



# Update on gastric cancer treatments and gene therapies

Alessio Biagioni<sup>1</sup> · Ileana Skalamera<sup>2</sup> · Sara Peri<sup>1,3</sup> · Nicola Schiavone<sup>1</sup> · Fabio Cianchi<sup>2</sup> · Elisa Giommoni<sup>4</sup> · Lucia Magnelli<sup>1</sup> · Laura Papucci<sup>1</sup>

Published online: 5 September 2019  
© Springer Science+Business Media, LLC, part of Springer Nature 2019

## Abstract

Gastric cancer is an active topic of clinical and basic research due to high morbidity and mortality. To date, gastrectomy and chemotherapy are the only therapeutic options for gastric cancer patients, but drug resistance, either acquired or primary, is the main cause for treatment failure. Differences in development and response to cancer treatments have been observed among ethnically diverse GC patient populations. In spite of major incidence, GC Asian patients have a significantly better prognosis and response to treatments than Caucasian ones due to genetic discordances between the two populations. Gene therapy could be an alternative strategy to overcome such issues and especially CRISPR/Cas9 represents one of the most intriguing gene-editing system. Thus, in this review article, we want to provide an update on the currently used therapies for the treatment of advanced GC.

**Keywords** Gastric cancer · Chemotherapy · Metastases · Gene therapy · CRISPR

## 1 Introduction

Gastric cancer (GC) is actually the fifth most frequently diagnosed cancer and the third leading cause of cancer death according to GLOBOCAN 2018, demonstrating to be a considerable welfare problem especially in Western Asian countries [1]. Being, the main risk factors the *Helicobacter pylori* infection [2], an unbalanced diet [3], alcohol consumption, and

tobacco smoking [4], we might be commonly inclined to identify GC as an environmental-induced disease, underestimating the role of the genetic components. The majority of GC are classified as adenocarcinomas (80–90%) [5]. While the WHO classified GCs into several subgroups including papillary, tubular, mucinous, signet-ring cell, and mixed carcinoma, the most common Laurén classification is composed by only intestinal, diffuse, and indeterminate type [6]. Thanks to the advances in the diagnostic tools and the deeper knowledge of the genetic profile of cancer, we may currently approach cancers, and in particular GCs, from a molecular point of view. Many different signaling pathways, such as Wnt [7] or miR-126 [8], just to report two of them, have been demonstrated to be aberrant regulated in GCs. Although surgical resection is currently the best method to treat gastric cancer, chemotherapy (CT) is still used as elective therapy, especially when surgery is not possible or in presence of metastatic lesions. However, the most common phenomenon responsible for the treatment failure is the acquisition of resistance to chemotherapy agents. The emerging genome editing technologies, ever more precise and quicker to perform, enable the design of new targeted cancer therapies, being CRISPR the spearhead among them. CRISPR allows for effective gene knockout and gene modifications aimed to repair defective oncogenic pathways, albeit with low *in vivo* efficiency. Better results were obtained in cancer immunotherapy where

---

Lucia Magnelli and Laura Papucci equally contributed to this manuscript as last authors

---

✉ Nicola Schiavone  
nicola.schiavone@unifi.it

✉ Fabio Cianchi  
fabio.cianchi@unifi.it

- <sup>1</sup> Department of Experimental and Clinical Biomedical Sciences, University of Florence, Viale G.B. Morgagni 50, 50134 Florence, Italy
- <sup>2</sup> Department of Experimental and Clinical Medicine, University of Florence, Largo Brambilla 3, 50134 Florence, Italy
- <sup>3</sup> Department of Biotechnology, Chemistry and Pharmacy, University of Siena, Via Aldo Moro 2, 53100 Siena, Italy
- <sup>4</sup> Medical Oncology Unit, Department of Oncology and Robotic Surgery, AOU Careggi, Largo Brambilla 3, 50134 Florence, Italy

cancer-specific T cells can be engineered in order to be potentiated as anticancer weapons [9]. In this context, GC does not make an exception [10]. Indeed, somatic cells have to overcome a number of obstacles and acquire additional abilities to become tumor cells, among these, the ability to avoid immunogenicity that occasionally arise as a result of DNA rearrangement. Many immunotherapy methods can be used to potentiate immunosurveillance or avoid the immunoescape of GC. The strategies comprise adoptive therapy *via* Chimeric Antigen Receptor T Cells (CAR-T) using engineered lymphocytes from patient expressing chimeric receptors toward tumor antigens, immune checkpoint inhibitors aimed to interfere with negative regulatory pathways of the immune response, and cancer vaccines aimed at inducing APCs, like dendritic cells, to present tumor antigens. Also, some approaches of immunotherapy could be considered an indirect gene therapy that benefit from DNA editing technologies and high throughput transcriptome analysis albeit its efficacy in solid tumors is waiting to be clearly demonstrated [11]. In particular, CRISPR/Cas9 gene editing could be applied in T cell-based immunotherapy in order to generate CAR-T lymphocytes [12]. In GC research and therapy, many, if not all the immunotherapy-based approaches are being under scrutiny, from adoptive immunotherapy using CAR-T, to cancer vaccines (for a comprehensive review about immunotherapy in GC, see [13]). Based on the above considerations, we can envision a future where highly efficient gene-editing approaches will be pivotal in gastric cancer therapy. Another promising field is the regaining of tumor suppressor function, silenced by epigenetic modifications. Artificial transcription factors engineered with transcriptional repressor or activator molecules have been developed and demonstrated to effectively activate and repress gene expression in mammalian cells [14]. Tumor suppressor gene classes such as MASPIN and REPRIMO have been reactivated in GC cell lines using these tools in combination with CRISPR, furnishing a proof of concept to exploit such technologies [15]. Studies on GC at a molecular level provide ever increasing opportunities for therapeutic intervention on GC by gene therapy. An example comes from the long noncoding RNAs (lncRNAs): PANDAR, a lncRNA responsible for blocking the expression of a cyclin-dependent kinase inhibitor (CDKN1A) by competitively binding to p53, was knocked out by CRISPR/Cas9 technology determining GC growth inhibition *in vivo* [16]. In this review article, we report an overview on the state of art of GC therapies.

## 2 Chemotherapy treatments for advanced gastric Cancer

In Western countries, about 50% of patients present with locally advanced or metastatic gastric cancer (AGC) at

