



## Inflamm-aging microRNAs may integrate signals from food and gut microbiota by modulating common signalling pathways

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### ABSTRACT

Human gut microbiota, which comprises an extremely diverse and complex community of microorganisms inhabiting the intestinal tract, may be associated with inflammation and age-related chronic health conditions. However, the mechanism underlying this association is only recently beginning to emerge. Transfer and modulation of gene expression by diet-derived microRNAs (miRs) in mammals might be involved in this communication. Through a bioinformatics approach, using on line tools and repositories, we searched for evidences that food-containing miRs, actually involved in the modulation of the inflammatory process, (inflamma-miRs), may contribute to mediate the anti-inflammatory effects exerted by some foods through the modulation of aging-related pathways and gut microbiota composition in a bidirectional communication. Supported by a “Pubmed” search and our previous research, a trio of experimentally validated inflamma-miRs were considered: miR-155, miR-146a and miR-21. Our *in silico* study supports the hypothesis that these inflamma-miRs could modulate some pathways, such as lysine degradation and lengthening of fatty acids which are involved in the modulation of microbiota composition, i.e. prevotella, ruminococcus and oscillibacter and *vice versa*. Food homologues to human miR-21, miR-155 and miR-146a were found in cow fat, cow milk, and eggs suggesting that they may be able of targeting, and probably exacerbating, inflammation related pathways. If these data will be experimentally validated, they will further support the relevance of a nutraceutical approach for a healthy aging.

### 1. Introduction

Although research on nutrition has focused on the interactions between nutrients and organism, it is becoming increasingly meaningful to study the effects of nutrient on host gut microbiota composition. Human organism hosts trillions microbial cells in a symbiotic relationship and, in the normal healthy state, they benefit from each other. Over 1000 different species of microorganism are making up the human microbiota. Ninety % of microbiota is concentrated in the gut, especially intestine. Emerging evidences indicated gut microbiota as an important component of human health preservation. The understanding the interrelationship among nutrient-host-microbiota may lead to the discovery of new mechanisms of disease onset and progression. Recent studies are indicating the microbiota as a principal key of healthy phenotype during aging (Franceschi et al., 2017; Claesson et al., 2012;

Forslund et al., 2015). A new intriguing hypothesis of aging is that bacteria influence the degree of systemic inflammation through chronic, low grade endotoxemia, thus promoting and sustaining inflamm-aging (Zhang et al., 2013). Growing evidence suggests chronic low-grade inflammation (LGI) as one of the most relevant possible mechanism underlying the aging process (Custodero et al., 2018). A major negative consequence of aging is immuno-senescence, which can be defined as a decline in the functionality of the immune system (Gruber et al., 2007), causing a chronic low-grade inflammatory status in the gut. Immuno-senescence can therefore cause unfavourable changes in the composition and structure of the gut microbiota in older people (Neish, 2009).

Gut microbiota can modulate the host gene expression, primarily the innate immune response (Tomkovich and Jobin, 2015). Recent evidences suggest that communication between the gut microbiota and

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host may in part occur via microRNAs miRs (Williams et al., 2017; Dalmasso et al., 2011). MiRs are a class of short (18–24 nt) regulatory RNAs that are widely evolutionary conserved among many species. These single-stranded, non-coding molecules mediate gene regulation at different levels. The best characterized function of miRs is the post-transcriptional repression of the targets mRNA by promoting cleavage or inhibiting its translation. In this way, miRs negatively regulate target genes expression, modulating all biological functions, including cell development, apoptosis, proliferation, differentiation and regulate inflammatory pathways (Cătană et al., 2015; Olivieri et al., 2013).

Several evidences showed that nutrients may modulate endogenous miRs (Carotenuto et al., 2016). In addition to endogenous miRs, recent fascinating results demonstrated that food-derived miRs enter the mammals' circulatory system and reach their target thus modulating specific mammalian protein expression (Wang et al., 2012; Sherman et al., 2015). This as an intriguing thesis that may pave the way for novel therapeutic approaches discovery. Recent studies confirmed this hypothesis demonstrating that miRs not only are synthesized endogenously, but also might be obtained from dietary sources (Philip et al., 2015; Baier et al., 2014). We hypothesized that ingested biological matter contributes directly to the miRs arsenal of body compartments or that the diet-derived exogenous miRs (referred as "xenomiRs") can affect total miRs profiles as part of a circulating miRs homeostasis that is altered in many diseases (Witwer, 2012). Epigenetic crosstalk between different kingdoms, has been extensively studied in the interaction between plants and bacteria, while evidences in humans are emerging only recently (Williams et al., 2017; Liu et al., 2016; Hua et al., 2018; Qin and Wade, 2017). Increasing evidence suggests the existence of a miRs-based crosstalk between microbiota, the host, and food (Witwer, 2012; Liu et al., 2016; Philip et al., 2015).

