



## Targeting antigen presentation in autoimmunity<sup>☆</sup>

Jason R. Lees<sup>a,\*</sup>

<sup>a</sup> Department of Medicine, Uniformed Services University of the Health Sciences, Bethesda, MD, United States



### ARTICLE INFO

#### Keywords:

Antigen presentation  
Autoimmunity  
Inflammation  
Immune regulation

### ABSTRACT

Autoimmune diseases are heterogeneous group of disorders that together represent an enormous societal and medical problem. CD4<sup>+</sup> T cells have critical roles in the initiation and pathogenesis of autoimmune disease. As such, modulation of T cell activity has proven to have significant therapeutic effects in multiple autoimmune settings. T cell activation is a complex process with multiple potential therapeutic targets, many of which have been successfully utilized to treat human disease. Current pharmacological treatment largely targets T cell intrinsic activities as a means of treating various autoimmune disorders. Here I review extant and potential therapeutic approaches that instead specifically target antigen presentation to CD4<sup>+</sup> T cells as a critical checkpoint in autoimmune responses. In addition, the contribution of antigen modulation components in current therapeutic approaches is considered along with the impact of new antigen targeted treatment modalities. Finally, potential challenges are considered in the context of the potential for antigen specific targeting of the antigen presentation process.

### 1. Introduction

Although the exact number of disorders that can be considered autoimmune diseases remains an area of contention, dozens of disorders are recognized as including a self-directed adaptive immune component. One commonality of characterized and putative autoimmune disease is that T cell effector function is a critical component to disease initiation and/or continuation [1,2]. So it is not surprising that many of the current therapies for autoimmune disorders target T cell activation and effector function. Multiple components of T cell activity have been targeted in an effort to ameliorate disease, including depletion of lymphocyte subsets, direct modulation of intrinsic T cell signaling, blocking of costimulatory signals critical for productive antigen recognition, and targeting of effector molecules such as cytokines. While these approaches have certainly proven fruitful, they have also tended to be relatively generalized approaches to immunomodulation, with concomitant impacts on immunity to infectious agents. Thus, while these broad immunosuppressive approaches have been enormously successful in transplantation settings, they have proven less universally accepted in autoimmune settings that represent less immediate risks of mortality.

In addition to general immunosuppressive protocols, advances have also been made in modulation of antigen-specific autoimmune

responses. Antigen specific modulation of autoimmunity is a highly sought goal, as it would allow therapeutic approaches that would not hinder immune reactivity to pathogens, preventing many of the adverse effects that limit current immune modulatory approaches to treatment of autoimmunity. Methods utilized often focus on targeting of T cells of known antigen specificity, either by directly targeting epitope specific lymphocytes or by induction or activation of regulatory cell populations that respond to the same antigen. The successes and challenges of these approaches have been extensively reviewed previously [3–6], and thus will not be covered in depth here. In addition several alternate approaches have been reported that attempted to directly target auto-antigen presentation and/or antigen-presenting cells in autoimmunity, with varying efficacy.

### 2. Antigen presentation as a risk factor for autoimmunity

The use of twin studies and the explosion in available genetic data have revealed fascinating insights into the role of genetics in autoimmunity [7–10]. Perhaps because autoimmunity is such a broad categorization, the impact of genetics on individual autoimmune manifestations is wildly divergent, with genetic predisposition accounting for anywhere between 5% and 90% of established risk for individual disorders. However, while sensitive genetic techniques have now

<sup>☆</sup> The content and views expressed in this paper are the sole responsibility of the author and do not necessarily reflect the views or policies of the Department of Defense or the US government. Mention of trade names, commercial products, or organizations does not imply endorsement by the US Government.

\* Address: 4301 Jones Bridge Rd, A-3073, Bethesda, MD 20814, United States.

E-mail address: [jason.lees@usuhs.edu](mailto:jason.lees@usuhs.edu).

revealed multiple potential contributors to disease, factors involved in antigen presentation are among the most highly represented.

Antigen presentation is a complex biological process, with many molecular contributors, involving the production, peptide-loading, and localization of MHC molecules to the cell surface where the MHC/peptide complex can interact with passing or recruited T cells in a cognate manner. Specific alleles of the both MHC class I and MHC class II molecules were some of the first identified and remain by far the most significant genetic risk factor for multiple autoimmune diseases [11–15]. Further, there is evidence to suggest that the risk of disease associated with expression of specific MHC proteins is modified by complex interactions between multiple MHC alleles [16].

Genetic differences in other aspects of the antigen presentation pathway, particularly the molecular mechanisms responsible for the peptide processing and loading that results in epitope selection, have also been linked to increased risk for some autoimmune diseases. As an example, allelic differences in endoplasmic reticulum aminopeptidase proteins (ERAP), which result in subtle changes in the peptides displayed on MHC, are associated with significantly increased risk of ankylosing spondylitis [17,18]. Other changes in peptide production or display, including some induced by pathogen driven inflammatory conditions, have also been shown to impact disease in multiple autoimmune settings [19,20]. Indeed, multiple studies have demonstrated that altering the ligands presented, often by providing large concentrations of modified peptides can greatly impact the T cell response [21–24].

Together, the studies discussed above provide strong evidence that MHC/T cell interactions are critical determinants of autoimmune disease. However, the genetic data cannot determine at what stage(s) of disease MHC/T cell interactions are required. Thus, an important question to consider is whether T cell/MHC interaction is required during disease initiation and/or on an ongoing basis throughout disease. To address this question early studies utilized antibodies specific for MHC class II molecules to both prevent and treat experimental autoimmune disease, suggesting that T cell/MHC interactions are important throughout the disease process [25–29]. Similarly, use of antibodies specific for the antigen receptor of clonal lymphocyte populations have been shown to modulate autoimmune disease [30,31].

In addition many other studies have made use of classic depletion methods to address the impact of different subsets of antigen presenting cells (APC) on disease induction and severity at different stages of disease. For instance, in models of the autoimmune disease type 1 diabetes, multiple types of APC appear to be critical to the initiation of disease [32,33]. However, it is important to note that these studies do not demonstrate the exact roles of the APC to disease development and thus the relative importance of antigen presentation, as opposed to other aspects of APC effector function to pathogenesis. Using a specific modulator of epitope generation and selection in diabetes models prevented pancreatic cell destruction, suggesting that antigen presentation is at least necessary to disease development [34–36]. Similarly, in experimental thyroiditis, use of a small molecule inhibits antigen presentation and subsequent disease [37].

### 3. Evidence of antigen presentation as a critical requirement for ongoing autoimmunity

Unfortunately, as neither patients nor physicians can accurately predict onset of autoimmunity, therapeutic intervention in established disease is typically required in the clinic. The critical requirements for T cell activity in disease initiation can make determining the impact of antigen-presentation in later phases of disease difficult. Because T cells generally rely on presentation of MHC/auto-antigen peptides within the target tissue to drive their effector functions, continued antigen presentation would appear to be a critical component in continuation of any established autoimmune setting that is known to be modulated

with T cell targeted therapeutics. Excellent examples of such disorders would be RA and RRMS, both of which can be treated with T cell targeted therapeutics. Models developed to examine experimental depletion of specific APC subsets after development of disease reveal that the ongoing presence of antigen presenting cells within the target tissue [38] is essential for pathogenesis [39–42].