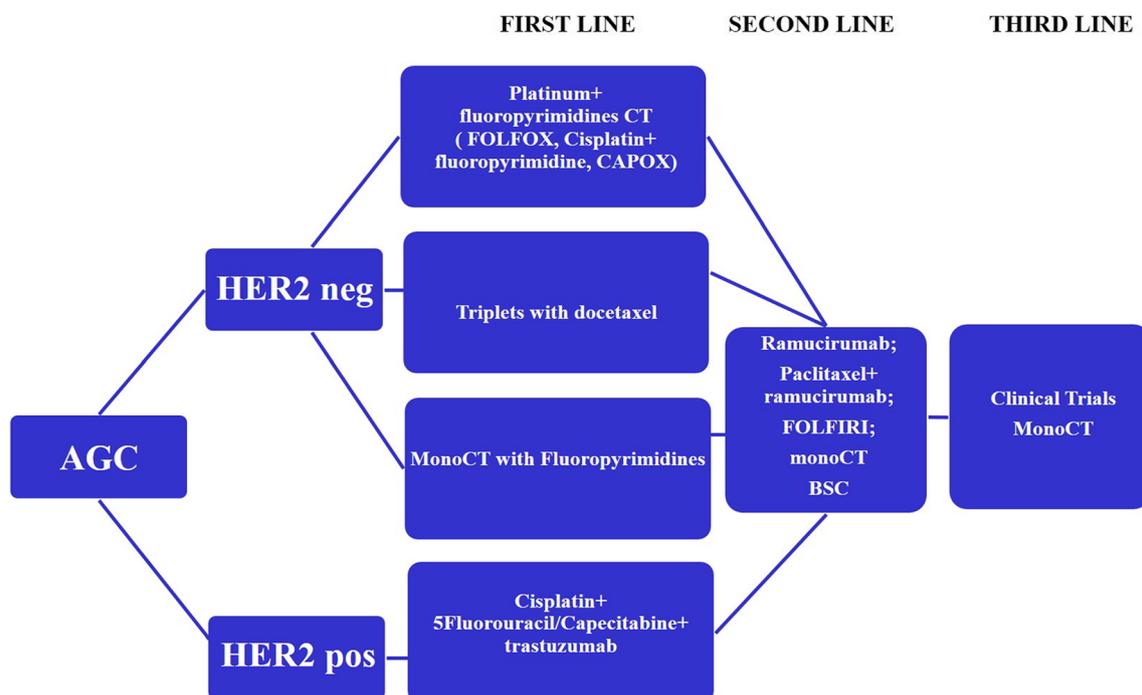
diagnosis, while about 40–60% of those radically resected relapses after surgery. In patients with AGC, palliative systemic treatment is the gold standard, with a median survival uncommonly exceeding 12 months. CT demonstrated to prolong survival and improve quality of life compared with best supportive care. However, median overall survival (OS) does not exceed 9–11 months with a 5-year OS of about 5–10% [17]. In patients with human epidermal growth factor receptor 2 (HER2) positive AGC, the addition of the anti-HER2 monoclonal antibody trastuzumab to CT, as demonstrated by the ToGA trial, achieved a survival benefit over CT alone (mOS 13.8 *vs.* 11.1 months, HR 0.74; 95% CI 0.60–0.91;  $p = 0.0046$ ), but the treatment is possible only in a limited subset of patients (15–20%) [18]. After the positive experience of the ToGA trial, first-line treatment with other targeted therapies failed to improve the outcome of AGC, while in second line, anti-VEGFR agent ramucirumab obtained a gain in terms of OS when used in monotherapy with respect to the best supportive care and in addition to CT *versus* CT alone. For first-line approach, international guidelines recommend regimens containing a platinum agent (cisplatin or oxaliplatin) and fluoropyrimidines (5-fluorouracil or capecitabine) in patients with HER2 negative disease. In case of HER2 positivity, trastuzumab is added to CT with cisplatin and fluoropyrimidines. In Eastern countries, S-1 (tegafur-gimstat-otastat potassium), an oral fluoropyrimidine, is used as mono CT or in combination, based on favorable results in trials involving Asiatic patients [19–21]. The addition of taxanes to first-approach CT, as in DCF (docetaxel, cisplatin and 5-fluorouracil) or FLOT (5-fluorouracil, oxaliplatin, and docetaxel) schedules resulted in higher response rates (RR) without impact on OS, but with relevant rate of toxicities: so triplets are reserved to young selected patients with good performance status to whom the goal of treatment is the downstaging (i.e., the neoadjuvant approach in limited disease, bulky and symptomatic disease, oligometastatic AGC) [22, 23]. Globally, after progression of disease to first-line CT, only about 40% of patients with AGC in Europe are able to receive second-line therapy [24]. Until the results with ramucirumab were presented, the standard of care for second line was mono CT, as confirmed by a meta-analysis of 10 randomized trials that demonstrated that doublet CT does not improve OS *versus* monotherapy, at the price of more toxicities [25]. In the REGARD trial, Ramucirumab *versus* placebo obtained an improvement in both OS (5.2 *vs.* 3.8 months, HR = 0.776,  $p = 0.047$ ) and progression-free survival PFS (2.1 *vs.* 1.3 months, HR = 0.48,  $p < 0.0001$ ). The efficacy of ramucirumab alone was comparable to the results of other phase III trials of second-line CT, with favorable toxicity profile. In the RAINBOW trial, the addition of ramucirumab to weekly paclitaxel significantly prolonged m OS (9.6 *vs.* 7.4 months, HR = 0.807,  $p = 0.017$ ) and PFS (4.4 *vs.* 2.9 months, HR = 0.635,  $p < 0.0001$ ) when compared with

paclitaxel monotherapy [26, 27]. When not contraindicated, ramucirumab is recommended by major International guidelines as second-line treatment as mono-CT or in combination with paclitaxel. Third-line treatment is not currently sustained by phase III evidences, so is reserved to selected patients. Trials with immunotherapy are ongoing in this setting of disease. It is also important to underline that nutritional support is a fundamental part of active treatment for patients with AGC, because an impaired nutritional status is associated with increased morbidity and mortality during anticancer treatment with major toxicities, dose reductions, delays, or treatment discontinuation. With a good support, we are able to increase the proportion of patients receiving second- and third-line treatments. We report in Fig. 1 the diagram of the therapeutic algorithm for AGC in Western countries.

### 3 Molecular insight for metastatic lesions

Metastatic GC is usually associated with incurable conditions. When anticancer therapies for primitive cancer other than surgery fail, interest in molecular mechanism related to metastasis development falls. This is the case of GC advanced disease, where CT can only slightly affect the patient survival. Indeed, until recently, GC databases reported incomplete information about GC spreading, restricting the field to Y or N, regardless of primary and secondary metastasization sites. An

extensive study about GC spreading performed on Swedish registers reports that the most common distant metastasization sites are the liver (48%), lung (15%), and bones (12%), while proximal peritoneal dissemination represents 32% [28]. Anyway, distribution of GC metastases sites can vary in different stratification GC patients' studies, probably depending on the cohort considered and heterogeneity in data acquisition. For example, peritoneal localization of GC cells by direct invasion from the gastric wall is elsewhere reported to be the most recurring, in comparison to distant metastatic settlements [29, 30]. Recently, whole-genome sequencing techniques that allowed to identify molecular subtypes in primitive gastric cancer cohorts have been applied to study the molecular signature of GC metastasis from different sites [31]. Recent evidences showed that CCNE1 amplification was found associated with liver metastasis in GC of the TP53 subtype [32] while mutations activating or inactivating specific genes were identified in peritoneal metastasis [33]. Circulating tumor DNA (ctDNA) detection is another innovative approach to analyze GC disease. Fang et al. showed that high ctDNA levels were associated with peritoneal recurrence and advanced GC [34]. Lastly, ctDNA harboring the same mutations of primitive GC were associated with late-stage GC. ctDNA can also furnish a tool to track data about the response to therapy in advanced GC, as in the case of amplification of the HER2 gene in trastuzumab-resistant metastatic HER2-positive GC [35].



**Fig. 1** A diagram reporting the current therapeutic algorithm for advanced gastric cancer in Western countries. AGC advanced gastric cancer, FOLFOX fluorouracil + oxaliplatin, Fluoropyrimidines capecitabine or 5-fluorouracil, mono CT irinotecan or paclitaxel, FOLFIRI 5-fluorouracil + irinotecan

## 4 Genetic discordances between Asian and Caucasian GC patients

Differences in development and response to treatments have been observed among ethnically diverse populations. In spite of major incidence, GC Asian patients have a better prognosis and response to treatments than Caucasian ones [36] with a 5-year survival rate of 90% in Japan and 10–30% in Europe [37, 38]. Cristescu et al. [31] classified GC from ethnically different cohorts into four distinct subtypes, on the basis of a pre-defined set of known cancer-related gene expression signatures, suggesting that oncogenic pathways are broadly conserved between populations. On the other hand, studies minimizing differences (clinical management, environmental habits) using SEER data of GC patients treated in the USA showed that the survival advantage of Asians over non-Hispanic whites would continue to hold after control for other well-known prognostic factors [39]. Differing tumor biology has been proposed as a potential explanation for the observed racial disparities in survival [40]. Most of the studies on GC have been conducted on Asian patients; however, even if results are promising, it remains questionable whether they can be fully extrapolated to Caucasian populations, as assessed, *i.e.*, by difference in the somatic mutation rates of several driver genes. Considering the increasing level of understanding of the molecular basis of tumor biology, several biomarkers have been identified for many types of tumors [41]. The frequencies of somatic mutations found between Asian and Caucasian populations were: APC (Asian: Caucasian 6.06 vs. 14.40%,  $p = 0.0076$ ), ARID1A (20.7 vs. 32.1%,  $p = 0.01$ ), KMT2A (4.04 vs. 12.35%,  $p = 0.003$ ), PIK3CA (9.6 vs. 18.52%,  $p = 0.01$ ), and PTEN (2.52 vs. 9.05%,  $p = 0.008$ ) [39]. Unfortunately, there are few defined biomarkers of prognostic and diagnostic use for gastric tumors [42]. Moreover, inter-individual variability of drug response together with tumor heterogeneity are particularly hard to overcome when treating GC. The ability to predict whether patients will respond to specific therapies would be of particular value and would allow for stratifying patients for personalized treatment strategies. Currently, only the status of HER2 for therapy with trastuzumab is approved [43]. The standard treatment for advanced GC is CT and the most widely accepted first-line regimen is a combination of fluoropyrimidine plus platinum with or without trastuzumab [44]. To better design new personalized therapies, especially gene targeting ones, it is indispensable to know in depth the genetic discordances between Caucasian and Asian populations.

