to baseline in all patients. Gastrointestinal symptoms, including nausea and diarrhoea, were common in patients receiving LNF. Interestingly, the dose-dependent effect of LNF was further demonstrated by the linear correlation between serum concentration of the drug and HDV-RNA decline. To increase concentrations of LNF and to overcome its toxicity, the combination of ritonavir (RTV), a CYP3A4 inhibitor, with 100 mg LNF was proposed. In a proof-of-concept study, Yurdaydin et al. showed that the RTV/LNF combination, administered twice daily, had better antiviral activity than LNF 300 mg twice daily with less adverse events [60]. In addition, in the same study a synergistic effect of the combination of LNF + peg-IFN α was shown, thus supporting further development of the drug. A multicentre phase 3 trial is now ongoing.

7. Blockade of hepatitis B surface antigen release/production

Nucleic acid polymers (NAPs) inhibit the release of HBV subviral particles; since HDV virions use the same secretion mechanism, a therapeutic effect on HDV infection was hypothesised. In vitro, these compounds blocked HDV infection in cultures of differentiated human hepatoma cells [61]. In a chronic duck HBV infection model in Pekin ducks, synergy with nucleos(t)ide analogues was noted [62].

Bazinet et al. treated 12 non-cirrhotic patients with chronic HDV infection (>6 months) with 500 mg of intravenous REP 2139 weekly for 15 weeks, followed by combined therapy with 250 mg of intravenous REP 2139 and 180 μ g of s.c. peg-IFN α 2a weekly for 15 weeks, and then monotherapy with 180 μ g of peg-IFN weekly for an additional 33 weeks [63]. All patients were followed-up for 1 year after the end of therapy. Six patients achieved an HBsAg level <0.05 IU/mL; the suppression was maintained at the end of follow-up and all patients developed significant hepatitis B surface antibody (anti-HBs) titres. Eleven patients became HDV-RNA negative during treatment, with seven of them remaining negative by the end of follow-up. Hair loss, dysphagia and dysgeusia have been reported with NAPs. Elevation of serum aminotransferases was recorded in all patients with HBsAg suppression at <1 IU/mL. This latter event was also observed in patients with HBV monoinfection receiving NAP with peg-IFN and tenofovir and may limit the use of this treatment in patients with cirrhosis. The mechanism of inhibition of HDV replication by NAPs is not fully clarified, although accumulation of HDV L-protein inside hepatocytes could play a role.

8. Interferon-lambda (IFN- λ)

IFN- λ could be an alternative to IFN- α since it binds the type 1 IFN receptor that is prevalently expressed on hepatocytes; as a consequence, less systemic adverse effects are expected. In HBV/HDV-infected humanised mice, both IFN- α and IFN- λ reduced intrahepatic HDV infection markers [64]. In a phase 2 study, 33 patients with chronic HDV infection were randomised to receive peg-IFN λ at a dose of either 120 μ g or 180 μ g by weekly s.c. injection for 48 weeks; all patients also received daily tenofovir or entecavir [65]. A 2 log decrease in HDV-RNA was achieved in 53% and 90% patients, respectively. Typical adverse effects seen with IFN- α were

fewer, but 10% of patients experienced hyperbilirubinemia and/or alanine aminotransferase increase; both effects were reversed by dose reduction. A phase 2a open-label study to evaluate the safety and antiviral effects of triple therapy with LNF/RTV and IFN- λ for a period of 24 weeks is ongoing.

9. Conclusions

For decades, treatment for chronic HDV infection has been based on IFN- α , leading to unsatisfactory results. New therapeutic approaches provide some hope for the near future, since efficacy results surpassed those achieved with IFN and tolerability looked better than for IFN. Controlled trials are ongoing and will quantify the advantages. However, most studies still combine the new agents with peg-IFN, which involves the limitations of this treatment. IFN- λ could be an alternative since it causes fewer adverse events than IFN- α . Theoretically, IFN-free combinations of two drugs with or without a nucleos(t)ide analogue could be an interesting option, which requires further investigation

Funding: None.

Competing Interest: None declared.

Ethical approval: Not required.

