



In vitro susceptibility of urinary *Escherichia coli* isolates to first- and second-line empirically prescribed oral antimicrobials: CANWARD surveillance study results for Canadian outpatients, 2007–2016[☆]

James A. Karlowsky^{a,b}, Philippe R.S. Lagacé-Wiens^{a,b}, Heather J. Adam^{a,b},
Melanie R. Baxter^a, Nancy M. Laing^a, Andrew J. Walkty^{a,b}, George G. Zhanel^{a,*}

^a Department of Medical Microbiology and Infectious Diseases, Max Rady College of Medicine, University of Manitoba, Winnipeg, Manitoba, Canada

^b Diagnostic Services, Shared Health, Winnipeg, Manitoba, Canada

ARTICLE INFO

Article history:

Received 22 November 2018

Accepted 23 April 2019

Keywords:

Escherichia coli

Urine

Outpatient

Susceptibility

CANWARD

ABSTRACT

Escherichia coli isolates ($n=2035$) from urine specimens of outpatients presenting to Canadian medical clinics and hospital emergency departments from 2007–2016 were collected as part of the CANWARD surveillance study. Isolate identification and antimicrobial susceptibility testing (AST) were performed at a central site (Health Sciences Centre, Winnipeg, Canada). AST of first- and second-line oral antimicrobial agents was performed using CLSI methods (M07, 11th ed, 2018); fosfomycin was tested by agar dilution and all other agents by broth microdilution. Minimum inhibitory concentrations (MICs) were interpreted using CLSI M100 (2018) criteria. Fosfomycin (99.2% of isolates susceptible), nitrofurantoin (97.5%) and cefalexin (93.6%) were the most active agents tested; amoxicillin/clavulanic acid (AMC) (85.6%), ciprofloxacin (83.0%) and trimethoprim/sulfamethoxazole (SXT) (77.0%) were less active. Annual percentages of isolates positive for extended-spectrum β -lactamases (ESBLs) or demonstrating multidrug-resistant (MDR) phenotypes increased from 0.8% (2007) to 10.1% (2016), and from 9.7% (2007) to 16.5% (2016), respectively, whilst the annual frequency of AmpC-positive isolates decreased from a high of 3.2% in 2008 to 0.7% in 2016. The most common MDR phenotype of *E. coli* was non-susceptibility to AMC, ciprofloxacin, and SXT, accounting for 12.7% (26/205) of all MDR isolates. Rates of susceptibility were higher for fosfomycin than for the five other oral agents tested against ESBL-positive (96.1% susceptible) and MDR (95.1%) isolates and were equal to nitrofurantoin (96.4%) against AmpC-positive isolates. Prudent use of antimicrobials and close monitoring of antimicrobial susceptibilities of clinical uropathogenic *E. coli* isolates are imperative to help preserve the utility of oral antimicrobials.

© 2019 Elsevier B.V. and International Society of Chemotherapy. All rights reserved.

1. Introduction

Urinary tract infections (UTIs) are among the most common bacterial infections in humans [1–5]. Uropathogenic *Escherichia coli* cause the majority of UTIs both in outpatients and inpatients [1–8]. Urine culture remains the gold standard for the identification of pathogens causing UTIs in symptomatic patients [1,9]; however, it is not recommended for otherwise healthy, non-pregnant, reproductive-age women presenting with symptomatic acute

uncomplicated cystitis as culture results would not alter patient management or outcome in this patient group. Instead, such patients are empirically prescribed oral therapy at their care provider visit. In the majority of these patients, empirical therapy is successful as there is a relatively low likelihood of encountering a resistant uropathogen and the risk of complications with this approach is low [1,5,7]. Urine culture and antimicrobial susceptibility testing (AST) of bacterial pathogens is generally only performed following failure of empirical therapy or in cases of upper UTI (e.g. pyelonephritis), recurrent infection, where there is a risk of infection with antimicrobial-resistant organisms, or for patients with complicated UTI (e.g. urinary obstruction, immunosuppression) given the potential for less common uropathogens, antimicrobial-resistant isolates and higher risk of complications in these patients [5,9]. Treatment of asymptomatic bacteriuria is only indicated for pregnant women and patients undergoing a genitourinary procedure [10,11].

[☆] This work was presented in part at the ASM Microbe 2018 Conference, 7–11 June 2018, Atlanta, GA [abstract Saturday-416].

* Corresponding author. Present address: Clinical Microbiology, Health Sciences Centre, MS673-820 Sherbrook Street, Winnipeg, Manitoba, R3A 1R9, Canada. Tel: +1 204 787 4902.

E-mail address: ggzhanel@pcs.mb.ca (G.G. Zhanel).

The Infectious Diseases Society of America (IDSA) currently recommends an empirical regimen of 5 days of nitrofurantoin, a 3-day course of double-strength trimethoprim/sulfamethoxazole (SXT) in settings where the prevalence of SXT resistance is <10–20%, or a 3 g single dose of fosfomycin trometamol for treating acute bacterial cystitis in otherwise healthy, adult, non-pregnant females; fluoroquinolones and oral β -lactams, e.g. amoxicillin/clavulanic acid (AMC) and cefalexin, are second-line therapies [5]. Empirical treatment choices should ideally be based on local cumulative AST data. Fosfomycin and nitrofurantoin are only approved for the treatment of uncomplicated UTIs. Discrepancies between fosfomycin minimum inhibitory concentrations (MICs) determined by the reference agar dilution method [12,13] and by currently available commercial methods have been identified [14,15].

Over the last 20 years, antimicrobial resistance among uropathogens, including uropathogenic *E. coli*, has become increasingly common and is an important consideration in the management of outpatient UTIs [7,16–19]. In the USA, multidrug-resistant (MDR) phenotypes among outpatient urine isolates of *E. coli* increased from 9% in 2001 to 17% in 2010 [17]. Care providers should be cognisant of antimicrobial resistance rates provided in their local antibiogram as well as clinical and epidemiological risk factors associated with MDR infections as treatment options may be limited and the presence of unanticipated resistant or MDR pathogens can lead to ineffective empirical prescribing and subsequently to poor clinical outcomes with low microbiological and clinical cure rates and increased patient morbidity and mortality [7,17,20].