## 5 Genome editing mechanisms and ways of action

In order to edit the human genome, to correct or knock out a specific gene expression, a discontinuity should be generated

in the DNA strands. Historically, the first studies and trials were conducted by the use of zinc finger nucleases (ZFNs) [45], meganucleases [46], or transcription activator-like effectors nucleases (TALENs) [47] which were able to generate double strand breaks (DSBs) in specific DNA target genes. Once the nick is created, cells start to repair such gap commonly exploiting the Non-Homologous End Join (NHEJ) repair system, inducing indel mutations and introducing a frameshift effect within the coding region of a gene [48]. As a result, the generation of a premature stop codon will interrupt the expression of the target gene. In case a single-stranded oligonucleotide is provided, acting as a donor template, the cell will repair the created nick by Homologous Directed Repair (HDR) allowing the insertion of new genetic material in the host DNA [49]. However, such nucleases needed to be specifically designed and engineered to recognize a single motif on the DNA double helix. The new approach to the gene editing with CRISPR has recently revolutionized the use of the nucleases. Such system is composed by two RNA elements called CRISPR RNAs (crRNAs), which is designed to be complementary to a specific target, and transactivating CRISPR RNA (tracrRNA) [50]. The hybrid of crRNA and tracrRNA binds a nuclease and helicase protein called CRISPR associated protein 9 (Cas9) commonly derived by *Streptococcus pyogenes*. The crRNA element guides the Cas9 to a specific target site exploiting the standard RNA-DNA complementarity Watson-Crick base-pairing rules, if adjacent to short sequences known as protospacer adjacent motifs (PAMs). Indeed, the single guide RNA (sgRNA) having to precede an NGG site, unlike the other nucleases described, it is not suitable to target any site in the genome. However, generally Cas9 is easier and faster to use than any other nucleases. Cas9 is a bi-lobed protein with the sgRNA nestled between the alpha-helical lobe and the nuclease lobe. There are two main domains located in the nuclease lobe: the RuvC, characterized by an RNase H fold structure [51], which cleaves the non-target DNA strand, and the HNH nuclease domain, which cleaves the target strand of DNA [52]. The nuclease also consists of a recognition lobe (REC) that matches the target sequence in the host DNA. A wild-type Cas9 performs a double strand break in the DNA site, while introducing D10A or H840A mutation into the RuvC- or HNH-like domains will make it incapable to perform a DSB, generating only a single cut per strand instead. These mutants also known as Nickase are extremely useful to improve the specificity of the cut, reducing off-target effects, albeit with a reduced efficiency, using two sgRNA instead of only one [53]. The easiest approach to CRISPR-mediated cancer treatment is to completely knockout a target gene exploiting the NHEJ repair system. In such a way, the nuclease should be directed toward genes involved in chemoresistance, proliferation, migration, invasion, and apoptosis resistance in order to inhibit metastasis and tumor growth.

## 6 CRISPR advancements in GCs

CRISPR is currently revolutionizing the world of gene therapy and even for GC there are some interesting applications of such technology. Yi-Qiang Zhang et al. recently focused on the role of the Prostate-derived Ets factor (PDEF) which is a member of the Ets family of transcription factors, playing an important role in tumorigenesis but with unclear function in GC [54]. They used CRISPR to stably knockout PDEF in the AGS cell line and such depletion inhibited cell proliferation, migration, and invasiveness, demonstrating that the PDEF gene may be involved in the initiation and development of GC and thus might be considered a possible therapeutic target. Benjamin Garcia-Bloj et al. while still using the CRISPR system, exploited a different approach waking up dormant tumor suppressor genes thanks to a mutant version of the Cas9 [15]. Indeed, mutation of two amino acids into the catalytic sites (RUVc and HNH) has generated an inactive form, or so called “deactivated” Cas9 (dCas9), which can be linked to different effector domains, such as p300 [55], an H3K9 acetylase core catalytic domain and VPR [56] for gene activation [57]. Exploiting such molecular mechanism, they reactivated two dormant class II tumor suppressor genes MASPIN and REPRIMO. Indeed, the last one, a novel tp53 dependent G2 arrest mediator candidate, has been proposed to have tumor suppressive functions in gastric [58, 59]. As reported above, CRISPR could also be used to potentiate the effect of primary T cells, disrupting PD-1 expression, upregulating cytokine production, and enhancing cytotoxicity [10]. Such an approach could be promising for future therapies, using immunotherapies of CRISPR-activated cytotoxic T lymphocytes in combination with standard radiotherapy [60].

## 7 Hereditary diffuse gastric cancer

Even if the majority of GCs are sporadic, 1–3% of GCs arise as a result of inherited cancer syndromes. It has now been widely established that germline mutations in the CDH1 gene, encoding the cell-cell adhesion protein E-cadherin, is one of the main genetic causes of the so called hereditary diffuse GC (HDGC) [61]. Heterozygous germline CDH1 mutations increase lifetime risk of developing diffuse GC [62]. HDGC is an autosomal dominant susceptibility illness causing a poorly differentiated adenocarcinoma that infiltrates into the stomach wall causing thickening of the wall (linitis plastica) without forming a distinct mass. In this particular condition, the use of a CRISPR/Cas9-based gene therapy might be an advantageous approach, designing an sgRNA specific for the CDH1 mutation and delivering the machinery *via* lentivirus or adenovirus [63]. However, such kind of therapies are often shown to produce immunogenic effects, inhibiting or nullifying the efficacy of the treatment [64, 65].

## 8 Clinical trials and biomarkers

Albeit CRISPR is rapidly revolutionizing the world of gene therapy, cancer, being a multigenic disease with genetically different cell subpopulations, is hard to be treated with such a system to date. However, gene-editing technologies could be applied to basic research to generate an accurate genetic profile of GC, in order to improve the available personalized medicine treatments, striking cancer-specific biomarkers. It is also important to remark that currently, most of GC literature is based on Asian patients and, as reported above, there are many genetic discordances reported between Asian and Caucasian populations [40]. The lack of specific markers affects not only gene therapies but also chemotherapies. Currently, the most common biomarker reported is HER2, which is overexpressed in some subset of breast, ovarian, gastric, colorectal, pancreatic, and endometrial cancers, and can be targeted by trastuzumab [66]. Many randomized controlled clinical trials with several CT mix have been performed in patients with GC but with little significant improvement [67–70]. In the era of biomarker-directed therapy, evaluating expression of gene signatures or multiple genes associated with patient clinical response and outcome in randomized clinical trials is an important challenge [71, 72]. Genomic sequencing of gastric tumors suggests that exploring molecular aberrations in selected patients may offer new avenues for targeted therapeutic opportunities [18, 73]. New genomic technologies such as the NanoString nCounter system have been used in several clinical trial studies for prognostic and predictive markers [74]. This could improve our understanding of the biological processes and also identify the predictors of CT prognosis and response and improve overall treatment outcomes. One of the most important tumor suppressor genes is the TP53 gene, which encodes p53, and plays a critical role in cell cycle regulation and apoptosis in response to cellular stress [75]. High frequency of TP53 mutations and overexpression of the p53 protein is an important biomarker for predicting prognosis and response to CT in GCs [76]. Some studies have shown that high levels of p53 expression are correlated with multidrug resistance in gastric cancer cell lines [77]. Yong Zha et al. have studied a specific codon and have observed that a polymorphism can be used to identify a subgroup of patients most likely to benefit from the regimens of paclitaxel plus capecitabine [78]. Kakoli Das et al. found a strong association between the higher expression level of the Wnt5A gene and the unresectable gastric tumors [79]. Additionally, low Wnt5A gene expression was a predictor of good response and better progression-free survival in S-1-treated GC patients. Another important theme was studied by Denil L. Fontes et al. which have focused on MET gene amplification that is one of the well-recognized mechanisms of c-MET overexpression and constitutive activation of the MET/HGF pathway and has been reported in 2 to 10% of gastroesophageal (GE) adenocarcinomas [80]. MET-amplified tumors display a higher

**Table 1** A short list of the most commonly mutated genes in GC by analyzing the data from The Cancer Genome Atlas Program ([www.cancer.gov](http://www.cancer.gov))