Again it is important to note that these findings do not necessarily demonstrate that antigen presentation itself is required for ongoing autoimmune disease, as APC can exert multiple impacts within a tissue beyond T cell activation. As such depletion methods may not accurately identify the actual role(s) antigen presentation plays in pathogenesis after autoimmune disease initiation. In support of an ongoing necessity for APC/T-cell interaction, in an autoimmune demyelination model, altered peptide ligands were found to modify T cell responses and ameliorate disease, suggesting that ongoing T cell activation is necessary for continued disease and could be targeted therapeutically [21–24]. Similar approaches were described in RA [43]. Further work has suggested several potential methods of modulating peptide usage in autoimmunity [44].

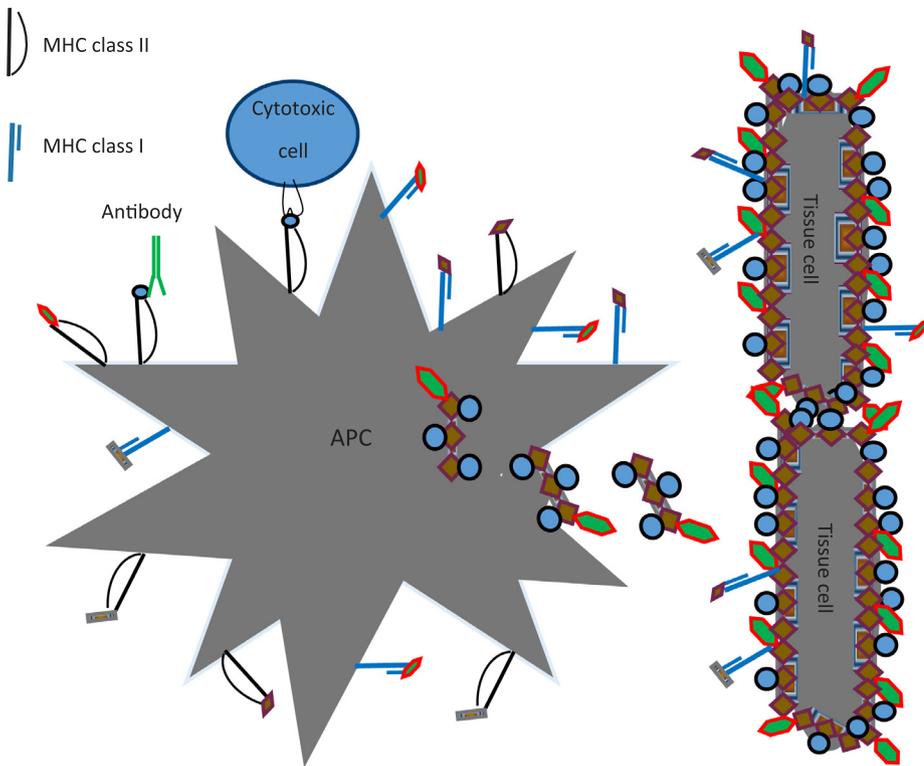
The nature of immune attack tends to result in increased antigen presentation during autoimmune inflammatory events [45]. During autoimmunity tissue destruction can result in antigen uptake and subsequent presentation, acting as a positive feedback loop to drive further autoimmunity through both increased antigen availability and production of new responses to previously unrecognized epitopes [46]. In several types of autoimmunity, these inflammation associated changes in presentation appear to be critical to the pathogenesis of disease [47]. The requirement for secondary responses to previously occult antigens, often called epitope spreading, in the pathogenesis of experimental neuroinflammation and arthritis models is a strong argument for the necessity of ongoing antigen presentation in established autoimmune disease.

### 4. Impact of current therapeutic approaches on antigen presentation

Given the heterogeneity of autoimmune disorders it is likely that blocking antigen presentation could be efficacious in some disorders while being completely ineffective in others. The efficacy of current therapeutics that are thought to work by modulating T cell activity would seem to be a reasonable guide. Broadly active therapeutics, such as corticosteroids, will have substantial effects on a multitude of cell types within the body, and have been shown to interfere with productive antigen presentation. Indeed any approach thought to act via modulation of innate immune parameters will most likely impact antigen presentation, as is clearly the case for newly recognized immunomodulators such as statins [48].

It is important to consider that, given the complex feedback mechanisms that regulate immunity, any therapeutic that modulates one aspect of the immune response, even by very narrow targeting of effector molecules, can have profound effects on other aspects of immunity. One example of this is the use of TNF $\alpha$  inhibitors in the treatment of multiple autoimmune disorders, including RA and Crohn's disease [49]. TNF $\alpha$  is a critical effector cytokine driving autoimmune pathology and as such its neutralization will have direct effects on disease outcomes [50]. In addition, TNF $\alpha$  is known to influence the antigen presentation capacity of antigen presenting cells. As such, TNF $\alpha$  inhibitors almost certainly have a secondary effect on antigen presentation [51], although the relative contribution of each effect of TNF $\alpha$  inhibition to disease amelioration is an open question.

Of the professional antigen presenting cells, B cells have been the focus of many recent studies. The capacity to deplete mature B cells has suggested that B cells contribute to multiple forms of autoimmune inflammation [52]. B cells can utilize multiple mechanisms in autoimmune responses, including production of antibody and cytokines [53,54], but recent data suggests that B cells may also have critical



**Fig. 1.** Targeting known MHC/peptide can also block presentation of additional associated tissue proteins. Proteins sampled from tissue cells will tend to be taken up as a complex resulting in multiple epitopes from a tissue being presented by the same antigen presenting cell. Targeting a single MHC class II/peptide can block presentation of multiple associated peptides while sparing MHC class II negative tissue cells.

antigen presenting roles in some forms of autoimmunity [55–57]. Examination of the role of specific B cell activities in autoimmune disease is extremely difficult, however studies using experimental autoimmune animal model systems have provided some important data. For instance, B cell antigen presentation appears to be a critical component to the initiation of spontaneous type I diabetes in the NOD model [58,59]. As a consequence depletion of B cells using a variety of methods results in reduced diabetes onset [60–63]. As an additional example, in MS the therapeutic success of anti-B cell antibody treatment [64,65] has suggested that B cells must play a critical role in ongoing neuroinflammation, however the exact mechanism is still unclear. Studies done in the experimental autoimmune encephalomyelitis model of MS has suggested that the critical role for B cells may be antigen presentation, with the caveat that B cell involvement in EAE disease is highly system dependent [66,67]. However, additional studies in EAE have suggested that B cell antigen presentation is sufficient to drive EAE responses when other APC are not present [68,69], suggesting that these cells can be a critical antigen presenting cell population during neuroinflammation.

Together, these data indicate that some current therapeutic approaches may partially rely on inhibition of antigen presentation to produce clinical benefit. This suggests that antigen presentation is a reasonable target for ongoing attempts to modulate immunity. Given the already mentioned advantages of antigen specific regulation, modulation of antigen presenting cell activity, in an antigen dependent manner, would appear to be an exciting goal.

## 5. Antigen specific targeting of antigen presenting cells

Early attempts to target antigen specific cell populations relied on antibody responses to unique determinants within the variable regions of clonal antigen receptors [30,31]. This is an effective method for targeting individual clonotypes, but requires identification of the specific idiotypes of dominant clones and the production of antibodies to those idiotypes. Conversely, there are several examples of the production of antibodies specific for peptide in the context of MHC [70]. These reagents should allow one to target specific antigen displayed on APC in

the context of MHC.