Reports providing antimicrobial susceptibility rates for uropathogens in outpatients receiving initial empirical therapy for community-acquired UTI are very limited because the majority of these patients (i.e. healthy, reproductive-age women presenting with symptomatic acute uncomplicated cystitis) are not requested to submit a urine specimen for culture [5,21]. Therefore, it should be assumed that many reports describing antimicrobial susceptibility of uropathogens in outpatients likely include isolates that are the result of treatment failures, recurrent infection, upper UTI or complicated infection [6,16–18,22–26]. AST surveillance data for uropathogenic *E. coli* isolated from Canadian patients were last published for isolates collected between 2010 and 2013 [22]. In the current study, uropathogenic *E. coli* isolates recovered from outpatients seeking care at medical clinics and hospital emergency departments in Canada from 2007–2016 were tested. The intent was to assess current rates of resistance to all commonly tested and reported empirically prescribed oral first- and second-line antimicrobial agents [5] as well as to document changes in susceptibility to these agents over time.

2. Materials and methods

2.1. Bacterial isolates

The *E. coli* isolates tested in this study ($n=2035$) were cultured from urine specimens of outpatients attending primary care and specialty medical clinics as well as hospital emergency departments and submitted to the annual CANWARD surveillance study from 2007–2016 [22,27]. If isolate identification at the coordinating laboratory (Health Sciences Centre, Winnipeg, Canada) [28] was not consistent with the identification provided by the submitting laboratory, the isolate was removed from the study. The low number of outpatient urine isolates per year (203–601 isolates/year; mean, 330 isolates/year) (Table 1) collected by the CANWARD surveillance study reflects that these isolates are not the sole focus of the study, however, we believe they are an important source of data to inform on changes in the prevalence of antimicrobial

resistance in uropathogenic *E. coli* infecting Canadian outpatients.

2.2. Antimicrobial susceptibility testing (AST)

AST of fosfomycin was performed by the Clinical and Laboratory Standards Institute (CLSI) agar dilution method with Mueller-Hinton agar supplemented with 25 $\mu\text{g}/\text{mL}$ glucose-6-phosphate; all other antimicrobial agents were tested in cation-adjusted Mueller-Hinton broth using in-house-prepared 96-well broth microdilution panels prepared according to CLSI guidelines [12,13]. Quality control was performed following CLSI recommendations, and MICs were interpreted using CLSI M100 breakpoints [12]. Cefazolin was tested as a surrogate for cefalexin as permitted by the CLSI M100 standard [12]. Extended-spectrum β -lactamase (ESBL)-producing *E. coli* isolates were screened for and were confirmed phenotypically following the CLSI method [12]. *Escherichia coli* isolates were deemed AmpC β -lactamase-positive if they demonstrated an MIC of ≥ 1 $\mu\text{g}/\text{mL}$ for ceftriaxone and/or ceftazidime (data not shown) and/or aztreonam (data not shown), an MIC of ≥ 32 $\mu\text{g}/\text{mL}$ for cefoxitin (data not shown) and were phenotypically negative for ESBL production [29]. MDR *E. coli* isolates were defined by non-susceptibility to agents from three or more antimicrobial classes according to a published guideline [30].

2.3. Statistical analysis

For the purpose of statistical analysis, isolates were defined as either susceptible or non-susceptible to an antimicrobial agent; isolates testing as intermediate were included in the non-susceptible group. A second-degree factorial multivariate stepwise regression analysis using available variables was performed to identify variables associated with non-susceptibility of *E. coli* isolates in this study. The following variables were included in the model: year of study; patient age; patient sex; and patient location (emergency room or hospital outpatient clinic). Initially, a stepwise regression model was created including all of the variables and second-degree interactions. To exclude variables without a significant impact on the model, variables were excluded from the final model in such a way as to minimise the corrected Akaike Information Criterion. Finally, a regression model was created including only the variables and interactions with a significant impact on resistance to an antimicrobial agent. In the multivariate model, a P -value of ≤ 0.05 was considered statistically significant.

Differences between fosfomycin and nitrofurantoin resistance within phenotypic subgroups of *E. coli* were assessed using Fisher's exact test. Significance was defined as $P < 0.05$ with a Bonferroni correction for multiple comparisons. Six phenotype subgroups were assessed (ESBL-positive, MDR, and non-susceptible to ciprofloxacin, SXT, cefalexin or AMC), therefore the P -value for significance was corrected to $P < 0.008$ for subgroup analyses. Statistical analysis was performed using JMP 14 software (SAS Institute Inc., Cary, NC).

3. Results

Escherichia coli was the most common urinary pathogen isolated in each year from 2007–2016 accounting for between 56.6% (2015) and 65.8% (2009) of isolates per annum (overall, 61.7%); the second most common pathogen isolated was *Klebsiella pneumoniae*, which accounted for approximately 10% of isolates per year [range, 6.7% (2008) to 12.9% (2013) per annum] (Table 1). Enterobacteriaceae accounted for between 77.2% (2007) and 84.7% (2016) of isolates per annum (overall, 80.4%). Gram-positive cocci (enterococci

Table 1

Annual prevalence of bacterial species isolated from urine specimens of outpatients in Canada, 2007–2016.