References

- [1] Hughes SA, Wedemeyer H, Harrison PM. Hepatitis delta virus. *Lancet* 2011;378:73–85.
- [2] Chen HY, Shen DT, Ji DZ, Han PC, Zhang WM, Ma JF, et al. Prevalence and burden of hepatitis D virus infection in the global population: a systematic review and meta-analysis. *Gut* 2018 Sep 18 [Epub ahead of print]. doi:10.1136/gutjnl-2018-316601.
- [3] Rizzetto M. Hepatitis delta: thirty years after. *J Hepatol* 2009;50:1043–50.
- [4] Wrانke A, Pinheiro Borzacov LM, Parana R, Lobato C, Hamid S, Ceausu E. Hepatitis Delta International Network. Clinical and virological heterogeneity of hepatitis delta in different regions world-wide: the Hepatitis Delta International Network (HDIN). *Liver Int* 2018;38:842–50.
- [5] Gaeta GB, Stroffolini T, Chiaramonte M, Ascione T, Stornaiuolo G, Lobello S, et al. Chronic hepatitis D: a vanishing disease? An Italian multicenter study. *Hepatology* 2000;32:824–7.
- [6] Gaeta GB, Stroffolini T, Smedile A, Niro G, Mele A. Hepatitis delta in Europe: vanishing or refreshing? *Hepatology* 2007;46:1312–13.
- [7] Cross TJS, Rizzi P, Horner M, Jolly A, Hussain MJ, Smith HM, et al. The increasing prevalence of hepatitis delta virus (HDV) infection in south London. *J Med Virol* 2008;80:277–82.
- [8] Wedemeyer H, Heidrich B, Mann MP. Hepatitis D virus infection—not a vanishing disease in Europe!. *Hepatology* 2007;45:1331–2.
- [9] Brancaccio G, Nardi A, Madonia S, Fasano M, Verucchi G, Massari M, et al. The present profile of chronic hepatitis B virus infection highlights future challenges: an analysis of the multicenter Italian MASTER-B cohort. *Dig Liver Dis* 2018;51:438–42.
- [10] Alvarado-Mora MV, Locarnini S, Rizzetto M, Pinho JR. An update on HDV: virology, pathogenesis and treatment. *Antivir Ther* 2013;18:541–8.
- [11] Rizzetto M, Verme G. Delta hepatitis—present status. *J Hepatol* 1985;1:187–93.
- [12] Raimondo G, Brunetto MR, Pontisso P, Smedile A, Maina AM, Saitta C. Associazione Italiana Studio Fegato Cooperative Group. Longitudinal evaluation reveals a complex spectrum of virological profiles in hepatitis B virus/hepatitis C virus-coinfected patients. *Hepatology* 2006;43:100–7.
- [13] Schaper M, Rodriguez-Frias F, Jardi R, Taberner D, Homs M, Ruiz G, et al. Quantitative longitudinal evaluations of hepatitis delta virus RNA and hepatitis B virus DNA shows a dynamic, complex replicative profile in chronic hepatitis B and D. *J Hepatol* 2010;52:658–64.
- [14] Lai MM. RNA replication without RNA-dependent RNA polymerase: surprises from hepatitis delta virus. *J Virol* 2005;79:7951–8.
- [15] Taylor JM. Virology of hepatitis D virus. *Semin Liver Dis* 2012;32:195–200.
- [16] Sureau C, Negro F. The hepatitis delta virus: replication and pathogenesis. *J Hepatol* 2016;64(Suppl 1):S102–16.
- [17] European Association for the Study of the Liver EASL 2017 clinical practice guidelines on the management of hepatitis B virus infection. *J Hepatol* 2017;67:370–98.
- [18] Gozlan J, Lacombe K, Gault E, Raguin G, Girard PM. Complete cure of HBV–HDV co-infection after 24 weeks of combination therapy with pegylated interferon and ribavirin in a patient co-infected with HBV/HCV/HDV/HIV. *J Hepatol* 2009;50:432–4.