MHC class I molecules are ubiquitously expressed while MHC class II molecules have a highly limited distribution. As outlined above, presentation of MHC class II (MHCII)/myelin peptides to  $CD4^+$  T cells within target tissue is thought to be a critical component of initiation and continuation of autoimmune inflammation. As such emerging protocols that would allow antibody mediated direct visualization and targeting of defined peptide/MHC II complexes may provide us with new ways to approach autoimmune treatment.

Recent work has delineated methods for successfully introducing new antigen receptors into mature cytotoxic T cells [71]. These cells have been found to be efficacious for antigenic cell elimination in a variety of settings. Antigen specificity was provided via transgenic expression of either cloned MHC class I/peptide responsive TCR or engineered antigen receptors that recognized native antigen without MHC restriction. The emergence of T cells transduced with defined antigen receptors has opened up new possibilities for targeting autoimmunity in an antigen specific manner [3,5]. Indeed, several exciting studies have emerged demonstrating T cell directed therapies produced by transduction of regulatory T cells [72].

While multiple groups examined the consequences of expressing a MHC class I restricted TCR in mature  $CD4^+$  T cells [73–75] no one has yet reported the functional consequences of expressing a MHCII restricted TCR in mature  $CD8^+$  cells. Interestingly, functional MHCII restricted  $CD8^+$  T cells have been reported in circumstances of  $CD4$  deficiency [76] or viral pathogenesis [77] and there is evidence that cytotoxicity can occur without  $CD8$  co-receptor activation [78]. Further, cytotoxic  $CD8^+$  T cells have been shown to modulate ongoing antigen presentation by killing antigen-decorated antigen presenting cells (APC) in a variety of systems [79–86]. These findings suggest that introduction of a MHCII/peptide responsive TCR into mature  $CD8^+$  T cells may allow cells to recognize and lyse cells bearing the cognate antigen, providing a new method for targeting APC displaying specific epitopes.

Regardless of the method used, one major advantage of targeting specific antigen(s) displayed on APC is that the targeted APCs would be expected to present multiple self-derived peptides from the same tissue,

due to the nature of antigen processing from phagocytized or endocytized complex material. Importantly, that would mean that targeting APC for modulation with a single defined antigen, using either antibody or transgenic cytotoxic cells, would also impact the presentation of any additional tissue specific antigens that APC would present (Fig. 1). This is critical as the inability to determine what antigenic responses are actually responsible for ongoing autoimmunity is one of the great challenges currently facing T cell targeted approaches to antigen specific tolerance induction.

## 6. Potential challenges

While there is clear evidence that failure of central tolerance can drive autoimmunity [87,88] there is also a growing understanding that at least some forms of autoimmunity are not the result of rare failures in central tolerance of lymphocytes, but rather represent a failure in the control of autoreactive cells that typically occur in the normal immune repertoire [89–92]. Further, there are indications that adaptive immune auto-reactivity is involved in normal physiological responses, suggesting that autoimmune reactions could, at least in some cases, represent dysregulation of otherwise beneficial biology [93,94]. Further, lessons from transplantation suggest that blockade of specific costimulatory molecules can decrease the capacity of naturally occurring regulatory mechanisms to modulate ongoing inflammation [95,96]. This data is supported in multiple other model systems that have demonstrated that blocking antigen presentation can also block the initiation of homeostatic peripheral tolerance mechanisms [97–101], even leading to *de novo* autoimmunity [102,103]. Any attempts to modulate antigen specific APC would also have to consider the emerging role of B cells as immune regulatory cells, with the capacity to modulate ongoing responses either through the production of cytokines such as IL-10 or by direct modulation of T cell function [104].

Given this data, it seems clear that attempts to modulate immune responses would have to be careful to avoid interference with active homeostatic regulatory processes. However, given the potential of this approach targeting tissue specific antigens displayed by antigen presenting cells appears to represent an emerging field of interest.

