



Exercise does not ameliorate cardiac dysfunction in obese mice exposed to fine particulate matter

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ABSTRACT

Background: Studies have demonstrated that exposure to fine particulate matter (PM_{2.5}) is linked to cardiovascular disease (CVD), which is exacerbated in patients with pre-existing conditions such as obesity. In the present study, we examined cardiac function of obese mice exposed to PM_{2.5} and determined if mild exercise affected cardiac function.

Methods: Obese mice (*ob/ob*) (leptin deficient, C57BL/6J background) were exposed to either filtered air (FA) or PM_{2.5} at an average concentration of 32 µg/m³ for 6 h/day, 5 days/week for 9 months. Following exposure, mice were divided into four groups: (1) FA sedentary, (2) FA treadmill exercise, (3) PM_{2.5} sedentary, and (4) PM_{2.5} treadmill exercise and all mice were analyzed after 8 weeks of exercise training.

Results: Echocardiography showed increased left ventricular end systolic (LVESd) and diastolic (LVEDd) diameters in PM_{2.5} sedentary mice compared to FA sedentary mice. There was increased expression of ICAM1, VCAM and CRP markers in sedentary PM_{2.5} mice compared to FA mice. Both FA and PM_{2.5} exercised mice showed decreased posterior wall thickness in systole compared to FA sedentary mice, coupled with altered expression of inflammatory markers following exercise.

Conclusion: Obese mice exposed to PM_{2.5} for 9 months showed cardiac dysfunction, which was not improved following mild exercise training.

1. Introduction

Airborne particulates have gained attention regarding their significance on human health as reports claim the environmental stressor ranks 9th in overall cause of mortality worldwide [1]. Notably, exposure to particulate matter < 2.5 µm in diameter (PM_{2.5}) has been shown to increase cardiovascular disease occurrence by penetrating deeply into the lungs and diffusing into the bloodstream to produce a host of deleterious effects [2]. The above mechanisms ultimately lead to an inflammatory response by the heart and lungs, a factor contributing to abnormal cardiac function [3].

According to the Center for Disease Control and Prevention, nearly 93.3 million people in the United States are affected by obesity, contributed in part by a poor diet and sedentary lifestyle. Obese individuals are predisposed to developing cardiovascular disease (CVD), and

obesity and air pollution exposure further act as comorbid factors leading to increased CVD incidence. One of the major contributors to CVD is the persistent chronic systemic inflammation which induces cardiac dysfunction [4–6]. In particular, left ventricular (LV) filling pressure, an index of LV function, is found to be impaired in obese individuals [7,8].

Exercise is an important therapeutic tool used as a preventative measure against obesity and the resultant cardiac dysfunction. Varying cardiac responses occur with respect to differing exercise intensities. Studies examining these differences typically note greater cardiac remodeling in high intensity regimens [9] compared to little or no cardiac remodeling in low intensity exercise regimens [10]. Emerging evidence suggests that exercise reverses adverse cardiac remodeling, ultimately leading to a lower occurrence of CVD [11]. In a mouse model of diet-induced obesity, exercise training has a beneficial effect on cardiac

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health by enhancing cardiac function and improving mechanical efficiency [12]. In contrast, another study showed that exercise was not able to reverse abnormalities in expression of L-type calcium channels in diet-induced obesity in rats, leading to continued cardiac dysfunction [13]. Similar studies have shown that endurance exercise in obese mice aggravated cardiac abnormalities through increased fibrosis and impaired mitochondrial biogenesis, although improved skeletal muscle metabolism was still observed [14]. Studies utilizing the *ob/ob* obese mouse model indicated varying results following exercise, including one by Sennott et al. that found improved blood glucose levels in mice assigned to voluntary running compared to both treadmill and sedentary groups [15].

Studies conducted on *ob/ob* mice reported cardiac hypertrophy and altered metabolism in response to defective leptin signaling [16,17]. Previous work in our lab has demonstrated adverse cardiac function due to PM_{2.5} exposure in both adult and adolescent mice through LV remodeling and impaired sarcomere function [18,19]. Introducing additional stressors, such as PM_{2.5} exposure, has not yet been explored in the *ob/ob* model. The aim of the current study was to examine cardiac effects in PM_{2.5}-exposed obese mice following mild treadmill exercise training.