Beyond opposing arguments regarding the cross-kingdom regulation of gene expression by plant miRs, concern on how these molecules can access the gastro-intestinal (GI) tract, enter the circulation and transport to cell have also been raised. Several endogenously originating miRs intercellular carriers have been identified, including microvesicle (MV), which are membrane-derived vesicles released from various cell types (Valadi et al., 2007; Zerneck et al., 2009; Zhang et al., 2015; Turchinovich et al., 2012). Encapsulation of miRs in exosomes and exosome-like particles confers protection against RNA degradation and creates a pathway for intestinal and vascular endothelial transport by endocytosis, as well as delivery to peripheral tissues. Exosome-like nanoparticles present in edible fruits and vegetables can be phagocytosed by intestinal macrophages and stem cells (Mu et al., 2014). Thus, considering the recent assumptions and evidences that exogenous miRs might be sufficiently stable to pass through the GI tract and enter circulation without losing functionality, we decided to study whether food-containing miRs, especially those that were identified in serum (exosome), can carry signals to host modulating immune system. For this purpose, we used a bioinformatics approach (*in silico*) that, through web-based tools and supplemental data available in literature, can predict the putative microRNAs involved in health microbiota. In this way, we were able to hypothesize new regulatory mechanisms associated to microbiota driven immune dysregulation and its implication with aging. Indeed, *in silico* biology which refers to computational models and simulations applied to complex biological phenomena is a potent tool to obtain quantitative insights into regulatory networks and processes. Due to the vast amounts of data generated by molecular analyses, computational biology is increasingly exploited to filter big amount of data. *In silico* applied to vast amounts of biological data uses sophisticated algorithms to advance scientific understanding. The retrieved information can then be experimentally validated or serve as a guide for future investigations. We consulted different databases for microRNAs pathway prediction, microbiota integrated pathways, clusters % match with other species (Dietary microRNA Database) and to filter common pathways (SID1.0). The aim of our *in silico* approach is to study the possible miRs-mediated interactions

between the healthy microbiota and host and to explore the impact of nutrition on maintaining the health status, with a specific focus on the inflamm-aging process.

## 2. Materials and methods

### 2.1. Data collection

We used PubMed free resource (<https://www.ncbi.nlm.nih.gov/pubmed/>) to collect papers from literature related to food and microbiota impacting to inflamm-aging. This data search helped us to collect miRs information contained in feces relevant for aging and inflammation. For this purpose, the combination of the following terms has been used in the search box: microbiota health interactions, immunity, inflamm-aging, microRNA, food and nutrition. We focused our attention on miRs able of shaping microbiota and of modulating inflamm-aging processes. These collected miRs were all experimentally validated in the papers we have selected.

### 2.2. Bioinformatics in silico analysis

We exploited different bioinformatics repositories and tools to analyze microbiota (Ruminococcus; Prevotella; Oscillibacter species) associated pathways, microRNAs modulated pathways, dietary microRNAs and common microbiota/microRNAs pathways. These tools are reported below.

DIANA tool (<http://snf-515788.vm.okeanos.grnet.gr/>) has been used for the prediction of the pathways modulated by microRNAs. Since DIANA can utilize experimentally validated miRNA interactions derived from DIANA-TarBase, we retrieved the KEGG Pathway IDs (strings) modulated by each miRNAs (Annex 1 in Supplementary data) and used them for SID1.0 analysis. MicroRNA nomenclatures used for the analysis were the following: hsa-miR-21-3p; hsa-miR-21-5p; hsa-miR-155-3p; hsa-miR-155-5p; hsa-miR-146a-3p; hsa-miR-146a-5p; hsa-miR-146b-3p; hsa-miR-146b-5p.

PATRIC (<https://patricbrc.org/>) tool has been used to find the metabolic pathways related to the genomes of Ruminococcus, Prevotella and Oscillibacter species. PATRIC puts together data from sources, data types, molecular entities and organisms. Data are collected from public repositories, for instance NCBI, the United States Department of Agriculture, the Centers for Disease Control (CDC) and other NIAID-funded projects, such as the System Biology for Infectious Disease Centers, as well as from studies published by researchers. The search tool was used to insert the name of each species and the KEGG IDs (Strings) has been used for SID1.0 analysis (Annex 2 in Supplementary data).

DMD database (Dietary microRNA Database: <http://sbbi-panda.unl.edu:5000/dmd/browse>) has been used to retrieve food containing microRNAs (miR-155, miR-146a and miR-21) with 100% clusters percentage match with human.

SID1.0 tool is a simple string identifier that has been used to filter the common microbiota/microRNAs pathways (strings of KEGG Pathway ID) obtained from PATRIC and DIANA tools retrieving common strings (Annex 3 in Supplementary data). All the information regarding the tools and repositories we used are reported in the webpage of the tools/repositories quoted. In the flow chart in Fig. 1 the followed road map is reported.