## 7. Conflict of interest statement

The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## References

- [1] K. Dornmair, N. Goebels, H.U. Weltzien, H. Wekerle, R. Hohlfeld, T-cell-mediated autoimmunity: novel techniques to characterize autoreactive T-cell receptors, *Am. J. Pathol.* 163 (2003) 1215–1226.
- [2] J.A. Bluestone, H. Bour-Jordan, M. Cheng, M. Anderson, T cells in the control of organ-specific autoimmunity, *J Clin Invest* 125 (2015) 2250–2260.
- [3] R.M. Pearson, L.M. Casey, K.R. Hughes, S.D. Miller, L.D. Shea, In vivo reprogramming of immune cells: technologies for induction of antigen-specific tolerance, *Adv. Drug Deliv. Rev.* 114 (2017) 240–255.
- [4] A.K. Shakya, K.S. Nandakumar, Antigen-specific tolerization and targeted delivery as therapeutic strategies for autoimmune diseases, *Trends Biotechnol.* 36 (2018) 686–699.
- [5] P.R. Adair, Y.C. Kim, A.H. Zhang, J. Yoon, D.W. Scott, human tregs made antigen specific by gene modification: the power to treat autoimmunity and antidrug antibodies with precision, *Front. Immunol.* 8 (2017) 1117.
- [6] H. Offner, S. Sinha, C. Wang, G.G. Burrows, A.A. Vandenbark, Recombinant T cell receptor ligands: immunomodulatory, neuroprotective and neuroregenerative effects suggest application as therapy for multiple sclerosis, *Rev. Neurosci.* 19 (2008) 327–339.
- [7] H. Erlich, A.M. Valdes, J. Noble, J.A. Carlson, M. Varney, P. Concannon, J.C. Mychaleckyj, J.A. Todd, P. Bonella, A.L. Fear, E. Lavant, A. Louey, P. Moonsamy, Type 1 Diabetes Genetics, C., HLA DR-DQ haplotypes and genotypes and type 1 diabetes risk: analysis of the type 1 diabetes genetics consortium families, *Diabetes* 57 (2008) 1084–1092.
- [8] J.A. Noble, A. Martin, A.M. Valdes, J.A. Lane, A. Galgani, A. Petrone, R. Lorini, P. Pozzilli, R. Buzzetti, H.A. Erlich, Type 1 diabetes risk for human leukocyte antigen (HLA)-DR3 haplotypes depends on genotypic context: association of DPB1 and HLA class I loci among DR3- and DR4-matched Italian patients and controls, *Hum. Immunol.* 69 (2008) 291–300.
- [9] M.D. Varney, A.M. Valdes, J.A. Carlson, J.A. Noble, B.D. Tait, P. Bonella, E. Lavant, A.L. Fear, A. Louey, P. Moonsamy, J.C. Mychaleckyj, H. Erlich, Type 1 Diabetes Genetics, C., HLA DPA1, DPB1 alleles and haplotypes contribute to the risk associated with type 1 diabetes: analysis of the type 1 diabetes genetics consortium families, *Diabetes* 59 (2010) 2055–2062.
- [10] S. Sawcer, G. Hellenthal, M. Pirinen, C.C. Spencer, N.A. Patsopoulos, L. Moutsianas, A. Dilthey, Z. Su, C. Freeman, S.E. Hunt, S. Edkins, E. Gray, D.R. Booth, S.C. Potter, A. Goris, G. Band, A.B. Oturai, A. Strange, J. Saarela, C. Bellenguez, B. Fontaine, M. Gillman, B. Hemmer, R. Gwilliam, F. Zipp, A. Jayakumar, R. Martin, S. Leslie, S. Hawkins, E. Giannoulaitou, S. D'Alfonso, C. Berthele, M. Boggild, J.P. Bradfield, D. Brassat, R. Bergamaschi, D. Buck, H. Butzkueven, R. Capra, W.M. Carroll, P. Cavalla, E.G. Celius, S. Cepok, R. Chiavacci, F. Clerget-Darpoux, K. Clysters, G. Comi, M. Cossburn, I. Cournu-Rebeix, M.B. Cox, W. Cozen, B.A. Cree, A.H. Cross, D.M.J. Cusi Daly, E. Davis, P.I. de Bakker, M. Debouverie, M.B. D'Hooghe, K. Dixon, R. Dobosi, B. Dubois, D. Ellinghaus, I. Elovaara, F. Esposito, C. Fontenille, S. Foote, A. Franke, D. Galimberti, A. Ghezzi, J. Glessner, R. Gomez, O. Gout, C. Graham, S.F. Grant, F.R. Guerini, H. Hakonarson, P. Hall, A. Hamsten, H.P. Hartung, R.N. Heard, S. Heath, J. Hobart, M. Hoshi, C. Infante-Duarte, G. Ingram, W. Ingram, T. Islam, M. Jagodic, M. Kabesch, A.G. Kermod, T.J. Kilpatrick, C. Kim, N. Klopp, K. Koivisto, M. Larsson, M. Lathrop, J.S. Lechner-Scott, M.A. Leone, V. Leppa, U. Liljedahl, I.L. Bomfim, R.R. Lincoln, J. Link, J. Liu, A.R. Lorentzen, S. Lupoli, F. Macciardi, T. Mack, M. Marriotti, V. Martinelli, D. Mason, J.L. McCauley, F. Mentch, I.L. Mero, T. Mihalova, X. Montalban, J. Mottershead, K.M. Myhr, P. Naldi, W. Ollier, A. Page, A. Palotie, J. Pelletier, L. Piccio, T. Pickersgill, F. Piehl, S. Pobywajlo, H.L. Quach, P.P. Ramsay, M. Reunanen, R. Reynolds, J.D. Rioux, M. Rodegher, S. Roesner, J.P. Rubio, I.M. Ruckert, M. Salvetti, E. Salvi, A. Santaniello, C.A. Schaefer, S. Schreiber, C. Schulze, R.J. Scott, F. Sellebjerg, K.W. Selmaj, D. Sexton, L. Shen, B. Simms-Acuna, S. Skidmore, P.M. Sleiman, C. Smestad, P.S. Sorensen, H.B. Sondergaard, J. Stankovich, R.C. Strange, A.M. Sulonen, E. Sundqvist, A.C. Syvanen, F. Taddeo, B. Taylor, J.M. Blackwell, P. Tienari, E. Bramon, A. Tourbah, M.A. Brown, E. Tronczynska, J.P. Casas, N. Tubridy, A. Corvin, J. Vickery, J. Jankowski, P. Villoslada, H.S. Markus, K. Wang, C.G. Mathew, J. Wason, C.N. Palmer, H.E. Wichmann, R. Plomin, E. Willoughby, A. Rautanen, J. Winkelmann, M. Wittig, R.C. Trembath, J. Yauquan, A.C. Viswanathan, H. Zhang, N.W. Wood, R. Zuvich, P. Deloukas, C. Langford, A. Duncanson, J.R. Oksenberg, M.A. Pericak-Vance, J.L. Haines, T. Olsson, J. Hillert, A.J. Ivinson, P.L. De Jager, L. Peltonen, G.J. Stewart, D.A. Hafler, S.L. Hauser, G. McVean, P. Donnelly, A. Compston, Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis, *Nature* 476 (2011) 214–219.
- [11] F. Menconi, M.C. Monti, D.A. Greenberg, T. Oashi, R. Osman, T.F. Davies, Y. Ban, E.M. Jacobson, E.S. Concepcion, C.W. Li, Y. Tomer, Molecular amino acid signatures in the MHC class II peptide-binding pocket predispose to autoimmune thyroiditis in humans and in mice, *Proc. Natl. Acad. Sci. U.S.A.* 105 (2008) 14034–14039.
- [12] M.J. Simmonds, J.M. Howson, J.M. Heward, H.J. Cordell, H. Foxall, J. Carr-Smith, S.M. Gibson, N. Walker, Y. Tomer, J.A. Franklyn, J.A. Todd, S.C. Gough, Regression mapping of association between the human leukocyte antigen region and Graves disease, *Am. J. Hum. Genet.* 76 (2005) 157–163.
- [13] M.M. Fernando, C.R. Stevens, E.C. Walsh, P.L. De Jager, P. Goyette, R.M. Plenge, T.J. Vyse, J.D. Rioux, Defining the role of the MHC in autoimmunity: a review and pooled analysis, *PLoS Genet.* 4 (2008) e1000024.
- [14] P. Forabosco, E. Bouzigon, M.Y. Ng, J. Hermanowski, S.A. Fisher, L.A. Criswell, C.M. Lewis, Meta-analysis of genome-wide linkage studies across autoimmune diseases, *Eur. J. Hum. Genet.* 17 (2009) 236–243.
- [15] V. Matzaraki, V. Kumar, C. Wijmenga, A. Zernakova, The MHC locus and genetic susceptibility to autoimmune and infectious diseases, *Genome Biol.* 18 (2017) 76.
- [16] L. Moutsianas, L. Jostins, A.H. Beecham, A.T. Dilthey, D.K. Xifara, M. Ban, T.S. Shah, N.A. Patsopoulos, L. Alfredsson, C.A. Anderson, K.E. Attfield, S.E. Baranzini, J. Barrett, T.M.C. Binder, D. Buck, E.G. Celius, C. Cotsapas, S. D'Alfonso, C.A. Dendrou, P. Donnelly, B. Dubois, B. Fontaine, L. Fugger, A. Goris, P.A. Gourraud, C. Graetz, B. Hemmer, J. Hillert, I.B.D.G.C. International, I. Kockum, S. Leslie, C.M. Lill, F. Martinelli-Boneschi, J.R. Oksenberg, T. Olsson, A. Oturai, J. Saarela, H.B. Sondergaard, A. Spurkland, B. Taylor, J. Winkelmann, F. Zipp, J.L. Haines, M.A. Pericak-Vance, C.C.A. Spencer, G. Stewart, D.A. Hafler, A.J. Ivinson, H.F. Harbo, S.L. Hauser, P.L. De Jager, A. Compston, J.L. McCauley, S. Sawcer, G. McVean, Class II HLA interactions modulate genetic risk for multiple sclerosis, *Nat. Genet.* 47 (2015) 1107–1113.
- [17] D.M. Evans, C.C. Spencer, J.J. Pointon, Z. Su, D. Harvey, G. Kochan, U. Oppermann, A. Dilthey, M. Pirinen, M.A. Stone, L. Appleton, L. Moutsianas, S. Leslie, T. Wordsworth, T.J. Kenna, T. Karaderi, G.P. Thomas, M.M. Ward, M.H. Weisman, C. Farrar, L.A. Bradbury, P. Danoy, R.D. Inman, W. Maksymowich, D. Gladman, P. Rahman, Spondyloarthritis Research Consortium of, C., A. Morgan, H. Marzo-Ortega, P. Bowness, K. Gaffney, J.S. Gaston, M. Smith, J. Bruges-Armas, A.R. Couto, R. Sorrentino, F. Paladini,