| Year (total n) | n (% of annual n) | | | | | | | |
|----------------|-------------------------|-------------------------------------|----------------------------------|---------------------------------------|--|-------------------------------|---------------------------------------|---|
| | <i>Escherichia coli</i> | <i>Klebsiella</i> spp. ^a | <i>Proteus</i> spp. ^b | <i>Enterobacter</i> spp. ^c | Other species of Enterobacteriaceae ^d | <i>Pseudomonas aeruginosa</i> | <i>Enterococcus</i> spp. ^e | <i>Staphylococcus</i> spp. ^f |
| 2016 (215) | 139 (64.7) | 27 (12.6) | 7 (3.3) | 6 (2.8) | 3 (1.4) | 4 (1.9) | 15 (7.0) | 9 (4.2) |
| 2015 (228) | 129 (56.6) | 29 (12.7) | 10 (4.4) | 4 (1.8) | 7 (3.1) | 3 (1.3) | 17 (7.5) | 20 (8.8) |
| 2014 (207) | 130 (62.8) | 19 (9.2) | 12 (5.8) | 2 (1.0) | 4 (1.9) | 5 (2.4) | 24 (11.6) | 6 (2.9) |
| 2013 (255) | 146 (57.3) | 33 (12.9) | 8 (3.1) | 11 (4.3) | 8 (3.1) | 5 (2.0) | 24 (9.4) | 14 (5.5) |
| 2012 (203) | 119 (58.6) | 26 (12.8) | 9 (4.4) | 6 (3.0) | 8 (3.9) | 5 (2.5) | 17 (8.4) | 10 (4.9) |
| 2011 (244) | 145 (59.4) | 27 (11.1) | 9 (3.7) | 5 (2.0) | 9 (3.7) | 3 (1.2) | 20 (8.2) | 15 (6.1) |
| 2010 (441) | 271 (61.5) | 42 (9.5) | 12 (2.7) | 12 (2.7) | 11 (2.5) | 6 (1.4) | 55 (12.5) | 20 (4.5) |
| 2009 (471) | 310 (65.8) | 46 (9.8) | 15 (3.2) | 11 (2.3) | 11 (2.3) | 5 (1.1) | 37 (7.9) | 17 (3.6) |
| 2008 (434) | 285 (65.7) | 29 (6.7) | 18 (4.1) | 7 (1.6) | 13 (3.0) | 8 (1.8) | 46 (10.6) | 20 (4.6) |
| 2007 (601) | 361 (60.1) | 59 (9.8) | 19 (3.2) | 6 (1.0) | 19 (3.2) | 11 (1.8) | 70 (11.6) | 32 (5.3) |
| All (3299) | 2035 (61.7) | 337 (10.2) | 119 (3.6) | 70 (2.1) | 93 (2.8) | 55 (1.7) | 325 (9.9) | 163 (4.9) |

^a The 337 *Klebsiella* spp. isolates comprised *Klebsiella pneumoniae* ($n=275$), *Klebsiella oxytoca/Raoultella* spp. ($n=54$) and unspciated *Klebsiella* ($n=8$).^b The 119 *Proteus* spp. isolates comprised *Proteus mirabilis* ($n=116$) and *Proteus vulgaris* ($n=3$).^c The 70 *Enterobacter* spp. isolates comprised *Enterobacter cloacae* ($n=44$), *Enterobacter aerogenes* ($n=23$), *Enterobacter agglomerans* ($n=1$), *Enterobacter asburiae* ($n=1$) and unspciated *Enterobacter* ($n=1$).^d The 93 other species of Enterobacteriaceae comprised *Citrobacter freundii* ($n=27$), *Morganella morganii* ($n=20$), *Citrobacter koseri* ($n=15$), *Serratia marcescens* ($n=9$), *Citrobacter amaloniticus* ($n=5$), *Providencia stuartii* ($n=5$), *Citrobacter braakii* ($n=2$), unspciated *Citrobacter* ($n=2$), *Hafnia alvei* ($n=2$), *Providencia rettgeri* ($n=2$), *Citrobacter diversus* ($n=1$), *Lelliottia amnigena* ($n=1$), *Salmonella enterica* ($n=1$) and *Serratia fonticola* ($n=1$).^e The 325 *Enterococcus* spp. isolates comprised *Enterococcus faecalis* ($n=164$), unspciated *Enterococcus* ($n=152$), *Enterococcus faecium* ($n=7$), *Enterococcus avium* ($n=1$) and *Enterococcus casseliflavus* ($n=1$).^f The 163 *Staphylococcus* spp. isolates comprised *Staphylococcus aureus* ($n=75$), unspciated coagulase-negative staphylococci ($n=48$), *Staphylococcus saprophyticus* ($n=33$), *Staphylococcus epidermidis* ($n=6$) and *Staphylococcus hominis* ($n=1$).**Table 2**Annual rates of in vitro susceptibility of first- and second-line empirically prescribed oral antimicrobial agents for outpatient urine isolates of *Escherichia coli* in Canada, 2007–2016.

| Year (n) | MIC ₉₀ (µg/mL)/% susceptible | | | | | |
|------------|---|------------------|----------|--------|---------|---------|
| | AMC | LEX ^a | CIP | FOS | NIT | SXT |
| 2016 (139) | 16/79.1 | >128/88.5 | >16/79.9 | 4/99.3 | 16/97.8 | >8/77.7 |
| 2015 (129) | 16/83.7 | 16/92.2 | >16/77.5 | 2/99.2 | 16/97.7 | >8/73.6 |
| 2014 (130) | 16/83.7 | >128/87.6 | >16/78.3 | 2/100 | 16/98.5 | >8/71.3 |
| 2013 (146) | 16/86.3 | 8/94.5 | 16/84.2 | 4/100 | 32/97.9 | >8/80.1 |
| 2012 (119) | 16/79.0 | 8/94.1 | >16/83.2 | 4/99.2 | 32/96.6 | >8/77.3 |
| 2011 (145) | 16/86.8 | 64/88.3 | >16/73.8 | 2/99.3 | 32/97.2 | >8/70.8 |
| 2010 (271) | 16/82.3 | 4/97.0 | >16/83.0 | 4/99.6 | 32/96.7 | >8/75.6 |
| 2009 (310) | 8/94.8 | 8/95.8 | >16/85.8 | 4/99.4 | 16/98.7 | >8/74.2 |
| 2008 (285) | 8/97.5 | 8/91.9 | >16/83.5 | 4/97.2 | 32/96.5 | >8/76.8 |
| 2007 (361) | 16/75.8 | 8/96.4 | >16/88.1 | 2/99.4 | 32/97.5 | >8/84.5 |
| All (2035) | 16/85.6 | 8/93.6 | >16/83.0 | 4/99.2 | 32/97.5 | >8/77.0 |

MIC₉₀, minimum inhibitory concentration required to inhibit 90% of the isolates; AMC, amoxicillin/clavulanic acid; LEX, cefalexin; CIP, ciprofloxacin; FOS, fosfomycin; NIT, nitrofurantoin; SXT, trimethoprim/sulfamethoxazole.^a LEX susceptibility was predicted by ceftazidime minimum inhibitory concentration (MIC) ≤ 16 µg/mL [12].

and staphylococci) accounted for between 11.2% (2016) and 17.0% (both 2007 and 2010) of isolates per annum (overall, 14.8%).