## 3. Results

### 3.1. MicroRNAs involved in the crosstalk between microbiota and host

From our data search we observed that feces contain miRs relevant for aging and inflammation processes. Based on our previous experiences we selected from the literature three miRs, namely miR-155, miR-21 and miR-146a as markers of inflamm-aging sensing. From the data

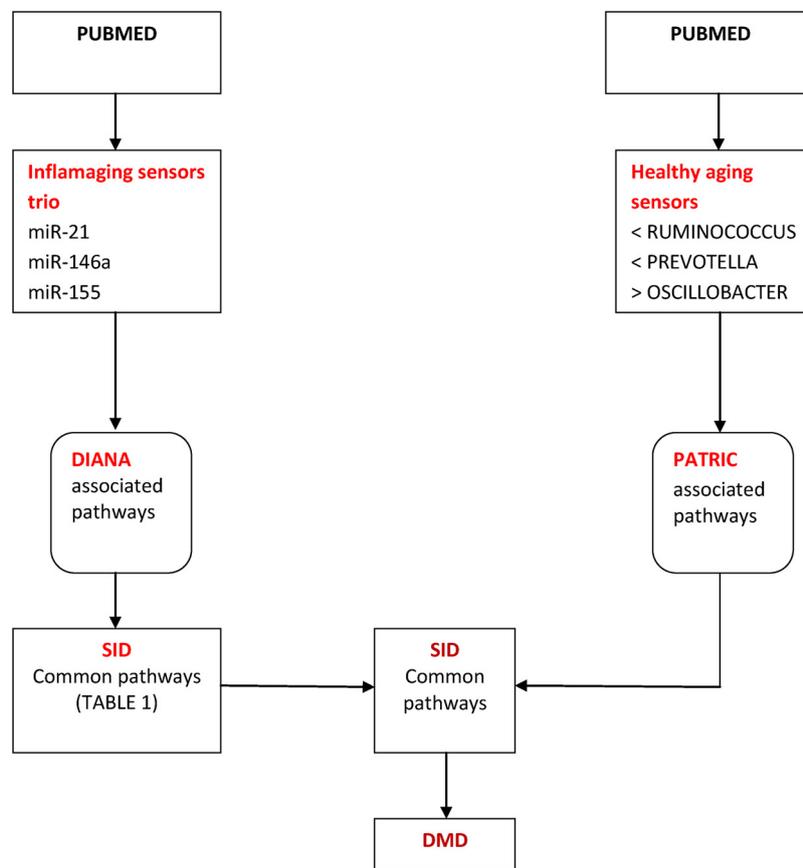


Fig. 1. Flowchart of visual representation of the sequence of steps applied to perform our *in silico* study.

Table 1

miR-21, miR-155 and miR-146a common pathways analyzed with SID1.0 (the pathways have been retrieved with DIANA tool). Common pathways were found only with the combination of 2 miRs out of three, no common pathways between all the trio-miRs have been found.

KEGG Pathway ID	Pathway name	Common microRNAs
hsa05142	Chagas disease (American trypanosomiasis)	miR-146a; miR-155
hsa05321	Inflammatory bowel disease (IBD)	miR-146a; miR-155
hsa04064	NF-kappa B signaling pathway	miR-146a; miR-155
hsa05161	Hepatitis B	miR-146a; miR-155
hsa05203	Viral carcinogenesis	miR-146a; miR-21
hsa04012	ErbB signaling pathway	miR-146a; miR-21
hsa04520	Adherens junction	miR-155; miR-21
hsa05223	Non-small cell lung cancer	miR-155; miR-21
hsa05219	Bladder cancer	miR-155; miR-21
hsa05212	Pancreatic cancer	miR-155; miR-21
hsa04110	Cell cycle	miR-146a; miR-21
hsa05210	Colorectal cancer	miR-155; miR-21
hsa00310	Lysine degradation	miR-146a; miR-21
hsa05220	Chronic myeloid leukemia	miR-155; miR-21
hsa00062	Fatty acid elongation	miR-155; miR-21

collected, these three candidate miRs were found to be able of modulating inflamm-aging processes (Olivieri et al., 2013) and of shaping microbiota. The pathways associated to each miR of the trio were retrieved using DIANA tool. Subsequently, the search for common pathways was carried out inquiring SID1.0. This analysis demonstrated that common pathways were found with two by two miRs combination only. As indicated in the Table 1, many of the pathways regulated by these miRs are involved in immune system processes indicating that these processes are mainly involved in the crosstalk between microbiota and host.

### 3.2. Pathways associated to healthy microbiota, miR-21, miR-155 and miR-146a

Literature search evidenced that gut microbiota composition correlates with diet and health in the elderly and it is mainly associated either to a decreased amount of Ruminococcus/Prevotell and to an increased amount of the Oscillibacter as reported by Claesson et al. Thus, these three enterobacters have been taken as indicative of healthy aging.

By inquiring PATRIC tool we were able to find the pathways associated to healthy aging bacteria (Annex 2 in Supplementary data). We combined with that of Table 1 (pathways associated to each of the trio miRs) and inquiring SID1.0 analysis we, thus, retrieved the common pathways to bacteria and miRs as demonstrated in Table 2. As showed from the data reported in Table 2 common pathways to all of them were not found. We found the common pathways with the combination of 5 and 4 out of the six items (Annex 3 in Supplementary data).

Our results indicate that the miRs we selected, and the bacteria considered may modulate: Amino Acid Metabolism, Lipid Metabolism, Cofactors and Vitamins Metabolism, Glycan Biosynthesis and Metabolism.