- M.A. Ferreira, H. Xu, Y. Liu, L. Jiang, C. Lopez-Larrea, R. Diaz-Pena, A. Lopez-Vazquez, T. Zayats, G. Band, C. Bellenguez, H. Blackburn, J.M. Blackwell, E. Bramon, S.J. Bumpstead, J.P. Casas, A. Corvin, N. Craddock, P. Deloukas, S. Dronov, A. Duncanson, S. Edkins, C. Freeman, M. Gillman, E. Gray, R. Gwilliam, N. Hammond, S.E. Hunt, J. Jankowski, A. Jayakumar, C. Langford, J. Liddle, H.S. Markus, C.G. Mathew, O.T. McCann, M.I. McCarthy, C.N. Palmer, L. Peltonen, R. Plomin, S.C. Potter, A.R. Rautanen Ravindrarajah, M. Ricketts, N. Samani, S.J. Sawcer, A. Strange, R.C. Trembath, A.C. Viswanathan, M. Waller, P. Weston, P. Whittaker, S. Widaa, N.W. Wood, G. McVean, J.D. Reveille, B.P. Wordsworth, M.A. Brown, P. Donnelly, Australo-Anglo-American Spondyloarthritis, C., Wellcome Trust Case Control, C., Interaction between ERAP1 and HLA-B27 in ankylosing spondylitis implicates peptide handling in the mechanism for HLA-B27 in disease susceptibility, *Nat. Genet.* 43 (2011) 761–767.
- [18] A. Martin-Esteban, A. Sanz-Bravo, P. Guasp, E. Barnea, A. Admon, J.A. Lopez de Castro, Separate effects of the ankylosing spondylitis associated ERAP1 and ERAP2 aminopeptidases determine the influence of their combined phenotype on the HLA-B\*27 peptidome, *J. Autoimmun.* 79 (2017) 28–38.
- [19] S.A. Jagessar, I.R. Holtman, S. Hofman, E. Morandi, N. Heijmans, J.D. Laman, B. Gran, B.W. Faber, S.I. van Kasteren, B.J. Eggen, B.A. Hart, Lymphocryptovirus Infection of Nonhuman Primate B Cells Converts Destructive into Productive Processing of the Pathogenic CD8 T Cell Epitope in Myelin Oligodendrocyte Glycoprotein, *J. Immunol.* 197 (2016) 1074–1088.
- [20] U. Kuckelkorn, T. Ruppert, B. Strehl, P.R. Jungblut, U. Zimny-Arndt, S. Lamer, I. Prinz, I. Drung, P.M. Kloetzel, S.H. Kaufmann, U. Steinhoff, Link between organ-specific antigen processing by 20S proteasomes and CD8(+) T cell-mediated autoimmunity, *J. Exp. Med.* 195 (2002) 983–990.
- [21] S. Brocke, K. Gijbels, M. Allegretta, I. Ferber, C. Piercy, T. Blankenstein, R. Martin, U. Utz, N. Karin, D. Mitchell, T. Veromaa, A. Waisman, A. Gaur, P. Conlon, N. Ling, P.J. Fairchild, D.C. Wraith, A. O'Garra, C.G. Fathman, L. Steinman, Treatment of experimental encephalomyelitis with a peptide analogue of myelin basic protein, *Nature* 379 (1996) 343–346.
- [22] N. Karin, D.J. Mitchell, S. Brocke, N. Ling, L. Steinman, Reversal of experimental autoimmune encephalomyelitis by a soluble peptide variant of a myelin basic protein epitope: T cell receptor antagonism and reduction of interferon gamma and tumor necrosis factor alpha production, *J. Exp. Med.* 180 (1994) 2227–2237.
- [23] N.K. Koehler, C.Y. Yang, J. Varady, Y. Lu, X.W. Wu, M. Liu, D. Yin, M. Bartels, B.Y. Xu, P.P. Roller, Y.Q. Long, P. Li, M. Kattah, M.L. Cohn, K. Moran, E. Tilley, J.R. Richert, S. Wang, Structure-based discovery of nonpeptidic small organic compounds to block the T cell response to myelin basic protein, *J. Med. Chem.* 47 (2004) 4989–4997.
- [24] N. Ji, A. Somanaboina, A. Dixit, K. Kawamura, N.J. Hayward, C. Self, G.L. Olson, T.G. Forsthuber, Small molecule inhibitor of antigen binding and presentation by HLA-DR2b as a therapeutic strategy for the treatment of multiple sclerosis, *J. Immunol.* 191 (2013) 5074–5084.
- [25] R.M. Smith, A. Morgan, D.C. Wraith, Anti-class II MHC antibodies prevent and treat EAE without APC depletion, *Immunology* 83 (1994) 1–8.
- [26] M.K. Waldor, S. Sriram, H.O. McDevitt, L. Steinman, In vivo therapy with monoclonal anti-I-A antibody suppresses immune responses to acetylcholine receptor, *PNAS* 80 (1983) 2713–2717.
- [27] M.K. Waldor, M. O'Hearn, S. Sriram, L. Steinman, Treatment of experimental autoimmune myasthenia gravis with monoclonal antibodies to immune response gene products, *Ann. N. Y. Acad. Sci.* 505 (1987) 655–668.
- [28] L. Steinman, J.T. Rosenbaum, S. Sriram, H.O. McDevitt, Prevention of EAE with monoclonal anti-I-A antibodies, *Trans Am. Neurol. Assoc.* 106 (1981) 237–238.
- [29] L. Steinman, J.T. Rosenbaum, S. Sriram, H.O. McDevitt, In vivo effects of antibodies to immune response gene products: prevention of experimental allergic encephalitis, *Proc. Natl. Acad. Sci. U.S.A.* 78 (1981) 7111–7114.
- [30] B.H. Hahn, F.M. Ebling, Suppression of murine lupus nephritis by administration of an anti-idiotypic antibody to anti-DNA, *J. Immunol.* 132 (1984) 187–190.
- [31] D. Teitelbaum, J. Rauch, B.D. Stollar, R.S. Schwartz, In vivo effects of antibodies against a high frequency idiotypic of anti-DNA antibodies in MRL mice, *J. Immunol.* 132 (1984) 1282–1285.
- [32] J.A. Carrero, D.P. McCarthy, S.T. Ferris, X. Wan, H. Hu, B.H. Zinselmeyer, A.N. Vomund, E.R. Unanue, Resident macrophages of pancreatic islets have a seminal role in the initiation of autoimmune diabetes of NOD mice, *Proc. Natl. Acad. Sci. U.S.A.* 114 (2017) E10418–E10427.
- [33] S.T. Ferris, J.A. Carrero, J.F. Mohan, K. Calderon, K.M. Murphy, E.R. Unanue, A minor subset of Batf3-dependent antigen-presenting cells in islets of Langerhans is essential for the development of autoimmune diabetes, *Immunity* 41 (2014) 657–669.
- [34] W. Yi, N.P. Seth, T. Martillotti, K.W. Wucherpfennig, D.B. Sant'Angelo, L.K. Denzin, Targeted regulation of self-peptide presentation prevents type 1 diabetes in mice without disrupting general immunocompetence, *J Clin Invest* 120 (2010) 1324–1336.
- [35] A. Geluk, K.E. van Meijgaarden, B.O. Roep, T.H. Ottenhoff, Altered peptide ligands of islet autoantigen Imogen 38 inhibit antigen specific T cell reactivity in human type-1 diabetes, *J. Autoimmun.* 11 (1998) 353–361.
- [36] A.W. Michels, D.A. Ostrov, L. Zhang, M. Nakayama, M. Fuse, K. McDaniel, B.O. Roep, P.A. Gottlieb, M.A. Atkinson, G.S. Eisenbarth, Structure-based selection of small molecules to alter allele-specific MHC class II antigen presentation, *J. Immunol.* 187 (2011) 5921–5930.
- [37] C.W. Li, F. Menconi, R. Osman, M. Mezei, E.M. Jacobson, E. Concepcion, C.S. David, D.B. Kastrinsky, M. Ohlmeyer, Y. Tomer, Identifying a small molecule blocking antigen presentation in autoimmune thyroiditis, *J. Biol. Chem.* 291 (2016) 4079–4090.