Overall from 2007–2016, fosfomycin (99.2% of isolates susceptible), nitrofurantoin (97.5%) and cefalexin (93.6%) were the most active agents against urinary *E. coli* isolates (Table 2); AMC (85.6% of isolates susceptible), ciprofloxacin (83.0%) and SXT (77.0%) were less active. Of the six antimicrobial agents tested against *E. coli*, a trend of decreasing susceptibility was observed for SXT ($P=0.0435$), cefalexin ($P=0.0004$) and ciprofloxacin ($P=0.0001$) but did not reach significance for AMC ($P=0.0881$) (Table 2). In the most recent year of data (2016), in vitro susceptibilities of urinary *E. coli* isolates to SXT (77.7%), AMC (79.1%) and ciprofloxacin (79.9%) were <80% and were compromised in comparison with fosfomycin (99.3%) and nitrofurantoin (97.8%).

Annual ESBL rates among isolates of *E. coli* were variable but increased significantly from 0.8% in 2007 to 10.1% in 2016 ($P < 0.0001$) (Table 3). MDR frequencies were also variable but increased significantly from 9.7% in 2007 to 16.5% in 2016 ($P=0.0009$), whilst AmpC-positive annual frequencies decreased non-significantly from 3.2% in 2008 to 0.7% in 2016 ($P=0.8005$).

The overall percent susceptible rate to fosfomycin was approximately 10% higher than to nitrofurantoin for ESBL-positive isolates (96.1% vs. 87.0% susceptible; $P=0.0393$) and MDR isolates (95.1% vs. 86.8% susceptible; $P=0.0026$) of *E. coli* (Table 4). The percent susceptible rate to fosfomycin was approximately 5–6% higher than to nitrofurantoin for *E. coli* isolates that were non-susceptible to SXT (98.3% vs. 94.0% susceptible; $P=0.0004$), cefalexin (96.2% vs. 90.8% susceptible; non-significant, $P=0.0652$), AMC (99.3% vs. 94.9% susceptible; $P=0.0001$) and ciprofloxacin (97.1% vs. 91.6% susceptible; $P=0.0013$).

Of the 2035 *E. coli* isolates, 1674 (82.3%) were from female patients and 539 (26.5%) were from female patients aged 18–45 years. Increasing age was significantly associated with non-susceptibility to cefalexin ($P=0.0024$), ciprofloxacin ($P < 0.0001$) and nitrofurantoin ($P=0.0001$) as well as being associated with isolation of ESBL-positive ($P=0.0021$) and MDR ($P < 0.0001$) isolates. *Escherichia coli* isolates with MDR (13.0% vs. 9.4%), ESBL-positive (5.3% vs. 3.5%) and AmpC-positive (1.9% vs. 1.3%) phenotypes were more commonly isolated from male patients than from female patients, however these differences did not reach

Table 3

Annual rates of multidrug-resistant (MDR), extended-spectrum β -lactamase (ESBL)-positive and AmpC-positive isolates among outpatient urine isolates of *Escherichia coli* in Canada, 2007–2016.

| Year (n) | % of annual n | | |
|-------------|----------------------|--------|--------|
| | % MDR ^{a,b} | % ESBL | % AmpC |
| 2016 (139) | 16.5 | 10.1 | 0.7 |
| 2015 (129) | 12.4 | 5.4 | 0.8 |
| 2014 (130) | 16.9 | 10.8 | 0.8 |
| 2013 (146) | 7.5 | 3.4 | 2.1 |
| 2012 (119) | 9.2 | 1.7 | 1.7 |
| 2011 (145) | 13.1 | 9.0 | 2.1 |
| 2010 (271) | 8.5 | 1.8 | 1.1 |
| 2009 (310) | 6.5 | 1.6 | 1.6 |
| 2008 (285) | 8.8 | 3.2 | 3.2 |
| 2007 (361) | 9.7 | 0.8 | NA |
| Mean (2035) | 10.1 | 3.8 | 1.4 |

SXT, trimethoprim/sulfamethoxazole; AMC, amoxicillin/clavulanic acid.

^a A MDR isolate was defined by a phenotype that was non-susceptible to agents from three or more antimicrobial classes [30]. The agents used to define *E. coli* isolates as MDR were gentamicin (aminoglycoside), piperacillin/tazobactam (antipseudomonal penicillin + β -lactamase inhibitor), meropenem (carbapenem) and ceftazidime [non-extended-spectrum cephalosporin; a ceftazidime minimum inhibitory concentration (MIC) of ≥ 32 μ g/mL was used to predict non-susceptibility to ceftazidime], ceftriaxone (extended-spectrum cephalosporin), ciprofloxacin (fluoroquinolone), SXT (folate pathway inhibitor), tigecycline (glycylcycline), AMC (penicillin + β -lactamase inhibitor), fosfomycin (phosphonic acid) and nitrofurantoin (nitrofurantoin) [30].

^b The five most common MDR phenotypes were: isolates non-susceptible to AMC, ciprofloxacin and SXT ($n=26$; 12.7% of MDR isolates); isolates non-susceptible to ciprofloxacin, gentamicin and SXT ($n=25$; 12.2% of MDR isolates); isolates non-susceptible to AMC, ciprofloxacin, gentamicin and SXT ($n=11$; 5.4% of MDR isolates); isolates non-susceptible to ceftazidime, ceftriaxone, ciprofloxacin, gentamicin and SXT ($n=11$; 5.4% of MDR isolates); and isolates non-susceptible to ceftazidime, ceftriaxone, ciprofloxacin and SXT ($n=10$; 4.9% of MDR isolates).

statistical significance in the multifactorial regression model because age was identified as a confounding variable that had a greater effect on non-susceptibility of isolates from males (median age 60 years) than females (median age 46 years) (Table 5). However, *E. coli* isolates non-susceptible to ciprofloxacin were significantly more common in males (24.1%) than in females (15.5%) independent of age ($P=0.0014$).