### 3.3. Food derived microRNA match with human microRNAs

Based on the fact that the analysis presented in Table 2 demonstrated that miR-21, miR-155 and miR-146a can probably modulate gut associated metabolism and could be involved in inflamm-aging process and on the observation that diet is emerging as the most critical determinant in human health and healthy aging, we inquired into the web platform DMD to retrieve information about foods containing such miRs. The results are reported in Table 3. As indicated in Table 3 cow fat/milk, chicken (and chicken egg), Atlantic salmon and pig possess

**Table 2**

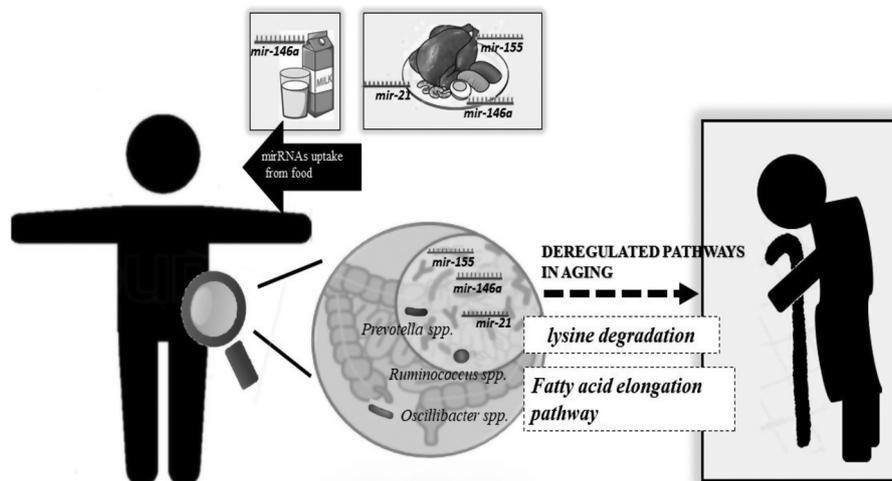
MiR-21, miR-155, miR-146a, Ruminococcus, Prevotell and Oscillibacter common pathways analyzed with SID1.0 (the pathways have been retrieved with Diana tool, mirpath v.3 for miRS and with PATRIC tool for microbiota). Common pathways to all of them were not found. We found the common pathways with the combination of 5 and 4 of them out of 6 elements (trio-miRs and trio-bacteria).

KEGG Pathway ID	Pathway name/Pathway class	Common microRNAs/microbiota
hsa00310	Lysine degradation/ Amino Acid Metabolism	miR-21; miR-146a/ Ruminococcus; Prevotell; Oscillibacter
hsa00062	Fatty acid elongation in mitochondria/ Lipid Metabolism	miR-21; miR-155/ Ruminococcus; Prevotell; Oscillibacter
hsa00670	One carbon pool by folate/ Metabolism of Cofactors and Vitamins	miR-146a/ Ruminococcus; Prevotell; Oscillibacter
hsa00601	Glycosphingolipid biosynthesis - lacto and neolacto series/ Glycan Biosynthesis and Metabolism	miR-146a/ Ruminococcus; Prevotell; Oscillibacter
hsa00290	Valine, leucine and isoleucine biosynthesis/ Amino Acid Metabolism	miR-21/ Ruminococcus; Prevotell; Oscillibacter
hsa00071	Fatty acid metabolism/ Lipid Metabolism	miR-21/ Ruminococcus; Prevotell; Oscillibacter
hsa00061	Fatty acid biosynthesis/ Lipid Metabolism	miR-21/ Ruminococcus; Prevotell; Oscillibacter
hsa00100	Steroid biosynthesis/ Lipid Metabolism	miR-155/ Ruminococcus; Prevotell; Oscillibacter

**Table 3**

DMD has been used to find the percentage match between food and human miR-21, miR-155 and miR-146a.