Symbol	Name	Number affected cases in Cohort	Number of mutations	Description
<i>TTN</i>	Titin	240/404 (59.41%)	783	Central role in contraction of striated muscle. Responsible for muscle passive elasticity. Also known as Connectin
<i>TP53</i>	Tumor protein p53	201/404 (49.75%)	150	Tumor suppressor. Binds to DNA and regulates gene expression to prevent genome mutations
<i>MUC16</i>	Mucin 16, cell surface associated	161/404 (39.85%)	290	Member of mucin family, plays an important role in forming a barrier protecting epithelial cells from pathogens
<i>SYNE1</i>	Spectrin repeat containing nuclear envelop1	133/404 (32.92%)	227	<b>Nuclear envelope</b> protein found in human myocytes and synapses that is involved in maintenance of nuclear organization and structural integrity
<i>CSMD3</i>	CUB and sushi multiple domain3	124/404 (30.69%)	169	Integral component of cells membrane. Biochemical properties and functions are unknown
<i>LRPBI</i>	Low-density lipoprotein receptor-related protein 1B	123/404 (30.45%)	182	Belongs to the LDL receptor family. Potential cell surface proteins that bind and internalize ligands in receptor-mediated endocytosis
<i>FLG</i>	Filaggrin	112/404 (27.72%)	166	Intermediate filament-associated protein that aggregates keratin intermediate filaments in mammalian epidermis
<i>ARID1A</i>	AT-rich interactive domain 1A (SWI-like)	111/404 (27.48%)	121	Nuclear protein that regulate genes transcription by chromatin structure alteration. Has been reported to act as tumor suppressor
<i>FAT4</i>	FAT atypical cadherin 4	111/404 (27.48%)	180	It is a member of the protocadherin family and plays a fundamental role in regulating planar cell polarity
<i>OBSCN</i>	Calmodulin and titin-interacting RhoGEF	100/404 (24.75%)	184	It belongs to the family of giant sarcomeric signaling proteins that includes titin and nebulin and may have a role in the organization of myofibrils
<i>HMCN1</i>	Hemicentin 1	93/404 (23.02%)	138	Plays a role in maintaining the architectural integrity of vertebrate tissues and organs. Serves a role in mitotic cytokinesis cleaving furrow maturation
<i>ZFHX4</i>	Zing finger homeobox4	92/404 (22.7%)	118	May play a role in neural and muscles differentiation. May be involved in transcriptional regulation
<i>FAT3</i>	FAT atypical cadherin 3	92/404 (22.77%)	124	May play a role in the interaction between neurites derived from specific subset of neurons during development
<i>CSMD1</i>	CUB and sushi multiple domains 1	90/404 (22.28%)	132	Has been proposed to have an active role in cell cycle regulation and controlling apoptosis. It may act as a potential tumor suppressor and reduce cell transformation and invasion
<i>PCLO</i>	Piccolo presynaptic cytomatrix protein	88/404 (21.78%)	135	It is part of the presynaptic cytoskeletal matrix and is involved in establishing active synaptic zones and in synaptic vesicle trafficking
<i>DNAH5</i>	Dynein, axonemal, heavy chain 5	87/404 (21.53%)	115	It is part of a microtubule-associated motor protein complex. It functions as a force-generating protein with ATPase activity
<i>RYR2</i>	Ryanodine receptor 2 (cardiac)	85/404 (21.04%)	120	It is the major mediator for sarcoplasmic release of stored calcium ions in the process of cardiac calcium-induced release. Mutations lead to heart failure
<i>SPTA1</i>	Spectrin, alpha, erythrocytic 1	84/404 (20.79%)	116	Links the plasma membrane to the actin cytoskeleton. It has a role in the determination of cell shape and organization of organelles. Mutations lead to hereditary red blood cell disorders
<i>RIMS2</i>	Regulating synaptic membrane exocytosis 2	82/404 (20.30%)	86	It is involved in synaptic membrane exocytosis and in neurotransmitter release
<i>DST</i>	Dystonin	81/404 (20.05%)	125	It is a cytoskeletal linker protein belonging to the plakin family of adhesion junction plaque proteins

We decided to analyze a large cohort including stomach and small intestine adenomas and adenocarcinomas comprising cases of both gender patients and without any filter based on race or ethnicity

pathologic grade and present at a more advanced stage [81]. In the phase I Clinical Trials Program who had MET amplification/mutation testing, a trend for a worse overall survival (OS) for MET positive GE cancers was detected [82]. Unfortunately, this difference was not statistically significant because MET abnormalities can be found in a small group of patients with GE adenocarcinoma and further studies are necessary to better characterize the prognostic and predictive impact of MET alterations [83]. Michael Stahl et al. investigated whether patients with gastric cancer may particularly benefit from EGFR-targeted therapy, since overexpression and high gene copy number of the epidermal growth factor receptor (EGFR) have been observed in a therapeutically relevant number of gastric carcinomas and therefore, it appeared to be a potentially valuable target in the context of perioperative therapy [84, 85]. They found that activation of cancer-driving pathways such as EGFR [86], fibroblast growth factors receptors (FGFR) [87], MET [88], and c-Kit [89] leads to shorter survival, but any significant difference in progression-free survival between the treatment arms was not found. Fibroblast growth factors (FGFs) and their receptors FGFRs are instrumental in a number of normal biologic processes and their dysregulation by mechanisms including activating gene mutations, gene amplification, and gene fusions is believed to drive human cancers, including GCs [90–92]. Van Cutsem et al. have hypothesized that AZD4547 has the potential to provide clinical benefit in patients with advanced gastric adenocarcinoma with tumors displaying FGFR2 polysomy or gene amplification since AZD4547 is a selective FGFR-1, 2, 3 tyrosine kinase inhibitor that has displayed potent activity in preclinical studies [93]. However, there was no clear correlation between the extent of sub-clonal heterogeneity and tumor shrinkage in response to AZD4547 and treatment with AZD4547 did not improve progression-free survival compared with paclitaxel in the overall population or in patients with FGFR2 amplification or polysomy according to FISH selection. Moreover, Y. Yamada et al. have studied the excision repair cross-complementation group 1 (ERCC1), which is an important component of the nuclear excision repair pathway which repairs DNA intra-strand, inter-strand, and DNA-protein crosslinks caused by cisplatin [94]. High mRNA levels of ERCC1 in primary gastric cancer may be associated with a lower response to cisplatin and poor survival. They observed that low ERCC1 expression was a significant independent favorable prognostic factor in patients with advanced gastric cancer who were receiving first-line CT regardless of the treatment regimen [95]. Furthermore, Min Deng et al. studied the role of miR-26a, suggesting that it might play a critical role in GC suppressing growth and metastasis by regulating FGF9 [96]. Ji Mei et al. focused on gene polymorphisms of GSTP1 G/G, XRCC1 A/A, and 5,10-MTHFR T/T and observed a clinical value for predicting the response to the doxorubicin, cisplatin, and 5-fluorouracil treatment for advanced gastric cancer [97]. Furthermore, an important role is

played by the epigenetic silencing of tumor suppressor genes (TSGs) that is commonly observed in GE cancer and is believed to be involved in oncogenesis, metastasis, and CT resistance [98–103]. Bryan J. Schneider et al., in a phase I study, observed that pre-treatment with a DHA can hypomethylate tumor DNA and induce expression of TSGs in cancer cells, thereby sensitizing them to cytotoxic CT [104]. Many genes have been identified as potential biomarkers for personalized cancer-targeted therapy but these studies are frequently limited by a small number of patient samples and usually focused on Asiatic patients. In Table 1 is reported a list of commonly mutated genes in GC. All these findings are encouraging but further evaluation is needed.

## 9 Conclusions

To date, gastrectomy and chemotherapy are still the best therapeutic approaches for GC patients even though the research for alternative strategies is more and more fueled by new intriguing molecular technologies. Indeed, being drug resistance the main cause for treatment failure, the deeper knowledge of new molecular targets and mutations is of vital value to better understand the genetic differences between ethnicity and thus to boost the personalized medicine treatments [105]. However, many issues need to be faced in order to make a step forward. First is the choice of the gene-editing technology. Indeed, very few clinical trials were successful for GC gene therapy and currently no therapies are available. Among the above cited methodologies, CRISPR represents the most affordable and easy to be used in such a kind of treatment due to the simplicity to design the sgRNA and the possibility to target any genes even the ones in heterochromatic loci. The other side of the coin, it is represented by the possibility to produce off-target effects and the need to use a safe way to deliver the molecular machinery into the tumor site avoiding healthy tissues. In such a way, of key importance is the choice of a surface antigen target to selectively deliver the CRISPR complex. One of the most probable targets might be the CarcinoEmbryonic Antigen (CEA) which is commonly expressed in many kinds of GCs [106] as well as the oncofetal antigen 5T4 [107] which has been linked with disease progression and poor clinical outcome. As a delivery system, the best ones available are Adenoviruses (AVs), Adeno-Associated Viruses (AAVs), and RiboNucleoProteins (RNPs) [108]. Indeed, while a virus-based approach is useful to maximize the infection efficacy, and thus the KO efficiency, it may generate a significant immune response neutralizing the biological effect. The RNPs are commonly well tolerated but they have a lower transfection efficiency so they need to be optimized for the use. Moreover, given the fact that CRISPR application *in vivo*, and especially in humans, has a very low working efficiency, more work needs to be done to make such approach

practicable. The most common gene-editing strategies to treat GC are “tumor-suppressor actions,” by which an irreversible gene KO leads to cell growth inhibition or to lose metastatic ability, “suicide gene therapy,” *i.e.*, the transduction of a gene processing a non-toxic “pro-drug” into a toxic one and the “immunomodulation” by which we can potentiate the host immune response against cancer cells. Being p53 one of the master regulator of the genome integrity and resulting mutated in about 60% of gastric cancer, its replacement with a wild-type version is fascinating and in 1999, Ohashi M et al. reported that such reintroduction, through AVs, led to a significant growth inhibition showing to be of clinical interest [109], while the two most common suicide gene therapies are based on the conversion of 5-fluorocytosine to 5-fluorouracil (5-FU), through expression of cytosine deaminase and the phosphorylation of ganciclovir, throughout the herpes simplex virus thymidine kinase, both used in combination with a CEA promoter to make their expression cancer-specific [110, 111]. It is our opinion that currently, molecular research needs to take several steps forward to use gene therapy constantly in clinical practice, especially by uncovering specific surface molecular targets to make such therapies more selective and avoiding any off-target effects. Lastly, we want to report an extremely interesting tool for the discovery of new biomarkers: the Genome-Wide Pooled CRISPR libraries. Such libraries, included in lentiviral plasmids, are used to knockout thousands of genes at the same time and randomly, thus once the cells are exposed to a selective pressure, *i.e.*, anoikis resistance, chemotherapies, hypoxia, *etc.*, we can identify which genes are fundamental for a certain phenotype [112].