2. Materials and methods

2.1. Animals and exposure

All animal experiments were performed in alignment with NIH guidelines approved by the Institutional Animal Care and Use Committee (IACUC) at The Ohio State University, Columbus, Ohio. Twelve week old male *ob/ob* mice (C57BL/6J background strain) were obtained from Jackson Laboratories (Bar Harbor, ME). The *ob/ob* mouse is leptin deficient as a result of a homozygous mutation in the *ob* gene. All mice were housed in our AAALAC-approved facility for one week before beginning the 9-month exposure period. Animals were exposed to concentrated PM_{2.5} from the Columbus, OH region using the system "Ohio's Air Pollution Exposure System for the Interrogation of Systemic Effects" (OASIS-1) aerosol concentration system located at the Ohio State University [18]. Animals were exposed either to PM_{2.5} or filtered air (FA), which included an identical system with the exception of a HEPA filtered inserted at the inlet to remove all ambient particulates. The average PM_{2.5} concentration that the animals were exposed to in this study was 32 µg/m³ over the exposure period, similar to concentrations seen in our previous studies [20,21]. Following exposure, all mice were housed in room air for the duration of treadmill exercise.

2.2. Treadmill exercise protocol

Following 9 months of exposure (12 months of age), the mice were separated into the following groups: 1) FA sedentary (FA Sed; n = 5), 2) PM_{2.5} sedentary (PM_{2.5} Sed; n = 5), 3) FA exercise (FA Ex; n = 3), and 4) PM_{2.5} exercise (PM_{2.5} Ex; n = 4). Mice were not exposed to PM_{2.5} during the exercise period. All animals were group housed under similar conditions. Initially, mice were exposed to adaptive training at 2.5 m/min for 10 min per session for the first three days. Exercise involved forced treadmill training 5 days/wk, consisting of 30 min of

daily training at a speed of 2.5 m/min at 0% incline for 8 weeks. Mice were monitored throughout the exercise periods and electrical stimulus used to motivate animals during exercise.

2.3. Echocardiography

Mice in all four groups underwent echocardiographic assessment using a 40 MHz transducer using the Vevo 2100 (Visualsonics, Toronto, ON, Canada). Isoflurane was administered at 1.5–2% (delivered with 100% O₂) as anesthesia through a nose cone and body temperature was maintained at 37 °C. Following fur removal, pre-warmed ultrasound gel (Aquasonic; Parker Laboratories, Fairfield, NJ) was applied onto the chest. A 15 MHz probe was placed in the parasternal, short axis orientation. M mode data was collected and analyzed with the average of three cine loops for the following parameters: LV end-systolic and end-diastolic internal dimensions (LVESd and LVEDd), as well as systolic and diastolic posterior wall thickness (PWTs and PWTd). Percent fractional shortening (%FS) was calculated using the equation: % FS = [(LVEDd - LVESd)/LVEDd * 100]. All analyses were performed in accordance with the American Heart Association defined technique by an investigator blinded to group assignments.

2.4. Quantitative real-time PCR

Total RNA was extracted from snap frozen left ventricular cardiac tissue via the RNeasy kit (Qiagen, Hilden, Germany). Concentrations were determined using a NanoDrop 2000c (Thermo-Scientific, Wilmington, DE). iScript Supermix kit (Bio-Rad, Hercules, CA) was used to reverse transcribe 1 ng of RNA to generate cDNA. This process used the CFX96 Thermocycler (Bio-Rad, Hercules, CA). Primers were used at a final concentration of 0.25–0.5 µM for target genes. Gene-specific primer sequences listed in Table 1 were used and normalized to *Gapdh* expression. The formula $2^{-\Delta\Delta Ct}$ was used to quantify relative gene expression (where C_t is threshold cycle) [22]. The three-step amplification protocol was as follows: denaturation at 95 °C for 10 min followed by 39 cycles of denaturation (95 °C, 1 s), annealing (65 °C, 10 s), and extension (72 °C, 20 s).

2.5. Statistical analyses

Data were assessed using Prism 6.0 (Graphpad Software, San Diego, CA) and differences were considered statistically significant when $P \leq 0.05$ via Student's *t*-test or one-way ANOVA followed by Tukey's *post hoc* analyses.

3. Results

3.1. PM_{2.5} exposure does not affect body weight or heart weight in obese sedentary or exercised mice

To study the effects of mild exercise on biometric parameters, we compared the exercised animals to not only age-matched sedentary controls but also evaluated the difference due to FA or PM_{2.5} exposure. Body weight (BW), heart weight (HW), HW/BW ratio remain unaltered in sedentary as well as exercised groups (Table 2).