	miR-21	miR-155	miR-146a	% Match (Dietary species in DMD)
<b>Cow microRNA fat (bta-)</b>	bta-miR-21-5p	bta-miR-155	bta-miR-146a	95.65% gga-miR-155 <b>100 % hsa-miR-155-5p</b> 100 % ssa-miR-155-5p
<b>Cow microRNA milk (bta-)</b>	bta-miR-21-5p bta-miR-21-3p	bta-miR-155	bta-miR-146a	95.65% gga-miR-155 <b>100 % hsa-miR-155-5p</b> 100 % ssa-miR-155-5p
<b>Orange microRNA fruit (csi-)</b>	-	-	-	-
<b>Chicken egg MicroRNA (gga-)</b>	gga-miR-21-5p	-	gga-miR-146a-5p	<b>100 % hsa-miR-21-5p</b> 100% ssc-miR-21 <b>100%hsa-miR-146a-5p</b> 100% ssc-miR-146a-5p
<b>Chicken MicroRNA (gga-)</b>	gga-miR-21-3p	gga-miR-155	gga-miR-146a-3p	95.45% bta-miR-155 <b>95.45%hsa-miR-155-p</b> 95.45% ssa-miR-155-5p
<b>Soybean MicroRNA (gma-)</b>	-	-	-	-
<b>Human MicroRNA, breastmilk (hsa-)</b>	hsa-miR-21-5p hsa-miR-21-3p	hsa-miR-155-5p	hsa-miR-146a-5p	100 % gga-miR-21-5p 100 % ssc-miR-21 100 % bta-miR-155 95.65% gga-miR-155 100 % ssa-miR-155-5p 100 % gga-miR-146a-5p 100 % ssc-miR-146a-5p
<b>Apple MicroRNA (mdm-)</b>	-	hsa-miR-155-3p	-	-
<b>Banana MicroRNA (msp-)</b>	msp-miR-21 msp-miR-21*	msp-miR-155 msp-miR-155*	msp-miR-146 msp-miR-146*	- -
<b>Rice MicroRNA (osa-)</b>	-	-	-	-
<b>Tomato MicroRNA (sly-)</b>	-	-	-	-
<b>Atlantic salmon MicroRNA (ssa-)</b>	ssa-miR-21a-5p ssa-miR-21a-3p ssa-miR-21a-2-3p ssa-miR-21b-5p ssa-miR-21b-3p	ssa-miR-155-5p	ssa-miR-146a-5p	100% bta-miR-155    95.65% gga-miR-155 <b>100% hsa-miR-155-5p</b>
<b>Pig MicroRNA (ssc-)</b>	ssc-miR-21	ssc-miR-155-3p ssc-miR-155-5p	ssc-miR-146a-3p ssc-miR-146a-3-3p ssa-miR-146b-5p ssa-miR-146b-3p ssa-miR-146d-5p ssa-miR-146d-3p ssa-miR-146d-2-3p	<b>100% gga-miR-21-5p</b> <b>100% hsa-miR-21-5p</b> 95.24% hsa-miR-146b-5p
<b>Wheat MicroRNA (tae-)</b>	-	-	-	-
<b>Grape MicroRNA (vvi-)</b>	-	-	-	-
<b>Corn MicroRNA (zma-)</b>	-	-	-	-



**Fig. 2.** Graphical representation of the results indicating an interaction among foods, host microbiota and aging through the common pathways lysine degradation and fatty acid elongation.

one or two inflamm-aging miRNAs with a 100% match with the same human microRNAs. All the other aliments indicated in the [Table 3](#) do not have these microRNAs or do not matches 100% with humans. It must be noted that not many foods have been tested yet for miRNAs libraries. It's interesting to note that human breast milk used only by neonates, has 100% matches for the 3 inflamm-aging miRNAs from animal foods. The fact that breast milk is limited to weaning probably indicates that this aliment is not a suitable food for adults, and this represents an interesting observation.

### 3.4. Gut microbiota, inflamm-aging miRNAs, diet and signaling pathways

Lysine degradation pathway and fatty acid elongation pathways were the two most representative pathways (five out of six elements in common) as demonstrated in [Table 2](#). Indeed, these pathways are found common to miR-21; miR-146a/ Ruminococcus; Prevotell; Oscillibacter. In [Fig. 2](#) are graphically represented these converging routes, taking together the results from [Tables 1–3](#).

## 4. Discussion

We demonstrated here, through a bioinformatics *in silico* study, that a cross-talk among i) gut microbiota, ii) human inflamm-aging miRNAs, and iii) foods might occur. This suggests that diet induces epigenetic modifications and these modifications work together in various combinations with gut microbiota and host, targeting signalling pathways relevant for contrasting or exacerbating inflammation associated senescence. The debate about gut microbiota miRNAs existence and their capacity to modulate human mRNA transcripts is a hot topic and a controversial issue. However, epigenetic crosstalk between different kingdoms, especially regarding exosome or vesicle like particles cross-talk between kingdoms is becoming more and more evident and it can be relevant for the aging phenotype outcome ([Williams et al., 2017](#)). In addition, recent studies suggest that miRNAs not only are synthesized endogenously, but also might be obtained from dietary sources ([Quintanilha et al., 2017](#)). We previously defined a set of miRNAs, namely: miR-146, miR-155, and miR-21, which play a central role in the interplay among DNA damage response, cell senescence and inflammation and yield a well-recognized role in aging and development of aging related diseases (ARDs) in humans ([Olivieri et al., 2015](#); [Carotenuto et al., 2016](#)). We found here that this trio-miRNAs shows inflammation related pathways in common ([Table 1](#)). Noteworthy, the analysis presented in [Table 2](#) demonstrated that miR-21, miR-155 and miR-146a share common molecular pathways with some of gut microbiota demonstrating that this trio-miRNAs can actually modulate the gut associated

metabolism. Among the common molecular pathways, we found that lysine degradation and fatty acid elongation in mitochondria are the most represented being common to 4 out of the five items considered, namely: miR-21; miR-146a; Ruminococcus; Prevotell; Oscillibacter (for lysine degradation pathway) and miR-21; miR-155; Ruminococcus; Prevotell; Oscillibacter (for fatty acid elongation in mitochondria). [Vital et al. \(2014\)](#) demonstrated that acetyl-coenzyme A (CoA) is the microbiota most prevalent pathway (mean, 79.7% of all pathways), followed by the lysine pathway (mean, 11.2%). On the other hand, acetyl-CoA is a lysine degradation pathway product. Indeed, in mammals, lysine is metabolized to acetyl-CoA; an acetyl-CoA rise induces the increase of the overall acetylation of cytoplasmic proteins, as well as the impairment of autophagy ([Mariño et al., 2014](#)). Acetyl-CoA is a central molecule connecting the catabolism of glucose, fatty acids, and amino acids, thus it is a key metabolite at the intersection of catabolic and anabolic metabolism being a major acetyl donor in cells.