**Authorship contribution** Conception and design: LM, LP, and AB. Writing, review, and/or revision of the manuscript: AB, LM, EG, NS, and IS; AB, NS, FC, SP, and IS searched the literature, edited the manuscript, and revised English draft. All authors read and approved the final manuscript.

**Financial support** Dr. Alessio Biagioni was supported by a postdoctoral fellowship of the Italian Foundation for Cancer Research (AIRC).

### Compliance with ethical standards

**Conflict of interest** The authors declare that they have no conflict of interest.

### References

- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., & Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: a Cancer Journal for Clinicians*, *68*, 394–424. <https://doi.org/10.3322/caac.21492>.
- Ishaq, S., & Nunn, L. (2015). Helicobacter pylori and gastric cancer: a state of the art review. *Gastroenterology and Hepatology from Bed to Bench*, *8*(Suppl 1), S6–S14.
- Kim, J., Cho, Y. A., Choi, W. J., & Jeong, S. H. (2014). Gene-diet interactions in gastric cancer risk: A systematic review. *World Journal of Gastroenterology*, *20*(28), 9600–9610. <https://doi.org/10.3748/wjg.v20.i28.9600>.
- World Cancer Research Fund/American Institute for Cancer Research (WCRF/AICR). Continuous update project report: diet, nutrition, physical activity and stomach cancer 2016. Revised 2018. London: World Cancer Research Fund International; 2018.
- Casamayor, M., Morlock, R., Maeda, H., & Ajani, J. (2018). Targeted literature review of the global burden of gastric cancer. *Ecancermedicalscience*, *12*, 883. <https://doi.org/10.3332/ecancer.2018.883> eCollection 2018.
- Berlth, F., Bollschweiler, E., Drebber, U., Hoelscher, A. H., & Moenig, S. (2014). Pathohistological classification systems in gastric cancer: diagnostic relevance and prognostic value. *World Journal of Gastroenterology*, *20*(19), 5679–5684. <https://doi.org/10.3748/wjg.v20.i19.5679>.
- Clements, W. M., Wang, J., Sarnaik, A., Kim, O. J., MacDonald, J., Fenoglio-Preiser, C., Groden, J., & Lowy, A. M. (2002). Beta-catenin mutation is a frequent cause of Wnt pathway activation in gastric cancer. *Cancer Research*, *62*(12), 3503–3506.
- Feng, R., Chen, X., Yu, Y., Su, L., Yu, B., Li, J., Cai, Q., Yan, M., Liu, B., & Zhu, Z. (2010). miR-126 functions as a tumour suppressor in human gastric cancer. *Cancer Letters*, *298*, 50–63.
- Moses, C., Garcia-Bloj, B., Harvey, A. R., & Blancafort, P. (2018). Hallmarks of cancer: The CRISPR generation. *European Journal of Cancer*, *93*, 10–18. <https://doi.org/10.1016/j.ejca.2018.01.002>.
- Su, S., Zou, Z., Chen, F., Ding, N., Du, J., Shao, J., Li, L., Fu, Y., Hu, B., Yang, Y., Sha, H., Meng, F., Wei, J., Huang, X., & Liu, B. (2016). CRISPR-Cas9-mediated disruption of PD-1 on human T cells for adoptive cellular therapies of EBV positive gastric cancer. *Oncotarget*, *6*(1), e1249558. <https://doi.org/10.1080/2162402X.2016.1249558>.
- Deng, X., & Nakamura, Y. (2017). Cancer precision medicine: from cancer screening to drug selection and personalized immunotherapy. *Trends in Pharmacological Sciences*, *38*(1), 15–24. <https://doi.org/10.1016/j.tips.2016.10.013>.
- Gao, Q., Dong, X., Xu, Q., Zhu, L., Wang, F., Hou, Y., & Chao, C. C. (2019). Therapeutic potential of CRISPR/Cas9 gene editing in engineered T-cell therapy. *Cancer Medicine*. <https://doi.org/10.1002/cam4.2257>.
- Dolcetti, R., De Re, V., Canzonieri, V. (2018). Immunotherapy for gastric cancer: time for a personalized approach? *International Journal of Molecular Sciences*, *19*(6). doi:<https://doi.org/10.3390/ijms19061602>.
- Cong, L., Zhou, R., Kuo, Y. C., Cunniff, M., & Zhang, F. (2012). Comprehensive interrogation of natural TALE DNA-binding modules and transcriptional repressor domains. *Nature Communications*, *3*, 968. <https://doi.org/10.1038/ncomms1962>.
- Garcia-Bloj, B., Moses, C., Sgro, A., Plani-Lam, J., Arooj, M., Duffy, C., Thiruvengadam, S., Sorolla, A., Rashwan, R., Mancera, R. L., Leisewitz, A., Swift-Scanlan, T., Corvalan, A. H., & Blancafort, P. (2016). Waking up dormant tumor suppressor genes with zinc fingers, TALEs and the CRISPR/dCas9 system. *Oncotarget*, *7*(37), 60535–60554. <https://doi.org/10.18632/oncotarget.11142>.
- Liu, J., Ben, Q., Lu, E., He, X., Yang, X., Ma, J., Zhang, W., Wang, Z., Liu, T., Zhang, J., & Wang, H. (2018). Long noncoding RNA PANDAR blocks CDKN1A gene transcription by competitive interaction with p53 protein in gastric cancer. *Cell Death & Disease*, *9*(2), 168. <https://doi.org/10.1038/s41419-017-0246-6>.