- [19] Yurdaydin C, Abbas Z, Buti M, Cornberg M, Esteban R, Etzioni O, et al. Treating chronic hepatitis delta: the need of surrogate markers of treatment efficacy. *J Hepatol* 2019;70:1008–15.
- [20] Le Gal F, Brichler S, Sahli R, Chevret S, Gordien E. International external quality assessment of hepatitis delta virus RNA quantification in plasma. *Hepatology* 2016;64:1483–94.
- [21] Farci P, Roskams T, Chessa L, Peddis G, Mazzoleni AP, Scioscia R, et al. Long-term benefit of interferon alpha therapy of chronic hepatitis D: regression of advanced hepatic fibrosis. *Gastroenterology* 2004;126:1740–9.
- [22] Yurdaydin C, Keskin O, Kalkan Ç, Karakaya F, Çaliskan A, Kabaçam G, et al. Effect on the natural course of the disease. *J Infect Dis* 2018;217:1184–92.
- [23] Wranke A, Serrano BC, Heidrich B, Kirschner J, Bremer B, Lehmann P, et al. Antiviral treatment and liver-related complications in hepatitis delta. *Hepatology* 2017;65:414–25.
- [24] Romeo R, Del Ninno E, Rumi M, Russo A, Sangiovanni A, de Franchis R, et al. A 28-year study of the course of hepatitis delta infection: a risk factor for cirrhosis and hepatocellular carcinoma. *Gastroenterology* 2009;136:1629–38.
- [25] Manesis EK, Vourli G, Dalekos G, Vasiliadis T, Manolaki N, Hounta A, et al. Prevalence and clinical course of hepatitis delta infection in Greece: a 13-year prospective study. *J Hepatol* 2013;59:949–56.
- [26] Niro GA, Smedile A, Ippolito AM, Ciancio A, Fontana R, Olivero A, et al. Outcome of chronic delta hepatitis in Italy: a long-term cohort study. *J Hepatol* 2010;53:834–40.
- [27] Abbas Z, Khan MA, Salih M, Jafri W. Interferon alpha for chronic hepatitis D. *Cochrane Database Syst Rev* 2011(12):CD006002.
- [28] Castelnauc C, Le Gal F, Ripault MP, Gordien E, Martinot-Peignoux M, Boyer N, et al. Efficacy of peginterferon alpha-2b in chronic hepatitis delta: relevance of quantitative RT-PCR for follow-up. *Hepatology* 2006;44:728–35.
- [29] Niro GA, Ciancio A, Gaeta GB, Smedile A, Marrone A, Olivero A, et al. Pegylated interferon alpha-2b as monotherapy or in combination with ribavirin in chronic hepatitis delta. *Hepatology* 2006;44:713–20.
- [30] Erhardt A, Gerlich W, Starke C, Wend U, Donner A, Sagir A, et al. Treatment of chronic hepatitis delta with pegylated interferon-alpha2b. *Liver Int* 2006;26:805–10.
- [31] Gheorghe L, Iacob S, Simionov I, Vadan R, Constantinescu I, Caruntu F, et al. Weight-based dosing regimen of peg-interferon α -2b for chronic hepatitis delta: a multicenter Romanian trial. *J Gastrointest Liver Dis* 2011;20:377–82.
- [32] Ormeci N, Bölükbas F, Erden E, Coban S, Ekiz F, Erdem H, et al. Pegylated interferon alpha-2B for chronic delta hepatitis: 12 versus 24 months. *Hepatogastroenterology* 2011;58:1648–53.
- [33] Wedemeyer H, Yurdaydin C, Dalekos GN, Erhardt A, Çakaloğlu Y, Değertekin H, et al. Peginterferon plus adefovir versus either drug alone for hepatitis delta. *N Engl J Med* 2011;364:322–31.
- [34] Samiullah S, Bikharam DN, Asreen. Treatment of chronic hepatitis delta virus with peg-interferon and factors that predict sustained viral response. *World J Gastroenterol* 2012;18:5793–8.
- [35] Karaca C, Soyer OM, Baran B, Ormeci AC, Gokturk S, Aydin E, et al. Efficacy of pegylated interferon- α treatment for 24 months in chronic delta hepatitis and predictors of response. *Antivir Ther* 2013;18:561–6.
