



Letter to the Editor

Chromosome-mediated *mcr-1* in *Escherichia coli* strain L73 from a goose

Sir,

Colistin is regarded as the drug of last resort to treat infections caused by multidrug-resistant (MDR) Gram-negative bacteria. Plasmid-mediated colistin resistance gene *mcr-1* was discovered in China in 2015 [1]. Since then, the following *mcr* genes have been described: *mcr-2* (LT598652), *mcr-3* (KY924928), *mcr-5* (KY807921), *mcr-6* (MF176240), *mcr-7* (MG267386) and *mcr-8* (MG736312). Mobile colistin resistance genes have received wide public attention; however, chromosome-mediated colistin resistance genes should not be overlooked. Although *mcr-1* has been reported in different sources, including in animals and humans, there is no report of *mcr-1* in the goose, which is an important farming animal in Jiangsu, China. In this study, we report a colistin-resistant *Escherichia coli* encoding a chromosomal *mcr-1* from a goose anus swab and characterize the genome with MinION long reads.

A total of 96 samples were collected from geese with no manifestation of disease from a goose farming center in Yangzhou, Jiangsu Province, China in 2017. A total of 162 Enterobacteriaceae isolates were obtained from these samples via screening on MacConkey Agar plates. *mcr* genes were identified by the multiplex polymerase chain reaction (PCR) method reported previously [2]. One isolate, L73, identified as *E. coli* by 16S rDNA sequencing, was positive for *mcr-1*. Antimicrobial susceptibility testing was performed on the *mcr-1*-positive *E. coli* using the broth microdilution method and interpreted according to the EUCAST breakpoints for Enterobacteriaceae. *E. coli* ATCC 25922 was the quality control. *E. coli* L73 was resistant to colistin (4 mg/L) and exhibited low susceptibility to streptomycin, doxycycline, amoxicillin, and florfenicol. Conjugation assay failed to transfer the colistin resistance phenotype to the azide-resistant *E. coli* J53 strain, which indicated the *mcr-1* in L73 was not in conjugative elements. Whole genome sequencing (WGS) was performed to genetically characterize *mcr-1*.

The genomic DNA of *E. coli* strain L73 was extracted using a TIANamp Bacteria DNA Kit. WGS was performed via Illumina and ONT MinION platforms. De novo assembly was performed by short-read and long-read data [3]. The Rapid Annotation using Subsystem Technology (RAST) was used to annotate the genome. Sequence analysis revealed that L73 harbored one chromosome (ST48) and two plasmids. *mcr-1* in L73 is located on the chromosome. A chromosomal segment containing *mcr-1* was extracted and BLASTn analysis was performed. The gene *yjbs* had been disrupted by insertion of the IS*Apl1*-*mcr-1*-orf-IS*Apl1* (Tn6330) exhibiting 100% identity to the Tn6330 in the *E. coli* HS30-1 chromosome (Figure 1a). The surrounding sequences of Tn6330 in L73 exhibited 99% identity (97% coverage) to the chromosome of *E. coli* str. K-12 (CP032667.1). This indicated that insertion

of Tn6330 occurred previously. A pair of reverse primers targeting *mcr-1* was used to test the ability of Tn6330 to generate circular intermediate as previously described [4]. Primers Tn-F (TTTCTGATTGACCAGCCCC) and Tn-R (CACCTTAACCTCTCG-GCA) targeting the surrounding sequences of Tn6330 in L73 were utilized to investigate whether the intact Tn6330 was consistently present in the bacterial population. The sequences of PCR products were obtained by Sanger sequencing. Gel electrophoresis showed there were two PCR products (~5 kb and ~1.4 kb) (Figure 1c), which indicated Tn6330 was unstable and still active, as described previously [4]. Tn6330 in *E. coli* L73 could generate a circular intermediate (IS*Apl1*-*mcr-1*-orf) (Figure 1d), which was detectable in the form of a ~2.5 kb amplicon (Figure 1e). A previous study indicated the IS*Apl1* upstream of *mcr-1* participated in the formation of the *mcr-1*-orf-IS*Apl1* circular intermediate on its own or via recombination with the IS*Apl1* downstream of *mcr-1* [5]. Based on the genetic context of *mcr-1*, the *mcr-1* was likely mobilized into this chromosome via the IS*Apl1* element.

WGS showed there were two circular plasmids, pL73-2 and pL73-3, in *E. coli* L73. pL73-2 was a 167 170-bp, IncN plasmid sharing 99% identity (41% coverage) to the sequence of *E. coli* strain WCHEC050613 plasmid pP_WCHEC050613 (CP019216.2) and 99% identity (41% coverage) to the sequence of *E. coli* strain AR_0157 plasmid (CP029729.1), and with an overall GC content of 50.48%. Multiple resistance genes, including *qnrS1*, *bla*_{TEM-1B}, *aph(6)-Ia*, *aph(3'')-Ib*, *sul2*, *dfrA14*, *floR* and two *tet(A)* genes, were observed in pL73-2 (Figure 1f). No resistance gene was found in plasmid pL73-3.

In conclusion, a chromosomal *mcr-1*-positive *E. coli* strain in the form of a composite transposon was identified from the goose. Nanopore sequencing can be utilized to accelerate the analysis of resistance genes in genomes without S1-pulsed-field gel electrophoresis (S1-PFGE) and Southern blot. To the best of our knowledge, this is the first report of an *mcr-1*-bearing *E. coli* from the goose. Active surveillance is essential to prevent further dissemination of *mcr-1*.

Accession numbers. The complete genome of *E. coli* strain L73 has been deposited in GenBank (CP033378, CP033379 and CP033380).

Declarations

None to declare.

Funding

This work was supported by the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD), Na-

tional Science Foundation of China (31872526 and 31872523) and the Natural Science Foundation of Jiangsu Province (BK20180900). We thank Jiayu Xiao and Ziyi Liu in Wang's Lab for sample collection.

Competing Interests

None to declare.

Ethical Approval

None to declare.

References

- [1] Liu YY, Wang Y, Walsh TR, Yi LX, Zhang R, Spencer J, et al. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. *Lancet Infect Dis* 2016;16:161–8.
- [2] Rebelo AR, Bortolaia V, Kjeldgaard JS, Pedersen SK, Leekitcharoenphon P, Hansen IM, et al. Multiplex PCR for detection of plasmid-mediated colistin resistance determinants, mcr-1, mcr-2, mcr-3, mcr-4 and mcr-5 for surveillance purposes. *Euro Surveill* 2018;23.
- [3] Li R, Xie M, Dong N, Lin D, Yang X, Wong MHY, et al. Efficient generation of complete sequences of MDR-encoding plasmids by rapid assembly of MinION barcoding sequencing data. *Gigascience* 2018;7:1–9.
- [4] Li R, Xie M, Lv J, Wai-Chi Chan E, Chen S. Complete genetic analysis of plasmids carrying mcr-1 and other resistance genes in an *Escherichia coli* isolate of animal origin. *J Antimicrob Chemother* 2016;72:696–9.
- [5] Zhao F, Feng Y, Lu X, McNally A, Zong Z. Remarkable diversity of *Escherichia coli* carrying mcr-1 from hospital sewage with the identification of two new mcr-1 variants. *Front Microbiol* 2017;8:2094.

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Received 23 November 2018

Accepted 2 March 2019