Table 1
Primer sequences used for PCR amplification.

Gene	Forward Primer	Reverse Primer
<i>Gapdh</i>	ATGGTGAAGGTCGGTGTGAACGG	AGGGTTCGTGTGATGGCAACAATCT
<i>CRP</i>	GTCTGCTACGGGATTGTAGA	CACCGCCATACGAGTCCTG
<i>ICAM-1</i>	TGCCTCTGAAGCTCGGATATAC	TCTGTGCAACTCCTCAGTCAC
<i>VCAM-1</i>	GTTCACGCGAGGGTCTACC	AACTCTTGGCAACATTAGGTTG

Table 2
Biometric data of FA and PM_{2.5} exposed sedentary and exercise mice.

	FA Sedentary (n = 5)	PM _{2.5} Sedentary (n = 5)	p-value	FA Exercise (n = 3)	PM _{2.5} Exercise (n = 4)	p-value
Body Weight, g	70.40 ± 2.52	64.91 ± 8.74	0.56	65.87 ± 2.97	67.85 ± 2.41	0.62
Heart Weight, mg	180 ± 10	200 ± 10	0.35	180 ± 20	180 ± 20	0.99
HW/BW, mg/g	2.61 ± 0.12	3.32 ± 0.39	0.11	2.70 ± 0.19	2.79 ± 0.19	0.77

FA, filtered air; PM_{2.5}, particulate matter (< 2.5 μm diameter).