17. Wagner, A. D., Unverzagt, S., Grothe, W., et al. (2010). Chemotherapy for advanced gastric cancer. *Cochrane Database of Systematic Reviews*, 3, CD004064.
18. Bang, Y. J., Van Cutsem, E., Feyereislova, A., Chung, H. C., Shen, L., Sawaki, A., Lordick, F., Ohtsu, A., Omuro, Y., Satoh, T., Aprile, G., Kulikov, E., Hill, J., Lehle, M., Ruschhoff, J., Kang, Y. K., et al. (2010). Trastuzumab in combination with chemotherapy versus chemotherapy alone for treatment of HER2-positive advanced gastric or gastroesophageal junction cancer (ToGA): a phase 3, open-label, randomised controlled trial. *Lancet*, 376(9742), 687–697.
19. Koizumi, W., Narahara, H., Hara, T., Takagane, A., Akiya, T., Takagi, M., Miyashita, K., Nishizaki, T., Kobayashi, O., Takiyama, W., Toh, Y., Nagaie, T., Takagi, S., Yamamura, Y., Yanaoka, K., Orita, H., & Takeuchi, M. (2008). S-1 plus cisplatin versus S-1 alone for first-line treatment of advanced gastric cancer (SPIRITS trial): a phase III trial. *The Lancet Oncology*, 9, 215–221.
20. Ajani, J. A., Buyse, M., Lichinitser, M., et al. (2013). Combination of cisplatin/S-1 in the treatment of patients with advanced gastric or gastroesophageal adenocarcinoma: results of noninferiority and safety analyses compared with cisplatin/5-fluorouracil in the First-Line Advanced Gastric Cancer study. *Eur J Cancer*, 49, 3616–3624.
21. Ajani, J. A., Rodriguez, W., Bodoky, G., Moiseyenko, V., Lichinitser, M., Gorbunova, V., Vynnychenko, I., Garin, A., Lang, I., & Falcon, S. (2010). Multicenter phase III comparison of cisplatin/S-1 with cisplatin/infusional fluorouracil in advanced gastric or gastroesophageal adenocarcinoma study: the FLAGS trial. *Journal of Clinical Oncology*, 28, 1547–1553.
22. Van Cutsem, E., Moiseyenko, V. M., Tjulandin, S., et al. (2006). Phase III study of docetaxel and cisplatin plus fluorouracil compared with cisplatin and fluorouracil as first-line therapy for advanced gastric cancer: a report of the V325 Study Group. *Journal of Clinical Oncology*, 24, 4991–4997.
23. Al-Batran, S. E., Homann, N., & Pauligk, C. (2019). Perioperative chemotherapy with fluorouracil plus leucovorin, oxaliplatin, and docetaxel versus fluorouracil or capecitabine plus cisplatin and epirubicin for locally advanced, resectable gastric or gastroesophageal junction adenocarcinoma (FLOT4): a randomised, phase 2/3 trial. *Lancet*, 393(10184), 1948–1957.
24. Catalano, V., Graziano, F., Santini, D., D'Emidio, S., Baldelli, A. M., Rossi, D., Vincenzi, B., Giordani, P., Alessandrini, P., Testa, E., Tonini, G., & Catalano, G. (2008). Second-line chemotherapy for patients with advanced gastric cancer: who may benefit? *British Journal of Cancer*, 99, 1402–1407.
25. Hang, Y., Ma, B., Huang, X. T., et al. (2016). Doublet versus single agent as second-line treatment for advanced gastric cancer: a meta-analysis of 10 randomized controlled trials. *Medicine*, 95, e2792.
26. Fuchs, C. S., Tomasek, J., Yong, C. J., Dumitru, F., Passalacqua, R., Goswami, C., Safran, H., dos Santos, L. V., Aprile, G., Ferry, D. R., Melichar, B., Tehfe, M., Topuzov, E., Zalcberg, J. R., Chau, I., Campbell, W., Sivanandan, C., Pikiel, J., Koshiji, M., Hsu, Y., Liepa, A. M., Gao, L., Schwartz, J. D., & Tabernero, J. (2014). Ramucirumab monotherapy for previously treated advanced gastric or gastro-oesophageal junction adenocarcinoma (REGARD): an international, randomised, multicentre, placebo-controlled, phase 3 trial. *Lancet*, 383, 31–39.
27. Wilke, H., Muro, K., Van Cutsem, E., et al. (2014). Ramucirumab plus paclitaxel versus placebo plus paclitaxel in patients with previously treated advanced gastric or gastro-oesophageal junction adenocarcinoma (RAINBOW): a double-blind, randomised phase 3 trial. *The Lancet Oncology*, 15, 1224–1235.
28. Riihimäki, M., Hemminki, A., Sundquist, K., Sundquist, J., & Hemminki, K. (2016). Metastatic spread in patients with gastric cancer. *Oncotarget*, 7(32), 52307–52316. <https://doi.org/10.18632/oncotarget.10740>.
29. Thomassen, I., van Gestel, Y. R., van Ramshorst, B., Luyer, M. D., Bosscha, K., Nienhuijs, S. W., Lemmens, V. E., & de Hingh, I. H. (2014). Peritoneal carcinomatosis of gastric origin: a population-based study on incidence, survival and risk factors. *International Journal of Cancer*, 134(3), 622–628. <https://doi.org/10.1002/ijc.28373>.
30. Kanda, M., & Kodera, Y. (2016). Molecular mechanisms of peritoneal dissemination in gastric cancer. *World Journal of Gastroenterology*, 22(30), 6829–6840. <https://doi.org/10.3748/wjg.v22.i30.6829> **Review**.
31. Cristescu, R., Lee, J., Nebozhyn, M., Kim, K. M., Ting, J. C., Wong, S. S., Liu, J., Yue, Y. G., Wang, J., Yu, K., Ye, X. S., Do, I. G., Liu, S., Gong, L., Fu, J., Jin, J. G., Choi, M. G., Sohn, T. S., Lee, J. H., Bae, J. M., Kim, S. T., Park, S. H., Sohn, I., Jung, S. H., Tan, P., Chen, R., Hardwick, J., Kang, W. K., Ayers, M., Hongyue, D., Reinhard, C., Loboda, A., Kim, S., & Aggarwal, A. (2015). Molecular analysis of gastric cancer identifies subtypes associated with distinct clinical outcomes. *Nature Medicine*, 21(5), 449–456. <https://doi.org/10.1038/nm.3850>.
32. Kim, B., Shin, H. C., Heo, Y. J., Ha, S. Y., Jang, K. T., Kim, S. T., Kang, W. K., Lee, J., & Kim, K. M. (2019). CCNE1 amplification is associated with liver metastasis in gastric carcinoma. *Pathology, Research and Practice*, 4, 152434. <https://doi.org/10.1016/j.prp.2019.152434>.
33. Zhang, J., Huang, J. Y., Chen, Y. N., Yuan, F., Zhang, H., Yan, F. H., Wang, M. J., Wang, G., Su, M., Lu, G., Huang, Y., Dai, H., Ji, J., Zhang, J., Zhang, J. N., Jiang, Y. N., Chen, S. J., Zhu, Z. G., & Yu, Y. Y. (2015). Erratum: whole genome and transcriptome sequencing of matched primary and peritoneal metastatic gastric carcinoma. *Scientific Reports*, 5, 15309. <https://doi.org/10.1038/srep15309>.
34. Fang, W. L., Lan, Y. T., Huang, K. H., Liu, C. A., Hung, Y. P., Lin, C. H., Jhang, F. Y., Chang, S. C., Chen, M. H., Chao, Y., Lin, W. C., Lo, S. S., Fen-Yau Li, A., Wu, C. W., Chiou, S. H., & Shyr, Y. M. (2016). Clinical significance of circulating plasma DNA in gastric cancer. *International Journal of Cancer*, 138(12), 2974–2983. <https://doi.org/10.1002/ijc.30018>.
35. Wang, R., Song, S., Harada, K., Ghazanfari Amlashi, F., Badgwell, B., Pizzi, M. P., Xu, Y., Zhao, W., Dong, X., Jin, J., Wang, Y., Scott, A., Ma, L., Huo, L., Vicente, D., Blum Murphy, M., Shanbhag, N., Tatlonghari, G., Thomas, I., Rogers, J., Kobayashi, M., Vykoukal, J., Estrella, J. S., Roy-Chowdhuri, S., Han, G., Zhang, S., Mao, X., Song, X., Zhang, J., Gu, J., Johnson, R. L., Calin, G. A., Peng, G., Lee, J. S., Hanash, S. M., Futreal, A., Wang, Z., Wang, L., & Ajani, J. A. (2019). Multiplex profiling of peritoneal metastases from gastric adenocarcinoma identified novel targets and molecular subtypes that predict treatment response. *Gut*. <https://doi.org/10.1136/gutjnl-2018-318070>.
36. Wang, J., Sun, Y., & Bertagnolli, M. M. (2015). Comparison of gastric cancer survival between Caucasian and Asian patients treated in the United States: results from the Surveillance Epidemiology and End Results (SEER) database. *Annals of Surgical Oncology*, 22(9), 2965–2971. <https://doi.org/10.1245/s10434-015-4388-4>.
37. Stock, M., & Otto, F. (2005). Gene deregulation in gastric cancer. *Gene*, 360(1), 1–19 **Review**.
38. Parkin, D. M., Bray, F., Ferlay, J., & Pisani, P. (2005). Global cancer statistics, 2002. *CA: a Cancer Journal for Clinicians*, 55(2), 74–108.
39. Jia, F., Teer, J. K., Knepper, T. C., Lee, J. K., Zhou, H. H., He, Y. J., & McLeod, H. L. (2017). Discordance of somatic mutations between Asian and Caucasian patient populations with gastric cancer. *Molecular Diagnosis & Therapy*, 21(2), 179–185. <https://doi.org/10.1007/s40291-016-0250-z>.