4. Discussion

Recent studies describing the antimicrobial susceptibilities of urinary *E. coli* isolates in North America and elsewhere have frequently reported rates of resistance to SXT and fluoroquinolones of >20% and rates of susceptibility to fosfomycin and nitrofurantoin in excess of 97% and 99%, respectively [6,16–18,22–26]. In addition, 10–20% of urinary *E. coli* isolates have been reported to harbour ESBLs [6,16,23,31,32] that inactivate penicillins and cephalosporins. The spread of *E. coli* sequence type 131 (ST131), which possesses a plasmid-mediated ESBL (CTX-M-14 or CTX-M-15) and chromosomal fluoroquinolone resistance-conferring mutations (in *gyrA/gyrB* genes), underlies the majority of increases in resistance noted for these two antimicrobial classes in patients both with community-onset and healthcare-associated diseases [6,16,23,31,32].

In the current study, the *E. coli* isolates tested likely represented at least partially a selected population given that only 61.7% of outpatient urinary isolates were *E. coli*, a percentage well below the expected prevalence of 75–95% [1–8]. However, these data do represent real-world isolates that can be expected to be grown from outpatient urine specimens submitted to clinical microbiology laboratories for culture and AST. We accept that the data may somewhat overestimate the prevalence of resistance,

ESBLs, AmpC β -lactamases and MDR isolates among uropathogenic *E. coli* in outpatients in Canada. Regardless of these caveats, fosfomycin (99.2% susceptible) and nitrofurantoin (97.5% susceptible) were both very active in vitro against the *E. coli* isolates tested. Current (2016) in vitro susceptibilities of urinary isolates of *E. coli* to SXT (77.7%) and ciprofloxacin (79.9%), two frequently prescribed empirical agents for outpatient UTIs, appear compromised compared with fosfomycin and nitrofurantoin. Of note, when comparing fosfomycin and nitrofurantoin, susceptibility to fosfomycin was $\geq 10\%$ higher than to nitrofurantoin both for ESBL-positive and MDR isolates of *E. coli*, and susceptibility to fosfomycin was $\geq 5\%$ higher than to nitrofurantoin for isolates of *E. coli* non-susceptible to SXT, ceftazidime, AMC and ciprofloxacin.

In the current study, fosfomycin (99.2% of isolates susceptible), nitrofurantoin (97.5%) and ceftazidime (93.6%) were observed to be the most active oral agents; AMC (85.6%), ciprofloxacin (83.0%) and SXT (77.0%) were less active. ESBL rates among isolates of *E. coli* increased from 0.8% in 2007 to 10.1% in 2016 and MDR rates increased from 9.7% in 2007 to 16.5% in 2016, whilst the annual frequency of AmpC-positive isolates decreased from a high of 3.2% in 2008 to 0.7% in 2016. Rates of susceptibility were higher for fosfomycin than for the other agents tested against ESBL-positive (96.1% susceptible) and MDR isolates (95.1% susceptible) and were equal to nitrofurantoin (96.4%) against AmpC-positive isolates. Several studies have shown that older patient age, male sex, recent hospitalisation, prior use of antimicrobials and specific geographical locations are risk factors associated with increased resistance rates to SXT, fluoroquinolones and penicillins as well as elevated rates of ESBL-producing and MDR isolates of uropathogenic *E. coli* both in community and hospitalised patients, whilst carbapenems, fosfomycin and nitrofurantoin remain effective treatments in patients with these risk factors [6,18,22–26].

This study has a few limitations that are important to identify. First, because urine specimens are not recommended to be collected from patients with uncomplicated UTI, we acknowledge that the rates of resistance calculated may overestimate the actual resistance rates in outpatients, as patients with complicated UTIs, co-morbidities or recent antimicrobial exposure, or healthcare-related infections may be present in the data set [33]. Second, we cannot account for the possible inclusion of isolates representing asymptomatic bacteriuria in the collection, which also may have had an impact on the resistance rates reported. Lastly, although we tried to restrict isolates to those that represented outpatient infections, it is possible that some isolates were from patients who had been hospitalised shortly before sample collection. Also, some of these outpatients may have received antimicrobial agents in the weeks prior to their current UTI, which potentially may have increased the isolation of resistant bacterial pathogens reported.

In conclusion, *E. coli* was the most common bacterial pathogen isolated from urine specimens of Canadian outpatients when culture was performed. The annual prevalence of Enterobacteriaceae, *P. aeruginosa* and Gram-positive cocci among urine specimens from Canadian outpatients was consistent from 2007–2016. In vitro susceptibility of urine isolates of *E. coli* to SXT was <80% for 8 of the 10 years between 2007 and 2016, and its use as empirical therapy should be reviewed based upon local antibiograms. Fosfomycin (99.2% of isolates susceptible) and nitrofurantoin (97.5% of isolates susceptible) were the most active antimicrobial agents in vitro against *E. coli* isolated from Canadian outpatients from 2007–2016. Based on these data as well as recent data generated by other investigators and expert reviews of the available literature [7,21,22,25], the activity of fosfomycin and nitrofurantoin remains high for most cases of *E. coli* UTI, including infections caused by MDR *E. coli* isolates.