- [38] B. Becher, I. Bechmann, M. Greter, Antigen presentation in autoimmunity and CNS inflammation: how T lymphocytes recognize the brain, *J Mol Med (Berl)* 84 (2006) 532–543.
- [39] M. Greter, F.L. Heppner, M.P. Lemos, B.M. Odermatt, N. Goebels, T. Laufer, R.J. Noelle, B. Becher, Dendritic cells permit immune invasion of the CNS in an animal model of multiple sclerosis, *Nat. Med.* 11 (2005) 328–334.
- [40] P. Kivisakk, J. Imitola, S. Rasmussen, W. Elyaman, B. Zhu, R.M. Ransohoff, S.J. Khoury, Localizing central nervous system immune surveillance: meningeal antigen-presenting cells activate T cells during experimental autoimmune encephalomyelitis, *Ann. Neurol.* 65 (2009) 457–469.
- [41] M.L. Krakowski, T. Owens, Naive T lymphocytes traffic to inflamed central nervous system, but require antigen recognition for activation, *Eur. J. Immunol.* 30 (2000) 1002–1009.
- [42] S.D. Miller, E.J. McMahon, B. Schreiner, S.L. Bailey, Antigen presentation in the CNS by myeloid dendritic cells drives progression of relapsing experimental autoimmune encephalomyelitis, *Ann. N. Y. Acad. Sci.* 1103 (2007) 179–191.
- [43] Z. Liu, B. Li, X. Li, L. Zhang, L. Lai, Identification of small-molecule inhibitors against human leukocyte antigen-death receptor 4 (HLA-DR4) through a comprehensive strategy, *J. Chem. Inf. Model* 51 (2011) 326–334.
- [44] M. Fridkis-Hareli, Design of peptide immunotherapies for MHC Class-II-associated autoimmune disorders, *Clin. Dev. Immunol.* 2013 (2013) 826191.
- [45] R.A. Sosa, C. Murphey, N. Ji, A.E. Cardona, T.G. Forsthuber, The kinetics of myelin antigen uptake by myeloid cells in the central nervous system during experimental autoimmune encephalomyelitis, *J. Immunol.* 191 (2013) 5848–5857.
- [46] P.V. Lehmann, T. Forsthuber, A. Miller, E.E. Sercarz, Spreading of T-cell autoimmunity to cryptic determinants of an autoantigen, *Nature* 358 (1992) 155–157.
- [47] S.H. Venkatesha, M. Durai, K.D. Moudgil, Epitope Spreading in Autoimmune Diseases. *Infection and Autoimmunity*, second ed., 2015, pp. 45–68.
- [48] J. Greenwood, L. Steinman, S.S. Zamvil, Statin therapy and autoimmune disease: from protein prenylation to immunomodulation, *Nat. Rev. Immunol.* 6 (2006) 358–370.
- [49] C. Monaco, J. Nanchahal, P. Taylor, M. Feldmann, Anti-TNF therapy: past, present and future, *Int. Immunol.* 27 (2015) 55–62.
- [50] K.D. Moudgil, D. Choubey, Cytokines in autoimmunity: role in induction, regulation, and treatment, *J. Interferon Cytokine Res.* 31 (2011) 695–703.
- [51] A.W. van Lieshout, P. Barrera, R.L. Smeets, G.J. Pesman, P.L. van Riel, W.B. van den Berg, T.R. Radstake, Inhibition of TNF alpha during maturation of dendritic cells results in the development of semi-mature cells: a potential mechanism for the beneficial effects of TNF alpha blockade in rheumatoid arthritis, *Ann. Rheum. Dis.* 64 (2005) 408–414.
- [52] P. Musette, J.D. Bouaziz, B cell modulation strategies in autoimmune diseases: new concepts, *Front. Immunol.* 9 (2018) 622.
- [53] M. Zouali, G. Tsay, Developing connections amongst B lymphocytes and deregulated pathways in autoimmunity, *Mol. Med.* 22 (2016) 705–712.
- [54] G.J. Tsay, M. Zouali, The interplay between innate-like B cells and other cell types in autoimmunity, *Front. Immunol.* 9 (2018) 1064.
- [55] K. Hofmann, A.K. Clauser, R.A. Manz, Targeting B cells and plasma cells in autoimmune diseases, *Front. Immunol.* 9 (2018) 835.
- [56] K. Lehmann-Horn, S. Kinzel, M.S. Weber, Deciphering the role of B cells in multiple sclerosis-towards specific targeting of pathogenic function, *Int. J. Mol. Sci.* 18 (2017).
- [57] S. Kinzel, M.S. Weber, B cell-directed therapeutics in multiple sclerosis: rationale and clinical evidence, *CNS Drugs* 30 (2016) 1137–1148.
- [58] E. Marino, M. Batten, J. Groom, S. Walters, D. Liuwantara, F. Mackay, S.T. Grey, Marginal-zone B-cells of nonobese diabetic mice expand with diabetes onset, invade the pancreatic lymph nodes, and present autoantigen to diabetogenic T-cells, *Diabetes* 57 (2008) 395–404.
- [59] E. Marino, J. Villanueva, S. Walters, D. Liuwantara, F. Mackay, S.T. Grey, CD4(+)CD25(+) T-cells control autoimmunity in the absence of B-cells, *Diabetes* 58 (2009) 1568–1577.
- [60] Y. Xiu, C.P. Wong, J.D. Bouaziz, Y. Hamaguchi, Y. Wang, S.M. Pop, R.M. Tisch, T.F. Tedder, B lymphocyte depletion by CD20 monoclonal antibody prevents diabetes in nonobese diabetic mice despite isotype-specific differences in Fc gamma R effector functions, *J. Immunol.* 180 (2008) 2863–2875.
- [61] G. Zekavat, S.Y. Rostami, A. Badkerhanian, R.F. Parsons, B. Koeberlein, M. Yu, C.D. Ward, T.S. Migone, L. Yu, G.S. Eisenbarth, M.P. Cancro, A. Naji, H. Noorchashm, In vivo BlyS/BAFF neutralization ameliorates islet-directed autoimmunity in nonobese diabetic mice, *J. Immunol.* 181 (2008) 8133–8144.
- [62] P. Fiorina, A. Vergani, S. Dada, M. Jurewicz, M. Wong, K. Law, E. Wu, Z. Tian, R. Abdi, I. Guleria, S. Rodig, K. Dunussi-Joannopoulos, J. Bluestone, M.H. Sayegh, Targeting CD22 reprograms B-cells and reverses autoimmune diabetes, *Diabetes* 57 (2008) 3013–3024.
- [63] C.Y. Hu, D. Rodriguez-Pinto, W. Du, A. Ahuja, O. Henegariu, F.S. Wong, M.J. Shlomchik, L. Wen, Treatment with CD20-specific antibody prevents and reverses autoimmune diabetes in mice, *J. Clin. Invest.* 117 (2007) 3857–3867.
- [64] X. Montalban, S.L. Hauser, L. Kappos, D.L. Arnold, A. Bar-Or, G. Comi, J. de Seze, G. Giovannoni, H.P. Hartung, B. Hemmer, F. Lublin, K.W. Rammohan, K. Selmaj, A. Traboulsee, A. Sauter, D. Masterman, P. Fontoura, S. Belachew, H. Garren, N. Mairon, P. Chin, J.S. Wolinsky, O.C. Investigators, Ocrelizumab versus placebo in primary progressive multiple sclerosis, *N. Engl. J. Med.* 376 (2017) 209–220.
- [65] S.L. Hauser, A. Bar-Or, G. Comi, G. Giovannoni, H.P. Hartung, B. Hemmer, F. Lublin, X. Montalban, K.W. Rammohan, K. Selmaj, A. Traboulsee, J.S. Wolinsky, D.L. Arnold, G. Klingelschmitt, D. Masterman, P. Fontoura, S. Belachew, P. Chin, N. Mairon, H. Garren, L. Kappos, I. Opera, O.I.C. Investigators, Ocrelizumab versus interferon beta-1a in relapsing multiple sclerosis, *N. Engl. J. Med.* 376 (2017) 221–234.
- [66] J.A. Lyons, M. San, M.P. Happ, A.H. Cross, B cells are critical to induction of