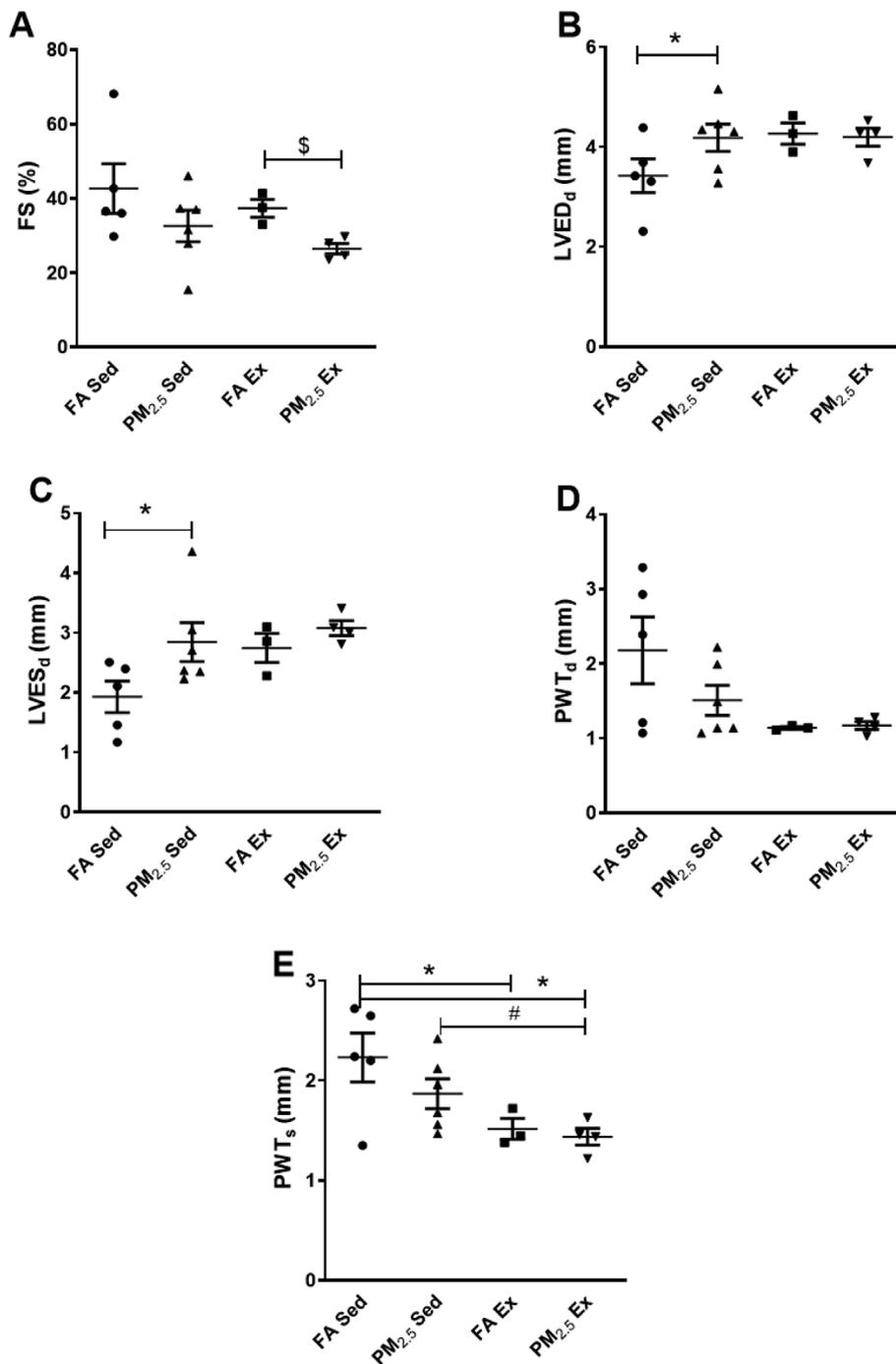


Fig. 1. Echocardiographic parameters comparing cardiac functional changes among FA (Sed: n = 5; Ex: n = 3) or PM_{2.5} (Sed: n = 5; Ex: n = 4) exposed sedentary and exercise mouse groups. A. Fractional shortening (%FS), B. Left ventricular end-diastolic diameter (LVEDd), C. Left ventricular end-systolic diameter (LVESd), D. Posterior wall thickness during diastole (PWTd), E. Posterior wall thickness during systole (PWTs). Five beat cycles were captured and three loops averaged per assessment. Data are expressed as ± S.E.M. *p ≤ 0.05 vs FA sedentary, #p < 0.05 vs PM_{2.5} sedentary, ^{ss}p < 0.01 vs FA exercise.

3.2. PM_{2.5} exposure leads to increased LV dimensions of obese sedentary mice

Echocardiographic analyses of sedentary mice exposed to PM_{2.5} demonstrated increased LVESd (1.93 ± 0.26 mm FA Sed; 2.84 ± 0.32 mm PM_{2.5} Sed; $p = 0.03$) and LVEDd (3.42 ± 0.33 mm FA Sed; 4.18 ± 0.27 mm PM_{2.5} Sed; $p = 0.05$) (Fig. 1B & C). Posterior wall thickness was not different during systole (PWTs) (2.23 ± 0.24 mm FA Sed; 1.86 ± 0.14 mm PM_{2.5} Sed; $p = 0.10$) or diastole (PWTd) (2.17 ± 0.44 mm FA Sed; 1.50 ± 0.20 mm PM_{2.5} Sed; $p = 0.90$) between both groups (Fig. 1D & E). A slight decrease in %FS (42.70 ± 6.70 FA Sed; 32.60 ± 4.24 PM_{2.5} Sed; $p = 0.10$) was observed in PM_{2.5} sedentary mice compared to FA exposed control mice (Fig. 1A).

3.3. PM_{2.5} leads to LV contractile dysfunction in obese exercise mice

Following PM_{2.5} exposure, 8 weeks of treadmill exercise resulted in a significant reduction in %FS (37.37 ± 2.41 FA Ex; 26.54 ± 1.42 PM_{2.5} Ex; $p = 0.009$) in PM_{2.5}Ex compared to FA Ex mice (Fig. 1A). While this decreased %FS is consistent with ventricular systolic dysfunction, other measurements such as LVESd (2.74 ± 0.24 mm FA Ex; 3.08 ± 0.12 mm PM_{2.5} Ex; $p = 0.12$), LVEDd (4.26 ± 0.21 mm FA Ex; 4.19 ± 0.18 mm PM_{2.5} Ex; $p = 0.81$), PWTs (1.51 ± 0.10 mm FA Ex; 1.43 ± 0.08 mm PM_{2.5} Ex; $p = 0.57$) and PWTd (1.14 ± 0.01 mm FA Ex; 1.17 ± 0.05 mm PM_{2.5} Ex; $p = 0.66$) for ventricular remodeling were not different (Fig. 1B–E).

When comparing sedentary groups (both FA and PM_{2.5}) with exercised groups (both FA and PM_{2.5}), we observed a significant difference only in PWTs (Fig. 1E), while the remaining parameters were unchanged.