Table 4

Cumulative in vitro activities of first- and second-line empirically prescribed oral antimicrobial agents against outpatient urine isolates of *Escherichia coli* in Canada, 2007–2016, stratified by resistance profile.

| Phenotype (n) | Antimicrobial agent | MIC (µg/mL) | | | CLSI M100 MIC interpretation | | |
|-------------------------------------|---------------------|-------------------|-------------------|--------------|------------------------------|-----------------|------|
| | | MIC ₅₀ | MIC ₉₀ | Range | %S | %I | %R |
| All isolates (2035) | AMC | 4 | 16 | ≤0.06 to >32 | 85.6 | 10.5 | 3.9 |
| | Cefalexin | 2 | 8 | ≤0.5 to >128 | 93.6 | NA ^a | 6.4 |
| | Ciprofloxacin | ≤0.06 | >16 | ≤0.06 to >16 | 83.0 | 0.1 | 16.9 |
| | Fosfomycin | ≤1 | 4 | ≤1 to >512 | 99.2 | 0.7 | 0.1 |
| | Nitrofurantoin | 16 | 32 | ≤1–256 | 97.5 | 1.6 | 0.9 |
| | SXT | ≤0.12 | >8 | ≤0.12 to >8 | 77.0 | NA | 23.0 |
| SXT-non-susceptible (468) | AMC | 8 | 16 | 1 to >32 | 71.4 | 22.8 | 5.8 |
| | Cefalexin | 4 | >128 | ≤0.5 to >128 | 85.3 | NA | 14.7 |
| | Ciprofloxacin | 0.25 | >16 | ≤0.06 to >16 | 56.6 | 0.5 | 42.9 |
| | Fosfomycin | ≤1 | 4 | ≤1–128 | 98.3 | 1.7 | 0 |
| | Nitrofurantoin | 16 | 32 | ≤1–256 | 94.0 | 3.4 | 2.6 |
| | SXT | >8 | >8 | 4 to >8 | 0 | NA | 100 |
| Nitrofurantoin-non-susceptible (51) | AMC | 8 | 16 | 1–32 | 70.6 | 23.5 | 5.9 |
| | Cefalexin | 4 | >128 | 1 to >128 | 76.5 | NA | 23.5 |
| | Ciprofloxacin | 16 | >16 | ≤0.06 to >16 | 43.1 | 0 | 56.9 |
| | Fosfomycin | 2 | 16 | ≤1–128 | 96.0 | 4.0 | 0 |
| | Nitrofurantoin | 64 | 128 | 64–256 | 0 | 62.7 | 37.3 |
| | SXT | >8 | >8 | ≤0.12 to >8 | 45.1 | NA | 54.9 |
| Fosfomycin-non-susceptible (17) | AMC | 4 | 16 | 2–32 | 88.2 | 5.9 | 5.9 |
| | Cefalexin | 2 | >128 | 1 to >128 | 70.6 | NA | 29.4 |
| | Ciprofloxacin | 16 | >16 | ≤0.06 to >16 | 41.2 | 0 | 58.8 |
| | Fosfomycin | 128 | 128 | 128 to >512 | 0 | 94.1 | 5.9 |
| | Nitrofurantoin | 16 | 64 | 4–64 | 88.2 | 11.8 | 0 |
| | SXT | 0.5 | >8 | ≤0.12 to >8 | 52.9 | NA | 47.1 |
| Cefalexin-non-susceptible (131) | AMC | 16 | >32 | ≤0.06 to >32 | 47.3 | 26.0 | 26.7 |
| | Cefalexin | >128 | >128 | 32 to >128 | 0 | NA | 100 |
| | Ciprofloxacin | >16 | >16 | ≤0.06 to >16 | 38.2 | 0 | 61.8 |
| | Fosfomycin | 2 | 4 | ≤1 to >512 | 96.2 | 3.0 | 0.8 |
| | Nitrofurantoin | 16 | 32 | ≤1–256 | 90.8 | 5.4 | 3.8 |
| | SXT | >8 | >8 | ≤0.12 to >8 | 47.3 | NA | 52.7 |
| AMC-non-susceptible (293) | AMC | 16 | 32 | 16 to >32 | 0 | 73.0 | 27.0 |
| | Cefalexin | 8 | >128 | 1 to >128 | 76.5 | NA | 23.5 |
| | Ciprofloxacin | ≤0.06 | >16 | ≤0.06 to >16 | 64.5 | 0 | 35.5 |
| | Fosfomycin | ≤1 | 4 | ≤1–128 | 99.3 | 0.7 | 0 |
| | Nitrofurantoin | 16 | 32 | ≤1–256 | 94.9 | 3.4 | 1.7 |
| | SXT | 0.5 | >8 | ≤0.12 to >8 | 54.3 | NA | 45.7 |
| Ciprofloxacin-non-susceptible (346) | AMC | 8 | 16 | 1 to >32 | 69.9 | 24.3 | 5.8 |
| | Cefalexin | 4 | >128 | 1 to >128 | 76.6 | NA | 23.4 |
| | Ciprofloxacin | >16 | >16 | 2 to >16 | 0 | 0.6 | 99.4 |
| | Fosfomycin | 2 | 4 | ≤1 to >512 | 97.1 | 2.6 | 0.3 |
| | Nitrofurantoin | 16 | 32 | ≤1–256 | 91.6 | 5.5 | 2.9 |
| | SXT | >8 | >8 | ≤0.12 to >8 | 41.3 | NA | 58.7 |
| MDR ^b (205) | AMC | 16 | 32 | 1 to >32 | 40.5 | 39.0 | 20.5 |
| | Cefalexin | 32 | >128 | 1 to >128 | 47.8 | NA | 52.2 |
| | Ciprofloxacin | >16 | >16 | ≤0.06 to >16 | 20.5 | 0 | 79.5 |
| | Fosfomycin | 2 | 4 | ≤1 to >512 | 95.1 | 4.4 | 0.5 |
| | Nitrofurantoin | 16 | 64 | ≤1–256 | 86.8 | 8.8 | 4.4 |
| | SXT | >8 | >8 | ≤0.12 to >8 | 22.4 | NA | 77.6 |
| ESBL-positive (77) | AMC | 8 | 16 | 4 to >32 | 59.7 | 32.5 | 7.8 |
| | Cefalexin | >128 | >128 | 16 to >128 | 1.3 | NA | 98.7 |
| | Ciprofloxacin | >16 | >16 | ≤0.06 to >16 | 18.2 | 0 | 81.8 |
| | Fosfomycin | 2 | 4 | ≤1–128 | 96.1 | 3.9 | 0 |
| | Nitrofurantoin | 16 | 64 | ≤1–256 | 87.0 | 9.1 | 3.9 |
| | SXT | >8 | >8 | ≤0.12 to >8 | 33.8 | NA | 66.2 |
| AmpC-positive (28) | AMC | 32 | >32 | 1 to >32 | 25.0 | 21.4 | 53.6 |
| | Cefalexin | 64 | >128 | ≤0.5 to >128 | 35.7 | NA | 64.3 |
| | Ciprofloxacin | 0.12 | >16 | ≤0.06 to >16 | 57.1 | 0 | 42.9 |
| | Fosfomycin | 2 | 8 | ≤1 to >512 | 96.4 | 0 | 3.6 |
| | Nitrofurantoin | 16 | 32 | 4–256 | 96.4 | 0 | 3.6 |
| | SXT | 0.25 | >8 | ≤0.12 to >8 | 64.3 | NA | 35.7 |