- experimental allergic encephalomyelitis by protein but not by a short encephalitogenic peptide, *Eur. J. Immunol.* 29 (1999) 3432–3439.
- [67] J.A. Lyons, M.J. Ramsbottom, A.H. Cross, Critical role of antigen-specific antibody in experimental autoimmune encephalomyelitis induced by recombinant myelin oligodendrocyte glycoprotein, *Eur. J. Immunol.* 32 (2002) 1905–1913.
- [68] C.R. Parker Harp, A.S. Archambault, J. Sim, S.T. Ferris, R.J. Mikesell, P.A. Koni, M. Shimoda, C. Linington, J.H. Russell, G.F. Wu, B cell antigen presentation is sufficient to drive neuroinflammation in an animal model of multiple sclerosis, *J. Immunol.* 194 (2015) 5077–5084.
- [69] C.R. Parker Harp, A.S. Archambault, J. Sim, M.J. Shlomchik, J.H. Russell, G.F. Wu, B cells are capable of independently eliciting rapid reactivation of encephalitogenic CD4 T cells in a murine model of multiple sclerosis, *PLoS ONE* 13 (2018) e0199694.
- [70] J.A. Spanier, D.R. Frederick, J.J. Taylor, J.R. Heffernan, D.I. Kotov, T. Martinov, K.C. Osum, J.L. Ruggiero, B.J. Rust, S.J. Landry, M.K. Jenkins, J.B. McLachlan, B.T. Fife, Efficient generation of monoclonal antibodies against peptide in the context of MHCII using magnetic enrichment, *Nat. Commun.* 7 (2016) 11804.
- [71] D.L. Porter, B.L. Levine, M. Kalos, A. Bagg, C.H. June, Chimeric antigen receptor-modified T cells in chronic lymphoid leukemia, *N. Engl. J. Med.* 365 (2011) 725–733.
- [72] Y.C. Kim, A.H. Zhang, J. Yoon, W.E. Culp, J.R. Lees, K.W. Wucherpfennig, D.W. Scott, Engineered MBP-specific human Tregs ameliorate MOG-induced EAE through IL-2-triggered inhibition of effector T cells, *J. Autoimmun.* (2018).
- [73] A. Chhabra, L. Yang, P. Wang, B. Comin-Anduix, R. Das, N.G. Chakraborty, S. Ray, S. Mehrotra, H. Yang, C.L. Hardee, R. Hollis, D.I. Dorsky, R. Koya, D.B. Kohn, A. Ribas, J.S. Economou, D. Baltimore, B. Mukherji, CD4+CD25- T cells transduced to express MHC class I-restricted epitope-specific TCR synthesize Th1 cytokines and exhibit MHC class I-restricted cytolytic effector function in a human melanoma model, *J. Immunol.* 181 (2008) 1063–1070.
- [74] T.L. Frankel, W.R. Burns, P.D. Peng, Z. Yu, D. Chinnasamy, J.A. Wargo, Z. Zheng, N.P. Restifo, S.A. Rosenberg, R.A. Morgan, Both CD4 and CD8 T cells mediate equally effective in vivo tumor treatment when engineered with a highly avid TCR targeting tyrosinase, *J. Immunol.* 184 (2010) 5988–5998.
- [75] S. Ghorashian, P. Velica, I. Chua, A.M. McNicol, B. Carpenter, A. Holler, E. Nicholson, M. Ahmadi, M. Zech, S.A. Xue, W. Uckert, E. Morris, R. Chakraverty, H.J. Stauss, CD8 T cell tolerance to a tumor-associated self-antigen is reversed by CD4 T cells engineered to express the same T cell receptor, *J. Immunol.* 194 (2015) 1080–1089.
- [76] E.L. Pearce, D.J. Shedlock, H. Shen, Functional characterization of MHC class II-restricted CD8+CD4- and CD8-CD4- T cell responses to infection in CD4-/- mice, *J. Immunol.* 173 (2004) 2494–2499.
- [77] S.G. Hansen, J.B. Sacha, C.M. Hughes, J.C. Ford, B.J. Burwitz, I. Scholz, R.M. Gilbride, M.S. Lewis, A.N. Gilliam, A.B. Ventura, D. Malouli, G. Xu, R. Richards, N. Whizin, J.S. Reed, K.B. Hammond, M. Fischer, J.M. Turner, A.W. Legasse, M.K. Axthelm, P.T. Edlfsen, J.A. Nelson, J.D. Lifson, K. Fruh, L.J. Picker, Cytomegalovirus vectors violate CD8+ T cell epitope recognition paradigms, *Science* 340 (2013) 1237874.
- [78] P.D. Holler, D.M. Kranz, Quantitative analysis of the contribution of TCR/pepMHC affinity and CD8 to T cell activation, *Immunity* 18 (2003) 255–264.
- [79] G. Guarda, M. Hons, S.F. Soriano, A.Y. Huang, R. Polley, A. Martin-Fonoteca, J.V. Stein, R.N. Germain, A. Lanzavecchia, F. Sallusto, L-selectin-negative CCR7-effector and memory CD8+ T cells enter reactive lymph nodes and kill dendritic cells, *Nat. Immunol.* 8 (2007) 743–752.
- [80] D.A. Price, J.M. Brenchley, L.E. Ruff, M.R. Betts, B.J. Hill, M. Roederer, R.A. Koup, S.A. Migueles, E. Gostick, L. Wooldridge, A.K. Sewell, M. Connors, D.C. Douek, Avidity for antigen shapes clonal dominance in CD8+ T cell populations specific for persistent DNA viruses, *J. Exp. Med.* 202 (2005) 1349–1361.
- [81] D.J. Zammit, L.S. Cauley, Q.M. Pham, L. Lefrancois, Dendritic cells maximize the memory CD8 T cell response to infection, *Immunity* 22 (2005) 561–570.
- [82] K.A. Andrew, H.M. Simkins, S. Witzel, R. Perret, J. Hudson, I.F. Hermans, D.S. Ritchie, J. Yang, F. Ronchese, Dendritic cells treated with lipopolysaccharide up-regulate serine protease inhibitor 6 and remain sensitive to killing by cytotoxic T lymphocytes in vivo, *J. Immunol.* 181 (2008) 8356–8362.