3.4. PM_{2.5} exposure increases inflammatory mRNA expression in obese sedentary mice with no effect in exercised mice

We analyzed markers of cardiac inflammation in sedentary mice exposed either to FA or PM_{2.5}. Measurements of gene expression demonstrated significantly increased C-reactive protein (CRP), an inflammatory protein associated with long term diseases and immune activation after PM_{2.5} exposure. Additionally, markers for intracellular adhesion molecule 1 (ICAM1) and vascular cell adhesion protein (VCAM), both important for leukocyte signaling, were increased in PM_{2.5} exposed mice compared to FA controls (Fig. 2). There was no difference observed in the expression of CRP, ICAM and VCAM between FA Ex and PM_{2.5} Ex mice (Fig. 2). Additionally, comparison of sedentary to exercised mice yielded no significant difference within either exposure group for each marker analyzed (Fig. 2).

4. Discussion

It is well known that a sedentary lifestyle and obesity coexist and that both are associated with CVD [23,24]. As obesity positively correlates with heart failure, atrial fibrillation, coronary artery disease, and hypertension [25,26], it is considered as one of the leading risk factors for CVD. Several studies have also indicated that aged obese mice have increased cardiomyocyte apoptosis and decreased survival [27], coupled with LV diastolic functional abnormalities [28–30]. PM_{2.5} exposure, on the other hand, has recently been determined to be a leading causative agent for the development of CVD [18,20,21]. Exercise is an effective treatment for improving cardiometabolic function in obese individuals; however, the effects of exercise on cardiac function after PM_{2.5} exposure in obese individuals has not been fully defined. The goal of the present study was to examine the effects of forced treadmill exercise on cardiac function of leptin deficient obese (*ob/ob*) mice exposed to PM_{2.5}. Following 8 weeks of mild treadmill exercise, we did not observe changes in biometric parameters between PM_{2.5} exposed sedentary and exercised mice. In PM_{2.5} exposed sedentary mice, there was evidence of contractile dysfunction manifested as altered LV internal dimensions both during systole and diastole, suggestive of contractile dysfunction potentially leading to LV volume overload. Our findings are similar to a cardiac phenotype observed in previous studies [28–30].

While molecular mechanisms remain unclear, activation of an inflammatory response (CRP, ICAM1, and VCAM) could be one of the mechanisms behind cardiac dysfunction observed in PM_{2.5} exposed sedentary mice as it is regarded as a stimulus to various cardiac pathophysiological events. Upregulation of inflammatory markers identifies additional physiological stress as inflammation has been identified as a response to myocardial damage [31]. Several studies demonstrated a link between pro-inflammatory cytokines and left ventricular hypertrophy and dysfunction [32]. This pro-inflammatory response is significant as chronic levels of inflammatory proteins have been observed to contribute to and exacerbate insulin resistance in cardiomyocytes, leading to impaired metabolic function [33]. Further, obese subjects exposed to particulate matter demonstrated changes in the DNA methylation of CD14 and TLR4, pathways heavily involved in an inflammatory response [34]. Increased CRP levels have been observed in those with obesity and pre-clinical diabetes [35], demonstrating a link between obesity and CRP regulation as seen in our study.

Data suggest that regular exercise has beneficial effects on cardiovascular function partly by eliciting anti-inflammatory properties, resulting in decreased CVD development and favorable cardiac remodeling [36]. Therefore, we evaluated the cardiac effects of exercise in PM_{2.5} exposed obese mice. A recent study found that exercise later in life benefitted LV function in middle-aged people who lived a sedentary lifestyle [37], suggesting exercise during later periods of life may