40. Kim, J., Sun, C. L., Mailey, B., Prendergast, C., Artinyan, A., Bhatia, S., Pigazzi, A., & Ellenhom, J. D. (2010). Race and ethnicity correlate with survival in patients with gastric adenocarcinoma. *Annals of Oncology*, 21(1), 152–160. <https://doi.org/10.1093/annonc/mdp290>.
41. Mehta, S., Shelling, A., Muthukaruppan, A., Lasham, A., Blenkinsop, C., Laking, G., & Print, C. (2010). Predictive and prognostic molecular markers for cancer medicine. *Therapeutic Advances in Medical Oncology*, 2(2), 125–148. <https://doi.org/10.1177/1758834009360519>.
42. Pinheiro Ddo, R., Ferreira, W. A., Barros, M. B., Araújo, M. D., Rodrigues-Antunes, S., & Borges, B. N. (2014). Perspectives on new biomarkers in gastric cancer: diagnostic and prognostic applications. *World Journal of Gastroenterology*, 20(33), 11574–11585. <https://doi.org/10.3748/wjg.v20.i33.11574> **Review**.
43. Abrahao-Machado, L. F., & Scapulatempo-Neto, C. (2016). HER2 testing in gastric cancer: an update. *World Journal of Gastroenterology*, 22(19), 4619–4625. <https://doi.org/10.3748/wjg.v22.i19.4619> **Review**.
44. Lordick, F., Lorenzen, S., Yamada, Y., & Ilson, D. (2014). Optimal chemotherapy for advanced gastric cancer: is there a global consensus? *Gastric Cancer*, 17(2), 213–225. <https://doi.org/10.1007/s10120-013-0297-z> **Review**.
45. Chou, S. T., Leng, Q., & Mixson, A. J. (2012). Zinc finger nucleases: tailor-made for gene therapy. *Drugs of the Future*, 37(3), 183–196.
46. Silva, G., Poirot, L., Galetto, R., Smith, J., Montoya, G., Duchateau, P., & Pâques, F. (2011). Meganucleases and other tools for targeted genome engineering: perspectives and challenges for gene therapy. *Current Gene Therapy*, 11(1), 11–27 **Review**.
47. Miller, J. C., Tan, S., Qiao, G., Barlow, K. A., Wang, J., Xia, D. F., Meng, X., Paschon, D. E., Leung, E., Hinkley, S. J., Dulay, G. P., Hua, K. L., Ankoudinova, I., Cost, G. J., Urnov, F. D., Zhang, H. S., Holmes, M. C., Zhang, L., Gregory, P. D., & Rebar, E. J. (2011). A TALE nuclease architecture for efficient genome editing. *Nature Biotechnology*, 29(2), 143–148. <https://doi.org/10.1038/nbt.1755>.
48. Lee, H. J., Kim, E., & Kim, J. S. (2010). Targeted chromosomal deletions in human cells using zinc finger nucleases. *Genome Research*, 20(1), 81–89. <https://doi.org/10.1101/gr.099747.109>.
49. Liang, F., Han, M., Romanienko, P. J., & Jasin, M. (1998). Homology-directed repair is a major double-strand break repair pathway in mammalian cells. *Proceedings of the National Academy of Sciences of the United States of America*, 95(9), 5172–5177.
50. Garneau, J. E., Dupuis, M. È., Villion, M., Romero, D. A., Barrangou, R., Boyaval, P., Fremaux, C., Horvath, P., Magadán, A. H., & Moineau, S. (2010). The CRISPR/Cas bacterial immune system cleaves bacteriophage and plasmid DNA. *Nature*, 468(7320), 67–71. <https://doi.org/10.1038/nature09523>.
51. Nishimasu, H., Ran, F. A., Hsu, P. D., Konermann, S., Shehata, S. I., Dohmae, N., Ishitani, R., Zhang, F., & Nureki, O. (2014). Crystal structure of Cas9 in complex with guide RNA and target DNA. *Cell*, 156(5), 935–949. <https://doi.org/10.1016/j.cell.2014.02.001>.
52. Haft, D. H., Selengut, J., Mongodin, E. F., & Nelson, K. E. (2005). A guild of 45 CRISPR-associated (Cas) protein families and multiple CRISPR/Cas subtypes exist in prokaryotic genomes. *PLoS Computational Biology*, 1(6), e60.
53. Dow, L. E., Fisher, J., O'Rourke, K. P., Muley, A., Kasthuber, E. R., Livshits, G., Tschaharganeh, D. F., Succi, N. D., & Lowe, S. W. (2015). Inducible in vivo genome editing with CRISPR-Cas9. *Nature Biotechnology*, 33(4), 390–394. <https://doi.org/10.1038/nbt.3155>.
54. Zhang, Y. Q., Pei, J. H., Shi, S. S., Guo, X. S., Cui, G. Y., Li, Y. F., Zhang, H. P., & Hu, W. Q. (2019). CRISPR/Cas9-mediated knockout of the PDEF gene inhibits migration and invasion of human gastric cancer AGS cells. *Biomedicine & Pharmacotherapy*, 111, 76–85. <https://doi.org/10.1016/j.biopha.2018.12.048>.
55. Hilton, I. B., D'Ippolito, A. M., Vockley, C. M., Thakore, P. I., Crawford, G. E., Reddy, T. E., & Gersbach, C. A. (2015). Epigenome editing by a CRISPR-Cas9-based acetyltransferase activates genes from promoters and enhancers. *Nature Biotechnology*, 33, 510–517.
56. Chavez, A., Scheiman, J., Vora, S., Pruitt, B. W., Tuttle, M., P R Iyer, E., Lin, S., Kiani, S., Guzman, C. D., Wiegand, D. J., Ter-Ovanesyan, D., Braff, J. L., Davidsohn, N., et al. (2015). Highly efficient Cas9-mediated transcriptional programming. *Nature Methods*, 12, 326–328.
57. Perez-Pinera, P., Kocak, D. D., Vockley, C. M., Adler, A. F., Kabadi, A. M., Polstein, L. R., Thakore, P. I., Glass, K. A., Ousterout, D. G., Leong, K. W., Guilak, F., Crawford, G. E., Reddy, T. E., & Gersbach, C. A. (2013). RNA-guided gene activation by CRISPR-Cas9-based transcription factors. *Nature Methods*, 10, 973–976.
58. Sandoval-Bórquez, A., Saavedra, K., Carrasco-Avino, G., Garcia-Bloj, B., Fry, J., Wichmann, I., & Corvalán, A. H. (2015). Noncoding genomics in gastric cancer and the gastric precancerous cascade: pathogenesis and biomarkers. *Disease Markers*, 2015, 1–14.
59. Bernal, C., Aguayo, F., Villarroel, C., Vargas, M., Díaz, I., Ossandon, F. J., Santibáñez, E., Palma, M., Aravena, E., Barrientos, C., & Corvalan, A. H. (2008). Reprimo as a potential biomarker for early detection in gastric cancer. *Clinical Cancer Research*, 14, 6264–6269.
60. Kanda, T., Furuse, Y., Oshitani, H., & Kiyono, T. (2016). Highly efficient CRISPR/Cas9-mediated cloning and functional characterization of gastric cancer-derived Epstein-Barr virus strains. *Journal of Virology*, 90(9), 4383–4393. <https://doi.org/10.1128/JVI.00060-16.Print2016May>.
61. Corso, G., Marrelli, D., & Roviello, F. (2012). Familial gastric cancer and germline mutations of E-cadherin. *Annali Italiani di Chirurgia*, 83(3), 177–182.
62. van der Post, R. S., Vogelaar, I. P., Carneiro, F., Guilford, P., Huntsman, D., Hoogerbrugge, N., Caldas, C., Schreiber, K. E., Hardwick, R. H., Ausems, M. G., Bardram, L., Benusiglio, P. R., Bisseling, T. M., Blair, V., Bleiker, E., Boussioutas, A., Cats, A., Coit, D., DeGregorio, L., Figueiredo, J., Ford, J. M., Heijkoop, E., Hermens, R., Humar, B., Kaurah, P., Keller, G., Lai, J., Ligtenberg, M. J., O'Donovan, M., Oliveira, C., Pinheiro, H., Raganath, K., Rasenberg, E., Richardson, S., Roviello, F., Schackert, H., Seruca, R., Taylor, A., Ter Huurne, A., Tischkowitz, M., Joe, S. T., van Dijck, B., van Grieken, N. C., van Hillegersberg, R., van Sandick, J. W., Vehof, R., van Krieken, J. H., & Fitzgerald, R. C. (2015). Hereditary diffuse gastric cancer: updated clinical guidelines with an emphasis on germline CDH1 mutation carriers. *Journal of Medical Genetics*, 52(6), 361–374. <https://doi.org/10.1136/jmedgenet-2015-103094>.
63. Biagioni, A., Laurenzana, A., Margheri, F., Chillà, A., Fibbi, G., & Del Rosso, M. (2018). Delivery systems of CRISPR/Cas9-based cancer gene therapy. *Journal of Biological Engineering*, 12, 33. <https://doi.org/10.1186/s13036-018-0127-2> **eCollection 2018. Review**.
64. Cepko, C., & Pear, W. (2001). Overview of the retrovirus transduction system. *Current Protocols in Molecular Biology*, 9, 9.9.
65. Annunziato, S., Kas, S. M., Nethe, M., Yücel, H., Del Bravo, J., Pritchard, C., Bin Ali, R., et al. (2016). Modeling invasive lobular breast carcinoma by CRISPR/Cas9-mediated somatic genome