- [83] J. Yang, S.P. Huck, R.S. McHugh, I.F. Hermans, F. Ronchese, Perforin-dependent elimination of dendritic cells regulates the expansion of antigen-specific CD8+ T cells in vivo, *PNAS* 103 (2006) 147–152.
- [84] S. Laffont, J.D. Coudert, L. Garidou, L. Delpy, A. Wiedemann, C. Demur, C. Coureau, J.C. Guery, CD8+ T-cell-mediated killing of donor dendritic cells prevents alloreactive T helper type-2 responses in vivo, *Blood* 108 (2006) 2257–2264.
- [85] B. Ludewig, W.V. Bonilla, T. Dumrese, B. Odermatt, R.M. Zinkernagel, H. Hengartner, Perforin-independent regulation of dendritic cell homeostasis by CD8(+) T cells in vivo: implications for adaptive immunotherapy, *Eur. J. Immunol.* 31 (2001) 1772–1779.
- [86] P. Wong, E.G. Pamer, Feedback regulation of pathogen-specific T cell priming, *Immunity* 18 (2003) 499–511.
- [87] Y. Ito, M. Hashimoto, K. Hirota, N. Ohkura, H. Morikawa, H. Nishikawa, A. Tanaka, M. Furu, H. Ito, T. Fujii, T. Nomura, S. Yamazaki, A. Morita, D.A. Vignali, J.W. Kappler, S. Matsuda, T. Mimori, N. Sakaguchi, S. Sakaguchi, Detection of T cell responses to a ubiquitous cellular protein in autoimmune disease, *Science* 346 (2014) 363–368.
- [88] M.S. Anderson, E.S. Venanzi, L. Klein, Z. Chen, S.P. Berzins, S.J. Turley, H. von Boehmer, R. Bronson, A. Dierich, C. Benoist, D. Mathis, Projection of an immunological self shadow within the thymus by the aire protein, *Science* 298 (2002) 1395–1401.
- [89] H. Wardemann, S. Yurasov, A. Schaefer, J.W. Young, E. Meffre, M.C. Nussenzweig, Predominant autoantibody production by early human B cell precursors, *Science* 301 (2003) 1374–1377.
- [90] A. Van der Aa, N. Hellings, C.C. Bernard, J. Raus, P. Stinissen, Functional properties of myelin oligodendrocyte glycoprotein-reactive T cells in multiple sclerosis patients and controls, *J. Neuroimmunol.* 137 (2003) 164–176.
- [91] M.P. Crawford, S.X. Yan, S.B. Ortega, R.S. Mehta, R.E. Hewitt, D.A. Price, P. Stastny, D.C. Douek, R.A. Koup, M.K. Racke, N.J. Karandikar, High prevalence of autoreactive, neuroantigen-specific CD8+ T cells in multiple sclerosis revealed by novel flow cytometric assay, *Blood* 103 (2004) 4222–4231.
- [92] W. Yu, N. Jiang, P.J. Ebert, B.A. Kidd, S. Muller, P.J. Lund, J. Juang, K. Adachi, T. Tse, M.E. Birnbaum, E.W. Newell, D.M. Wilson, G.M. Grotenbreg, S. Valitutti, S.R. Quake, M.M. Davis, Clonal deletion prunes but does not eliminate self-specific alpha beta CD8(+) T lymphocytes, *Immunity* 42 (2015) 929–941.
- [93] K. Baruch, M. Schwartz, CNS-specific T cells shape brain function via the choroid plexus, *Brain Behav. Immun.* 34 (2013) 11–16.
- [94] M. Schwartz, K. Baruch, Breaking peripheral immune tolerance to CNS antigens in neurodegenerative diseases: boosting autoimmunity to fight-off chronic neuroinflammation, *J. Autoimmun.* 54 (2014) 8–14.
- [95] J.F. Bach, Immunotherapy of type 1 diabetes: lessons for other autoimmune diseases, *Arthritis Res.* 4 (Suppl. 3) (2002) S3–S15.
- [96] J.F. Bach, L. Chatenoud, A historical view from thirty eventful years of immunotherapy in autoimmune diabetes, *Semin. Immunol.* 23 (2011) 174–181.
- [97] J.G. Henderson, A. Opejin, A. Jones, C. Gross, D. Hawiger, CD5 instructs extrathymic regulatory T cell development in response to self and tolerizing antigens, *Immunity* 42 (2015) 471–483.
- [98] A. Jones, A. Opejin, J.G. Henderson, C. Gross, R. Jain, J.A. Epstein, R.A. Flavell, D. Hawiger, Peripherally induced tolerance depends on peripheral regulatory T cells that require Hoxp to inhibit intrinsic IL-2 expression, *J. Immunol.* 195 (2015) 1489–1497.
- [99] C.A. Iberg, A. Jones, D. Hawiger, Dendritic cells as inducers of peripheral tolerance, *Trends Immunol.* 38 (2017) 793–804.
- [100] A. Jones, D. Hawiger, Peripherally induced regulatory T Cells: recruited protectors of the central nervous system against autoimmune neuroinflammation, *Front. Immunol.* 8 (2017) 532.
- [101] A.K. Hopp, A. Rupp, V. Lukacs-Kornek, Self-antigen presentation by dendritic cells in autoimmunity, *Front. Immunol.* 5 (2014) 55.
- [102] M. Takenaka, V. Tiriveedhi, V. Subramanian, K. Hoshinaga, A.G. Patterson, T. Mohanakumar, Antibodies to MHC class II molecules induce autoimmunity: critical role for macrophages in the immunopathogenesis of obliterative airway disease, *PLoS ONE* 7 (2012) e42370.
- [103] M. Takenaka, V. Subramanian, V. Tiriveedhi, D. Phelan, R. Hachem, E. Trulock, A.E. Gelman, G.A. Patterson, K. Hoshinaga, T. Mohanakumar, Complement activation is not required for obliterative airway disease induced by antibodies to major histocompatibility complex class I: implications for chronic lung rejection, *J. Heart Lung Transplant.* 31 (2012) 1214–1222.
- [104] E.C. Rosser, C. Mauri, Regulatory B cells: origin, phenotype, and function, *Immunity* 42 (2015) 607–612.