- [36] Wedemeyer H, Yurdaydin C, Hardtke S, Caruntu FA, Curescu MG, Yalcin K, et al. Peginterferon alfa-2a plus tenofovir disoproxil fumarate for hepatitis D (HIDIT-II): a randomised, placebo controlled, phase 2 trial. *Lancet Infect Dis* 2019;19:275–86.
- [37] Heller T, Rotman Y, Koh C, Clark S, Haynes-Williams V, Chang R, et al. Long-term therapy of chronic delta hepatitis with peginterferon alfa. *Aliment Pharmacol Ther* 2014;40:93–104.
- [38] Borzacov LM, de Figueiredo Nicolette LD, Souza LF, Dos Santos AO, Vieira DS, Salcedo JM. Treatment of hepatitis delta virus genotype 3 infection with peg-interferon and entecavir. *Int J Infect Dis* 2016;46:82–8.
- [39] Bahcecioglu IH, Ispiroglu M, Demirel U, Yaniz M. Pegylated interferon α therapy in chronic delta hepatitis: a one-center experience. *Hepat Mon* 2015;15:e24366.
- [40] Yurdaydin C, Keskin O, Kalkan Ç, Karakaya F, Çaliskan A, Kabaçam G, et al. Interferon treatment duration in patients with chronic delta hepatitis and its effect on the natural course of the disease. *J Infect Dis* 2018;217:1184–92.
- [41] Heidrich B, Yurdaydin C, Kabaçam G, Ratsch BA, Zachou K, Bremer BHIDIT-1 Study Group. Late HDV RNA relapse after peginterferon alpha-based therapy of chronic hepatitis delta. *Hepatology* 2014;60:87–97.
- [42] Niro GA, Smedile A, Fontana R, Olivero A, Ciancio A, Valvano MR, et al. HBsAg kinetics in chronic hepatitis D during interferon therapy: on-treatment prediction of response. *Aliment Pharmacol Ther* 2016;44:620–8.
- [43] Keskin O, Wedemeyer H, Tuzün A, Zachou K, Deda X, Dalekos GN, et al. Association between level of hepatitis D virus RNA at Week 24 of pegylated interferon therapy and outcome. *Clin Gastroenterol Hepatol* 2015;13:2342–9.
- [44] Lutterkort GL, Wranke A, Hengst J, Yurdaydin C, Stift J, Bremer B, et al. Viral dominance patterns in chronic hepatitis delta determine early response to interferon alpha therapy. *J Viral Hepat* 2018;25:1384–94.
- [45] Ricco G, Popa D.C., Cavallone D., Iacob S., Salvati A., Tabacelia D., et al. Quantification of serum markers of hepatitis B (HBV) and delta virus (HDV) infections in patients with chronic HDV infection. *J Viral Hepat* 2018;25:911–9.
- [46] Niro GA, Ciancio A, Tillman HL, Lagget M, Olivero A, Perri F, et al. Lamivudine therapy in chronic delta hepatitis: a multicentre randomized-controlled pilot study. *Aliment Pharmacol Ther* 2005;22:227–32.
- [47] Canbakan B, Senturk H, Tabak F, Akdogan M, Tahan V, Mert A, et al. Efficacy of interferon alpha-2b and lamivudine combination treatment in comparison to interferon alpha-2b alone in chronic delta hepatitis: a randomized trial. *J Gastroenterol Hepatol* 2006;21:657–63.
- [48] Yurdaydin C, Bozkaya H, Onder FO, Sentürk H, Karaaslan H, Akdoğan M, et al. Treatment of chronic delta hepatitis with lamivudine vs lamivudine + interferon vs interferon. *J Viral Hepat* 2008;15:314–2.
- [49] Kabaçam G, Onder FO, Yakut M, Seven G, Karatayli SC, Karatayli E, et al. Entecavir treatment of chronic hepatitis D. *Clin Infect Dis* 2012;55:645–50.
- [50] Abbas Z, Memon MS, Umer MA, Abbas M, Shazi L. Co-treatment with pegylated interferon alfa-2a and entecavir for hepatitis D: a randomized trial. *World J Hepatol* 2016;8:625–31.
- [51] Brancaccio G, Fasano M, Grossi A, Santantonio TA, Gaeta GB. Clinical outcomes in patients with hepatitis D, cirrhosis and persistent hepatitis B virus replication, and receiving long-term tenofovir or entecavir. *Aliment Pharmacol Ther* 2019;49:1071–6. doi:10.1111/apt.15188.