It is worthwhile to note that the mammalian degradation of lysine proceeds via two distinct routes, the saccharopine and the pipecolic acid route. In mammalian brain an age-dependent lysine degradative pathway occurs, and the pipecolate pathway emerged as the main catabolic pathways, whereas the saccharopine pathway prevails in developing brain ([Hallen et al., 2013](#)). Indeed, lysine degradation pathway dysregulation has been associated to neurodegeneration in elderly. In addition, lysine plays several others important roles in humans, like proteinogenesis, crosslinking of collagen polypeptides, uptake of essential mineral nutrients, production of carnitine, which is key element in fatty acid metabolism and elongation. The growing fatty acid chain is elongated by the sequential addition of two-carbon units derived from acetyl-CoA. Genes of fatty acid elongation pathway were decreased in intestinal microbiome of patients suffering from neovascular age-related macular degeneration ([Zinkernagel et al., 2017](#)). Indeed, it has been suggested that fatty acid chain elongation is decreased with age ([Jimenez et al., 1997](#)).

Apart from the above discussed cross-talk between microbiota and host organism, diet is the other key element modulating host response to inflammation. Diet is emerging as the most critical determinant in human health and healthy aging, thus suggesting that the study of the effects of food derived inflamm-aging miRNAs are urgently needed to elucidate whether some foods should be avoided in the aging people diet since they could harmfully regulate both inflammation related metabolism and healthy gut, thus contributing to unhealthy aging. With this in mind, we inquired the DMD (Dietary MicroRNA Database) web platform to retrieve foods derived miR-21, miR-155 and miR-146a that are 100% homologues to human ([Table 3](#)). Our results suggest that cow fat, cow milk, and eggs are rich in this trio-miRNAs (100% homologues to

human) and thus might target inflammation related pathways. Conversely none of the plant aliments in DMD yielded the inflamma-miRs trio. Thus, the information retrieved by enquiring the DMD repository would suggest restricting cow fat, milk and eggs and increase plant aliments which do not contain any of these trio-miRs. Of note is that human breast milk has 100% matches for the 3 inflamma-miRs from animal foods. Human breast milk is a particular nutrient used from birth to weaning only, and it is interesting that has a 100% match with animals (pig, caws, atlantic salmon, gallus) homologous miRs to be avoided in diet.

In order to uncover the network linking gut microbiota, inflamma-miRs, and diet we speculate on the possible converging signalling pathways. In Fig. 2 are reported some of these converging routes, crossing the results of Tables 1 and 2. As discussed above, lysine degradation pathway and fatty acids elongation pathways were the two most representative pathways (four out of six common elements). A cross talk between these two pathways is also evidenced from biochemical studies, as for example through carnitine production (lysine degradation) and fatty acids elongation (Longo et al., 2016).

Furthermore, some of the direct target of the inflamma-miRs trio are straightforwardly involved in the modulation of oxidative stress and autophagy. As for example miR-21 has as direct target BCL2 an anti-oxidant proteins family. A reduction of this protein suppresses mitochondria.

## 5. Conclusions

We demonstrated here, through a bioinformatics *in silico* research, that a cross-talk among gut microbiota, humans inflamm-aging miRs, and foods may occur through the sharing of signalling pathways. These pathways are relevant for contrasting or, *viceversa*, exacerbating inflammation associated senescence. Indeed, we postulate by this computational study that a putative multi-directional communication among i) microbiota; ii) dietary metabolism and iii) host metabolism exists. Dietary microRNAs affect pathways that regulate microbiota composition and gut microbiota affect these pathways as well. MicroRNAs act as a link between the two systems and both tune the host metabolism. Obviously, this observation must be validated experimentally, however we believe that this hypothesis is worthy and pave the way to the study of cross-talk among the systems.

Unfortunately, this scientific approach is still in its infancy, not many nutrimitomics studies are yet carried out and some show contradictory results. In addition a very few miRs food repositories are available as yet (Quintanilha et al., 2017). DMD for example contains miRs library for 15 aliments only. This fact highlights the urgent need for more research devoted to the study of the epigenetic role of foods and microbiota and the importance of nutrition for inflammation related aging.

## Ethical statement for solid state ionics – diffusion and reactions

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Testify that:

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- 3) All authors have been personally and actively involved in substantive work leading to the manuscript and will hold themselves jointly and individually responsible for its content.

## Declaration of Competing Interest

The authors declare no commercial or financial conflict of interest.