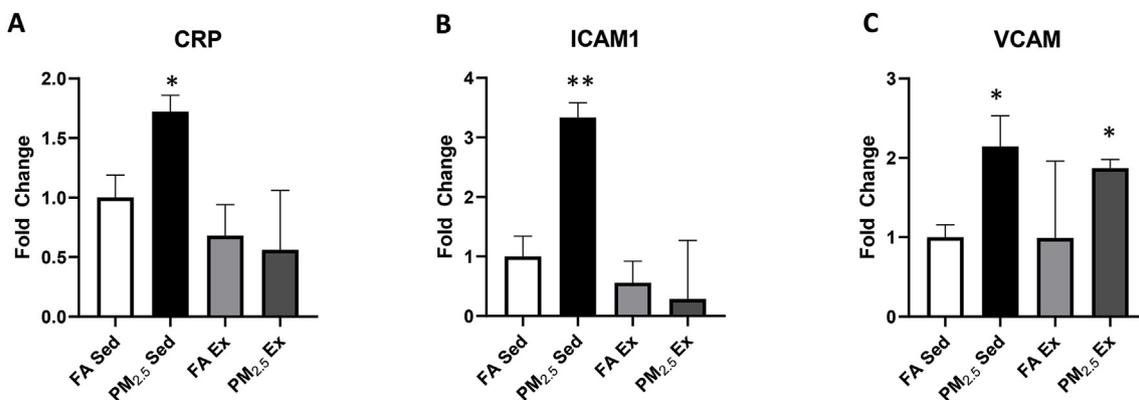


Fig. 2. Quantitative polymerase chain reaction (qPCR) analysis of RNA samples comparing changes among FA or PM_{2.5} exposed sedentary and exercise mouse groups. A. CRP, B. ICAM1, C. VCAM. Results are mean \pm S.E.M. Data are expressed as \pm S.E.M. * $p < 0.05$, ** $p < 0.01$ vs FA sedentary.

produce cardiac benefits. In our study, however, we found no improvement in cardiac function when mice underwent mild exercise training. Exercised *ob/ob* mice exposed to PM_{2.5} showed even worse cardiac function with significantly decreased %FS consistent with that observed in heart failure. There was also no change in the expression of inflammatory markers following exercise training in FA and PM_{2.5} exposed *ob/ob* mice. Therefore, contrary to previous evidence, mild exercise may have introduced an additional stressor to cause a further decline in cardiac function.

5. Conclusion and future directions

The present study aimed at identifying the potential CVD burden in PM_{2.5} exposed obese mice. The national ambient air quality standard (NAQS) set by the U.S. Environmental Protection Agency (U.S. EPA) for PM_{2.5} are 35 µg/m³/per day and 12 µg/m³/year. The levels achieved in 9 months of exposure in this study are significantly more than the annual limit. This suggests that highly polluted areas which are exceeding EPA standards pose serious risks to those who have risk factors for CVD such as obesity. *ob/ob* sedentary mice demonstrated contractile dysfunction following PM_{2.5} exposure. Mild exercise intervention for 8 weeks failed to improve cardiac function following 9 months of exposure to *ob/ob* PM_{2.5} mice. Exercise training has been shown to exert beneficial effects on cardiovascular function, and the positive outcomes have been substantiated in various experimental models of exercise training [38,39]. However, our results indicated that mild forced exercise does not result in cardioprotection in the PM_{2.5} exposed *ob/ob* mouse model, exacerbating rather than attenuating cardiac dysfunction. We acknowledge factors that may contribute to disease exacerbation due to exercise (1): long-term PM_{2.5} exposure resulted in significant aging of the mice which is considered as the largest risk factor for CVD [40]. In this study, animals were aged by the time they were subjected to mild treadmill training, which could have served as an additional deleterious factor along with PM_{2.5} exposures compared to young mice used in other positive outcome studies (2): type of exercise could also have beneficial or adverse effects on the cardiac outcomes. In our study, we used a mild exercise training protocol that did not exert beneficial cardiac effects (3): we observed impaired exercise tolerance in *ob/ob* mice and consider it as the limiting factor in exercise associated beneficial effect. Leptin selectively stimulates phosphorylation and activation of the α2 catalytic subunit of AMPK (α2-AMPK), an enzyme that appears to have a significant role in the promotion of exercise tolerance in skeletal muscle [41]. Our present study utilized leptin deficient *ob/ob* mice and hence impaired α2-AMPK activity that may directly contribute to reduced exercise capacity resulting in non-beneficial effects in both FA and PM_{2.5} exposed exercised groups. We additionally acknowledge three limitations of the study design (1): A single echocardiographic time point did not permit analysis of cardiac changes during PM_{2.5} exposure or exercise (2): Low animal numbers, especially in the FA Ex group, may have contributed to a large error bar in VCAM data; and (3): qPCR data were not confirmed with protein or histological experiments. Thus, on the basis of these findings, further studies are required to determine whether leptin deficiency is contributing to the aberrant response to exercise. Future studies should also examine the effect of early interventional strategies in CVD animal models exposed long term to high levels of ambient particulates.

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