- [52] Yan H, Zhong G, Xu G, He W, Jing Z, Gao Z, et al. Sodium taurocholate cotransporting polypeptide is a functional receptor for human hepatitis B and D virus. *Elife* 2012;1:e00049.
- [53] Schieck A, Müller T, Schulze A, Haberkorn U, Urban S, Mier W. Solid-phase synthesis of the lipopeptide Myr-HBVpreS/2-78, a hepatitis B virus entry inhibitor. *Molecules* 2010;15:4773–83.
- [54] Bogomolov P, Alexandrov A, Voronkova N, Macievich M, Kokina K, Petrachenkova M, et al. Treatment of chronic hepatitis D with the entry inhibitor myrcludex B: first results of a phase Ib/IIa study. *J Hepatol* 2016;65:490–8.
- [55] Wedemeyer H, Bogomolov P, Blank A, Allweiss L, Dandri-Petersen M, Bremer B, et al. Final results of a multicenter, open-label phase 2b clinical trial to assess safety and efficacy of myrcludex B in combination with tenofovir in patients with chronic HBV/HDV infection. *J Hepatol* 2018;68(Suppl 1):S3.
- [56] Loglio A, Ferenci P, Uceda Renteria SC, Tham CYL, van Bömmel F, Borghi M, et al. Excellent safety and effectiveness of high-dose myrcludex-b monotherapy administered for 48 weeks in HDV-related compensated cirrhosis: a case report of 3 patients. *J Hepatol* 2019;71:834–9. doi:10.1016/j.jhep.2019.07.003.
- [57] Bordier BB, Marion PL, Ohashi K, Kay MA, Greenberg HB, Casey JL, et al. A prenylation inhibitor prevents production of infectious hepatitis delta virus particles. *J Virol* 2002;76:10465–72.
- [58] Bordier BB, Ohkanda J, Liu P, Lee SY, Salazar FH, Marion PL, et al. In vivo antiviral efficacy of prenylation inhibitors against hepatitis delta virus. *J Clin Invest* 2003;112:407–14.
- [59] Koh C, Canini L, Dahari H, Zhao X, Uprichard SL, Haynes-Williams V, et al. Oral prenylation inhibition with lonafarnib in chronic hepatitis D infection: a proof-of-concept randomised, double-blind, placebo-controlled phase 2A trial. *Lancet Infect Dis* 2015;15:1167–74.
- [60] Yurdaydin C, Keskin O, Kalkan Ç, Karakaya F, Çaliskan A, Karatayli E, et al. Optimizing lonafarnib treatment for the management of chronic delta hepatitis: the LOWR HDV-1 study. *Hepatology* 2018;67:1224–36.
- [61] Vaillant A. Nucleic acid polymers: broad spectrum antiviral activity, antiviral mechanisms and optimization for the treatment of hepatitis B and hepatitis D infection. *Antiviral Res* 2016;133:32–40.
- [62] Quinet J, Jamard C, Burtin M, Lemasson M, Guerret S, Sureau C, et al. Nucleic acid polymer REP 2139 and nucleos(T)ide analogues act synergistically against chronic hepadnaviral infection in vivo in Pekin ducks. *Hepatology* 2018;67:2127–40.
- [63] Bazinet M, Pântea V, Cebotarescu V, Cojuhari L, Jimbei P, Albrecht J, et al. Safety and efficacy of REP 2139 and pegylated interferon alfa-2a for treatment-naïve patients with chronic hepatitis B virus and hepatitis D virus co-infection (REP 301 and REP 301-LTF): a non-randomized, open label, phase 2 trial. *Lancet Gastroenterol Hepatol* 2017;2:877–89 Erratum in: *Lancet Gastroenterol Hepatol* 2018;3:e1.
- [64] Giersch K, Homs M, Volz T, Helbig M, Allweiss L, Lohse AW, et al. Both interferon alpha and lambda can reduce all intrahepatic HDV infection markers in HBV/HDV infected humanized mice. *Sci Rep* 2017;7:3757.
- [65] Hamid SS, Etzioni O, Lurie Y, et al. A phase 2 randomized clinical trial to evaluate the safety and efficacy of pegylated interferon lambda monotherapy in patients with chronic hepatitis delta virus infection. Interim results from the LIMT HDV Study (abstract). *Hepatology* 2017;66:496A.