MIC, minimum inhibitory concentration; MIC_{50/90}, MIC required to inhibit 50% and 90% of the isolates, respectively; CLSI, Clinical and Laboratory Standards Institute; S, susceptible; I, intermediate; R, resistant; AMC, amoxicillin/clavulanic acid; SXT, trimethoprim/sulfamethoxazole; MDR, multidrug-resistant; ESBL, extended-spectrum β-lactamase.

^a NA, not available (intermediate MIC interpretative breakpoints are not published by the CLSI for this antimicrobial agent) [12].

^b A MDR isolate was defined by a phenotype that was non-susceptible to agents from three or more antimicrobial classes [30]. See footnote ^b under Table 3 for a detailed description of the most common MDR isolates identified in this study.

Table 5
Sex and age group analysis of *Escherichia coli* isolates cultured from urine specimens of outpatients in Canada, 2007–2016.

| Sex (n)/age group (n) | % of sex total | MDR | | ESBL | | AmpC | |
|-----------------------|----------------|-----|-----------------------------|------|-----------------------------|------|-----------------------------|
| | | n | % of sex/age group isolates | n | % of sex/age group isolates | n | % of sex/age group isolates |
| Female | | | | | | | |
| 0–17 years (297) | 17.7 | 17 | 5.7 | 8 | 2.7 | 2 | 0.7 |
| 18–45 years (539) | 32.2 | 42 | 7.8 | 17 | 3.2 | 6 | 1.1 |
| 46–64 years (301) | 18.0 | 30 | 10.0 | 12 | 4.0 | 6 | 2.0 |
| ≥65 years (537) | 32.1 | 69 | 12.8 | 21 | 3.9 | 7 | 1.3 |
| Total (1674) | 100 | 158 | 9.4 | 58 | 3.5 | 21 | 1.3 |
| Male | | | | | | | |
| 0–17 years (76) | 21.1 | 3 | 3.9 | 0 | 0 | 1 | 1.3 |
| 18–45 years (42) | 11.6 | 8 | 19.0 | 3 | 7.1 | 0 | 0 |
| 46–64 years (95) | 26.3 | 12 | 12.6 | 4 | 4.2 | 1 | 1.1 |
| ≥65 years (148) | 41.0 | 24 | 16.2 | 12 | 8.1 | 5 | 3.4 |
| Total (361) | 100 | 47 | 13.0 | 19 | 5.3 | 7 | 1.9 |
| Grand total | | 205 | | 77 | | 28 | |

MDR, multidrug-resistant; ESBL, extended-spectrum β -lactamase.

Acknowledgments

The authors would like to thank the investigators and staff at the contributing clinical microbiology laboratories for their participation in the CANWARD surveillance study from 2007–2016.

Funding

The CANWARD surveillance study was supported in part by the Health Sciences Centre (Winnipeg, Manitoba, Canada), the University of Manitoba (Winnipeg, Manitoba, Canada), the Public Health Agency of Canada – National Microbiology Laboratory (Winnipeg, Manitoba, Canada) and Paladin Labs Inc. (St-Laurent, Québec, Canada).

Competing interests

None declared.

Ethical approval

The CANWARD study receives annual approval by the University of Manitoba Research Ethics Board [H2009:059].