Guardare se questa Rivista lo vuole scritto qui o su una dichiarazione a parte

## Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.mad.2019.111127>.

## References

- Baier, S.R., Nguyen, C., Xie, F., Wood, J.R., Zemleni, J., 2014. MicroRNAs are absorbed in biologically meaningful amounts from nutritionally relevant doses of cow milk and affect gene expression in peripheral blood mononuclear cells, HEK-293 kidney cell cultures, and mouse livers. *J. Nutr.* 144 (10), 1495–1500. <https://doi.org/10.3945/jn.114.196436>.
- Carotenuto, F., Albertini, M., Coletti, D., Vilmercati, A., Campanella, L., Darzynkiewicz, Z., Teodori, L., 2016. How diet intervention via modulation of DNA damage response through MicroRNAs may have an effect on cancer prevention and aging, an *in silico* study. *Int. J. Mol. Sci.* 17 (5), 752. <https://doi.org/10.3390/ijms17050752>.
- Cătană, C., Calin, G.A., Neagoe, I., 2015. Inflammation-miRs in aging and breast cancer: are they reliable players? *Front. Med.* 2. <https://doi.org/10.3389/fmed.2015.00085>.
- Claesson, M.J., Jeffery, I.B., Conde, S., Power, S.E., O'Connor, E.M., Cusack, S., et al., 2012. Gut microbiota composition correlates with diet and health in the elderly. *Nature* 488 (7410), 178–184. <https://doi.org/10.1038/nature11319>.
- Custodero, D., Mankowski, R.T., Lee, S.A., Chen, Z., Wu, S., Manini, T.M., et al., 2018. Evidence-based nutritional and pharmacological interventions targeting chronic low-grade inflammation in middle-age and older adults: a systematic review and meta-analysis. *Ageing Res. Rev.* 46, 42–59. <https://doi.org/10.1016/j.arr.2018.05.004>.
- Dalmasso, G., Nguyen, H.T.T., Yan, Y., Laroui, H., Charania, M.A., Ayyadurai, S., et al., 2011. Microbiota modulate host gene expression via microRNAs. *PLoS One* 6 (4), e19293. <https://doi.org/10.1371/journal.pone.0019293>.
- Forslund, K., Hildebrand, F., Nielsen, T., Falony, G., Le Chatelier, E., et al., 2015. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. *Nature* 528 (7581), 262–266. <https://doi.org/10.1038/nature15766>.
- Franceschi, C., Garagnani, P., Vitale, G., Capri, M., Salvioli, S., 2017. Inflammaging and “Garb-aging”. *Trends Endocrinol. Metab.* 28 (3), 199–212. <https://doi.org/10.1016/j.tem.2016.09.005>.
- Gruver, A., Hudson, L., Sempowski, G., 2007. Immunosenescence of ageing. *J. Pathol.* 211 (2), 144–156. <https://doi.org/10.1002/path.2104>.
- Hallen, A., Jamie, J.F., Cooper, A.J.L., 2013. Lysine metabolism in mammalian brain: an update on the importance of recent discoveries. *Amino Acids* 45 (6), 1249–1272. <https://doi.org/10.1007/s00726-013-1590-1>.
- Hua, C., Zhao, J.-H., Guo, H.-S., 2018. Trans-kingdom RNA silencing in plant–fungal pathogen interactions. *Mol. Plant* 11 (2), 235–244. <https://doi.org/10.1016/j.molp.2017.12.001>. <http://sbbi-panda.unl.edu:5000/dmd/browse>, <http://snf-515788.vml.okeanos.grnet.gr/>, <https://www.ncbi.nlm.nih.gov/pubmed>. <https://pubmed.ncbi.nlm.nih.gov/31313131/>.
- Jimenez, J.A.L., Bordoni, A., Lorenzini, A., Rossi, C.A., Biagi, P.L., Hrelia, S., 1997. Linoleic acid metabolism in primary cultures of adult rat cardiomyocytes is impaired by aging. *Biochem. Biophys. Res. Commun.* 237 (1), 142–145. <https://doi.org/10.1006/bbrc.1997.7101>.
- Liu, S., da Cunha, A.P., Rezende, R.M., Cialic, R., Wei, Z., Bry, L., et al., 2016. The host shapes the gut microbiota via fecal microRNA. *Cell Host Microbe* 19 (1), 32–43. <https://doi.org/10.1016/j.chom.2015.12.005>.
- Longo, N., Frigeni, M., Pasquali, M., 2016. Carnitine transport and fatty acid oxidation. *Biochim. Biophys. Acta (BBA) – Mol. Cell Res.* 1863 (10), 2422–2435. <https://doi.org/10.1016/j.bbmr.2016.01.023>.
- Mariño, G., Pietrocola, F., Eisenberg, T., Kong, Y., Malik, S.A., Andryushkova, A., et al., 2014. Regulation of autophagy by cytosolic acetyl-coenzyme A. *Mol. Cell* 53 (5), 710–725. <https://doi.org/10.1016/j.molcel.2014.01.016>.
- Mu, J., Zhuang, X., Wang, Q., Jiang, H., Deng, Z., Wang, B., et al., 2014. Interspecies communication between plant and mouse gut host cells through edible plant derived exosome-like nanoparticles. *Mol. Nutr. Food Res.* 58 (7), 1561–1573. <https://doi.org/10.1002/mnfr.201300729>.
- Neish, A.S., 2009. Microbes in gastrointestinal health and disease. *Gastroenterology* 136 (1), 65–80. <https://doi.org/10.1053/j.gastro.2008.10.080>.
- Olivieri, F., Albertini, M.C., Orciani, M., Ceka, A., Cricca, M., Procopio, A.D., Bonafè, M., 2015. DNA damage response (DDR) and senescence: shuttled inflamma-miRNAs on the stage of inflamm-aging. *Oncotarget* 6 (34). <https://doi.org/10.18632/oncotarget.5899>.
- Olivieri, F., Rippon, M.R., Procopio, A.D., Fazioli, F., 2013. Circulating inflamma-miRs in aging and age-related diseases. *Front. Genet.* 4. <https://doi.org/10.3389/fgene.2013.00121>.
- Philip, A., Ferro, V.A., Tate, R.J., 2015. Determination of the potential bioavailability of plant microRNAs using a simulated human digestion process. *Mol. Nutr. Food Res.* 59 (10), 1962–1972. <https://doi.org/10.1002/mnfr.201500137>.
- Qin, Y., Wade, P.A., 2017. Crosstalk between the microbiome and epigenome: messages from bugs. *J. Biochem.* 163 (2), 105–112. <https://doi.org/10.1093/jb/mvx080>.
- Quintanilha, B., Reis, B., Duarte, G., Cozzolino, S., Rogero, M., 2017. Nutrimitomics: role of microRNAs and nutrition in modulating inflammation and chronic diseases. *Nutrients* 9 (11), 1168. <https://doi.org/10.3390/nu9111168>.
- Sherman, J.H., Choudhuri, S., Vicini, J.L., 2015. Transgenic proteins in agricultural biotechnology: the toxicology forum 40th annual summer meeting. *Regul. Toxicol. Pharmacol.* 73 (3), 811–818. <https://doi.org/10.1016/j.yrtph.2015.10.014>.

- Tomkovich, S., Jobin, C., 2015. Microbiota and host immune responses: a love-hate relationship. *Immunology* 147 (1), 1–10. <https://doi.org/10.1111/imm.12538>.
- Turchinovich, A., Weiz, L., Burwinkel, B., 2012. Extracellular miRNAs: the mystery of their origin and function. *Trends Biochem. Sci.* 37 (11), 460–465. <https://doi.org/10.1016/j.tibs.2012.08.003>.
- Valadi, H., Ekström, K., Bossios, A., Sjöstrand, M., Lee, J.J., Lötvall, J.O., 2007. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat. Cell Biol.* 9 (6), 654–659. <https://doi.org/10.1038/ncb1596>.
- Vital, M., Howe, A.C., Tiedje, J.M., 2014. Revealing the bacterial butyrate synthesis pathways by analyzing (meta)genomic data. *mBio* 5 (2). <https://doi.org/10.1128/mbio.00889-14>.
- Wang, K., Li, H., Yuan, Y., Etheridge, A., Zhou, Y., Huang, D., et al., 2012. The complex exogenous RNA spectra in human plasma: an interface with human gut biota? *PLoS One* 7 (12), e51009. <https://doi.org/10.1371/journal.pone.0051009>.
- Williams, M.R., Stedtfeld, R.D., Tiedje, J.M., Hashsham, S.A., 2017. MicroRNAs-based inter-domain communication between the host and members of the gut microbiome. *Front. Microbiol.* 8. <https://doi.org/10.3389/fmicb.2017.01896>.
- Witwer, K.W., 2012. Circulating MiRNA profiles. *RNA Biol.* 9 (9), 1147–1154.
- Zernecke, A., Bidzhekov, K., Noels, H., Shagdarsuren, E., Gan, L., Denecke, B., et al., 2009. Delivery of microRNA-126 by apoptotic bodies induces CXCL12-dependent vascular protection. *Sci. Signal.* 2 (100). <https://doi.org/10.1126/scisignal.2000610>.
- Zhang, C., Li, S., Yang, L., Huang, P., Li, W., Wang, S., et al., 2013. Structural modulation of gut microbiota in life-long calorie-restricted mice. *Nat. Commun.* 4 (1). <https://doi.org/10.1038/ncomms3163>.
- Zhang, J., Li, S., Li, L., Li, M., Guo, C., Yao, J., Mi, S., 2015. Exosome and exosomal microRNA: trafficking, sorting, and function. *Genomics Proteom. Bioinform.* 13 (1), 17–24. <https://doi.org/10.1016/j.gpb.2015.02.001>.
- Zinkernagel, M.S., Zysset-Burri, D.C., Keller, I., Berger, L.E., Leichtle, A.B., Largiadèr, C.R., et al., 2017. Association of the intestinal microbiome with the development of neovascular age-related macular degeneration. *Sci. Rep.* 7 (1). <https://doi.org/10.1038/srep40826>.