References

- [1] Bader MS, Loeb M, Brooks AA. An update on the management of urinary tract infections in the era of antimicrobial resistance. *Postgrad Med* 2017;129:242–58.
- [2] Nicolle LE. Epidemiology of urinary tract infections. *Clin Microbiol Newsl* 2001;24:135–40.
- [3] Foxman B. Urinary tract infection syndromes: occurrence, recurrence, bacteriology, risk factors, and disease burden. *Infect Dis Clin North Am* 2014;28:1–13.
- [4] Echols RM, Tosiello RL, Haverstock Tice AD. Demographic, clinical, and treatment parameters influencing the outcome of acute cystitis. *Clin Infect Dis* 1999;29:113–19.
- [5] Gupta K, Hooton TM, Naber KG, Wullt B, Colgan R, Miller LG, et al. International clinical practice guidelines for the treatment of uncomplicated acute cystitis and pyelonephritis in women: a 2010 update by the Infectious Diseases Society of America and the European Society for Microbiology and Infectious Diseases. *Clin Infect Dis* 2011;52:e103–20.
- [6] Lob SH, Nicolle LE, Hoban DJ, Kazmierczak KM, Badal RE, Sahn DF. Susceptibility patterns and ESBL rates of *Escherichia coli* from urinary tract infections in Canada and the United States, SMART 2010–2014. *Diagn Microbiol Infect Dis* 2016;85:459–65.
- [7] Walker E, Lyman A, Gupta K, Mahoney MV, Snyder GM, Hirsch EB. Clinical management of an increasing threat: outpatient urinary tract infections due to multidrug-resistant uropathogens. *Clin Infect Dis* 2016;63:960–5.
- [8] Hooton TM, Roberts PL, Cox ME, Stapleton AE. Voided midstream urine culture and acute cystitis in premenopausal women. *N Engl J Med* 2013;369:1883–91.
- [9] Yarborough ML. Impact of reflex algorithms on urine culture utilization. *Clin Microbiol Newsl* 2018;40:19–24.
- [10] Nicolle LE, Bradley S, Colgan R, Rice JC, Schaeffer A, Hooton TM. Infectious Diseases Society of America guidelines for the diagnosis and treatment of asymptomatic bacteriuria in adults. *Clin Infect Dis* 2005;40:643–654.
- [11] Association of Medical Microbiology and Infectious Disease Canada. Choosing Wisely Canada – Medical Microbiology. Canada: Choosing Wisely; 2017.
- [12] Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. 28th ed. Wayne, PA: CLSI; 2018. CLSI supplement M100.
- [13] Clinical and Laboratory Standards Institute. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. 11th ed. Wayne, PA: CLSI; 2018. CLSI standard M07.
- [14] van den Bijllaardt W, Schijffelen MJ, Bosboom RW, Stuart JC, Dierderen B, Kampinga G, et al. Susceptibility of ESBL *Escherichia coli* and *Klebsiella pneumoniae* to fosfomycin in the Netherlands and comparison of several testing methods including Etest, MIC test strip, Vitek2, Phoenix and disk diffusion. *J Antimicrob Chemother* 2018;73:2380–7.
- [15] Camarlinghi G, Parisio EM, Antonelli A, Nardone M, Coppi M, Giani T, et al. Discrepancies in fosfomycin susceptibility testing of KPC-producing *Klebsiella pneumoniae* with various commercial methods. *Diagn Microbiol Infect Dis* 2019;93:74–6.
- [16] Sanchez GV, Master RN, Karlowsky JA, Bordon JM. In vitro antimicrobial resistance of urinary *Escherichia coli* isolates among U.S. outpatients from 2000 to 2010. *Antimicrob Agents Chemother* 2012;56:2181–3.
- [17] Sanchez GV, Baird AMG, Karlowsky JA, Master RN, Bordon JM. Nitrofurantoin retains antimicrobial activity against multidrug-resistant urinary *Escherichia coli* from US outpatients. *J Antimicrob Chemother* 2014;69:3259–62.
- [18] Sanchez GV, Babiker A, Master RN, Luu T, Mathur A, Bordon JM. Antibiotic resistance among urinary isolates from female outpatients in the United States in 2003 and 2012. *Antimicrob Agents Chemother* 2016;60:2680–3.
- [19] US Centers for Disease Control and Prevention (CDC). Biggest threats and data. Atlanta, GA: CDC; 2018 https://www.cdc.gov/drugresistance/biggest_threats.html.
- [20] Sahn DF, Brown NP, Yee YC, Evangelista AT. Stratified analysis of multidrug-resistant *Escherichia coli* in US health care institutions. *Postgrad Med* 2008;120:53–9.
- [21] Rossignol L, Vaux S, Maugat S, Blake A, Barlier R, Heym B, et al. Incidence of urinary tract infections and antibiotic resistance in the outpatient setting: a cross-sectional study. *Infection* 2017;45:33–40.
- [22] Karlowsky JA, Denisuk AJ, Lagacé-Wiens PR, Adam HJ, Baxter MR, Hoban DJ, et al. In vitro activity of fosfomycin against *Escherichia coli* isolated from patients with urinary tract infections in Canada as part of the CANWARD surveillance study. *Antimicrob Agents Chemother* 2014;58:1252–6.
- [23] Karlowsky JA, Lagacé-Wiens PR, Simner PJ, DeCorby MR, Adam HJ, Walkty A, et al. Antimicrobial resistance in urinary tract pathogens in Canada from 2007 to 2009: CANWARD surveillance study. *Antimicrob Agents Chemother* 2011;55:3169–75.
- [24] Zhanel GG, Hisanaga TL, Laing NM, DeCorby MR, Nichol KA, Weshnowski B, et al. Antibiotic resistance in *Escherichia coli* outpatient urinary isolates: final results from the North American Urinary Tract Infection Collaborative Alliance (NAUTICA). *Int J Antimicrob Agents* 2006;27:468–75.
- [25] Kresken M, Körber-Irrgang B, Biedenbach DJ, Batista N, Besard V, Cantón R, et al. Comparative in vitro activity of oral antimicrobial agents against Enterobacteriaceae from patients with community-acquired urinary tract infections in three European countries. *Clin Microbiol Infect* 2016;22:63 e1–5.
- [26] Lagacé-Wiens PRS, Simner PJ, Forward KR, Tailor F, Adam HJ, DeCorby M, et al. Analysis of 3789 in- and outpatient *Escherichia coli* isolates from across Canada—results of the CANWARD 2007–2009 study. *Diagn Microbiol Infect Dis* 2011;69:314–19.

- [27] Zhanel GG, Adam HJ, Baxter MR, Fuller J, Nichol K, Denisuk A, et al. Antimicrobial susceptibility of 22,746 pathogens from Canadian hospitals; results of the CANWARD 2007–2011 study. *J Antimicrob Chemother* 2013;68(Suppl 1):7–22.
- [28] Clinical and Laboratory Standards Institute. Abbreviated identification of bacteria and yeast; approved guideline. second edition. Wayne, PA: CLSI; 2008. CLSI document M35-A2.
- [29] Simner PJ, Zhanel GG, Pitout J, Taylor F, McCracken M, Mulvey MR, et al. Prevalence and characterization of extended-spectrum β -lactamase- and AmpC β -lactamase-producing *Escherichia coli*: results of the CANWARD 2007–2009 study. *Diagn Microbiol Infect Dis* 2011;69:326–34.
- [30] Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 2012;18:268–81.
- [31] Weiner LM, Webb AK, Limbago B, Dudeck MA, Patel J, Kallen AJ, et al. Antimicrobial-resistant pathogens associated with healthcare-associated infections: summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2011–2014. *Infect Control Hosp Epidemiol* 2016;37:1288–301.
- [32] Puttagunta S, Gupta V, Murray J, Dunne M. Prevalence of extended-spectrum β -lactamase producing and quinolone non-susceptible Enterobacteriaceae in inpatient and outpatient settings in the USA from 2011–2017. IDWeek 2017, VA: Infectious Diseases Society of America; 2017. 4–8 October 2017; San Diego, CA. Arlington[abstract 400].
- [33] Kronenberg A, Koenig S, Droz S, Muhlemann K. Active surveillance of antibiotic resistance prevalence in urinary tract and skin infections in the outpatient setting. *Clin Microbiol Infect* 2011;17:1845–51.