- editing of the mammary gland. *Genes & Development*, 30(12), 1470–1480.
66. English, D. P., Roque, D. M., & Santin, A. D. (2013). HER2 expression beyond breast cancer: therapeutic implications for gynecologic malignancies. *Molecular Diagnosis & Therapy*, 17(2), 85–99. <https://doi.org/10.1007/s40291-013-0024-9> **Review**.
  67. Koizumi, W., Tanabe, S., Saigenji, K., Ohtsu, A., Boku, N., Nagashima, F., Shirao, K., Matsumura, Y., & Gotoh, M. (2003). Phase I/II study of S-1 combined with cisplatin in patients with advanced gastric cancer. *British Journal of Cancer*, 89(12), 2207–2212.
  68. Ohtsu, Y., Shimada, K., Shirao, N., Boku, I., Hyodo, H., & Saito, (et al.). Randomized phase III trial of fluorouracil alone versus fluorouracil plus cisplatin versus uracil and tegafur plus mitomycin in patients with unresectable, advanced gastric cancer: the Japan Clinical Oncology Group Study (JCOG9205). *Journal of Clinical Oncology*, 21(2003), 54e59.
  69. Van Cutsem, E., Moiseyenko, V. M., Tjulandin, S., Majlis, A., Constenla, M., Boni, C., et al. (2006). Phase III study of docetaxel and cisplatin plus fluorouracil compared with cisplatin and fluorouracil as first-line therapy for advanced gastric cancer: a report of the V325 Study Group. *Journal of Clinical Oncology*, 24, 4991e4997.
  70. Narahara, H., Iishi, H., Imamura, H., Tsuburaya, A., Chin, K., Imamoto, H., Esaki, T., Furukawa, H., Hamada, C., & Sakata, Y. (2011). Randomized phase III study comparing the efficacy and safety of irinotecan plus S-1 with S-1 alone as first-line treatment for advanced gastric cancer (study GC0301/TOP-002). *Gastric Cancer*, 14(1), 72–80. <https://doi.org/10.1007/s10120-011-0009-5>.
  71. Das, K., Chan, X. B., Epstein, D., Teh, B. T., Kim, K. M., Kim, S. T., Park, S. H., Kang, W. K., Rozen, S., Lee, J., & Tan, P. (2016). NanoString expression profiling identifies candidate biomarkers of RAD001 response in metastatic gastric cancer. *ESMO Open*, 1, e000009.
  72. Liu, S., Chapman, J. A., Burnell, M. J., Levine, M. N., Pritchard, K. I., Whelan, T. J., Rugo, H. S., Albain, K. S., Perez, E. A., Virk, S., Barry, G., Gao, D., O'Brien, P., Shepherd, L. E., Nielsen TO, & Gelmon, K. A. (2015). Prognostic and predictive investigation of PAM50 intrinsic subtypes in the NCIC CTG MA.21 phase III chemotherapy trial. *Breast Cancer Research and Treatment*, 149(2), 439–448. <https://doi.org/10.1007/s10549-014-3259-1>.
  73. Dulak, A. M., Schumacher, S. E., van Lieshout, J., Imamura, Y., Fox, C., Shim, B., Ramos, A. H., Saksena, G., Baca, S. C., Baselga, J., Taberero, J., Barretina, J., Enzinger, P. C., Corso, G., Roviello, F., Lin, L., Banda, S., Luketich, J. D., Pennathur, A., Meyerson, M., Ogino, S., Shivdasani, R. A., Beer, D. G., Godfrey, T. E., Beroukhi, R., & Bass, A. J. (2012). Gastrointestinal adenocarcinomas of the esophagus, stomach, and colon exhibit distinct patterns of genome instability and oncogenesis. *Cancer Research*, 72(17), 4383–4393.
  74. Lee, J., Sohn, I., Do, I. G., Kim, K. M., Park, S. H., Park, J. O., Park, Y. S., Lim, H. Y., Sohn, T. S., Bae, J. M., Choi, M. G., Lim, D. H., Min, B. H., Lee, J. H., Rhee, P. L., Kim, J. J., Choi, D. I., Tan, I. B., Das, K., Tan, P., Jung, S. H., Kang, W. K., & Kim, S. (2014). Nanostring-based multigene assay to predict recurrence for gastric cancer patients after surgery. *PLoS One*, 9, e90133.
  75. Chen, K., Yang, D., Li, X., et al. (2015). Mutational landscape of gastric adenocarcinoma in Chinese: implications for prognosis and therapy. *Proceedings of the National Academy of Sciences of the United States of America*, 112, 1107e1112.
  76. Busuttill, R. A., Zapparoli, G. V., Haupt, S., et al. (2014). Role of p53 in the progression of gastric cancer. *Oncotarget*, 5, 12016e12026. 4.
  77. Endo, F., Nishizuka, S. S., Kume, K., Ishida, K., Katagiri, H., Ishida, K., Sato, K., Iwaya, T., Koeda, K., & Wakabayashi, G. (2014). A compensatory role of NF- $\kappa$ B to p53 in response to 5-FU-based chemotherapy for gastric cancer cell lines. *PLoS One*, 9, e90155.
  78. Zha, Y., Gan, P., Liu, Q., & Yao, Q. (2016). TP53 codon 72 polymorphism predicts efficacy of paclitaxel plus capecitabine chemotherapy in advanced gastric cancer patients. *Archives of Medical Research*, 47(1), 13–18. <https://doi.org/10.1016/j.arcmed.2015.12.001> **Epub 2015 Dec 13**.
  79. Das, K., Taguri, M., Imamura, H., Sugimoto, N., Nishikawa, K., Yoshida, K., Tan, P., & Tsuburaya, A. (2018). Genomic predictors of chemotherapy efficacy in advanced or recurrent gastric cancer in the GC0301/TOP002 phase III clinical trial. *Cancer Letters*, 412, 208–215. <https://doi.org/10.1016/j.canlet.2017.10.011>.
  80. Jardim, D. L., de Melo, G. D., Falchook, G. S., Janku, F., Zinner, R., Wheler, J. J., Subbiah, V., Piha-Paul, S. A., Fu, S., Murphy, M. B., Ajani, J., Tang, C., Hess, K., Hamilton, S. R., Roy-Chowdhuri, S., Kurzrock, R., Meric-Bernstam, F., & Hong, D. S. (2014). MET aberrations and c-MET inhibitors in patients with gastric and esophageal cancers in a phase I unit. *Oncotarget*, 5(7), 1837–1845.
  81. Comoglio, P. M., Giordano, S., & Trusolino, L. (2008). Drug development of MET inhibitors: targeting oncogene addiction and expedience. *Nature Reviews Drug Discovery*, 7(6), 504–516.
  82. Tuyenman, J. B., Lagarde, S. M., Ten Kate, F. J., Richel, D. J., & van Lanschot, J. J. (2008). Met expression is an independent prognostic risk factor in patients with oesophageal adenocarcinoma. *British Journal of Cancer*, 98(6), 1102–1108.
  83. Lennerz, J. K., Kwak, E. L., Ackerman, A., Michael, M., Fox, S. B., Bergethon, K., Lauwers, G. Y., Christensen, J. G., Wilner, K. D., Haber, D. A., Salgia, R., Bang, Y. J., Clark, J. W., Solomon, B. J., & Iafrate, A. J. (2011). MET amplification identifies a small and aggressive subgroup of esophagogastric adenocarcinoma with evidence of responsiveness to crizotinib. *Journal of Clinical Oncology*, 29(36), 4803–4810.
  84. Stahl, M., Maderer, A., Lordick, F., Mihaljevic, A. L., Kanzler, S., Hoehler, T., Thuss-Patience, P., Mönig, S., Kunzmann, V., Schroll, S., Sandermann, A., Tannapfel, A., Meyer, H. J., Schuhmacher, C., Wilke, H., Moehler, M., & Arbeitsgemeinschaft Internistische Onkologie (AIO) Oesophageal and Gastric Cancer Working Group and the Chirurgische Arbeitsgemeinschaft Onkologie (CAOGI/DGAV) of the German Cancer Society. (2018). Perioperative chemotherapy with or without epidermal growth factor receptor blockade in unselected patients with locally advanced oesophagogastric adenocarcinoma: randomized phase II study with advanced biomarker program of the German Cancer Society (AIO/CAO STO-0801). *European Journal of Cancer*, 93, 119–126. <https://doi.org/10.1016/j.ejca.2018.01.079>.
  85. Kim, M. A., Lee, H. S., Lee, H. E., Jeon, Y. K., Yang, H. K., & Kim, W. H. (2008). EGFR in gastric carcinomas: prognostic significance of protein overexpression and high gene copy number. *Histopathology*, 52(6), 738–746. <https://doi.org/10.1111/j.1365-2559.2008.03021.x>.
  86. Cacina, C., Arian, S., Duzkoylu, Y., Dogan, M. B., Okay, E., Turan, S., et al. (2015). Analyses of EGF A61G gene variation and serum EGF level on gastric cancer susceptibility and clinicopathological parameters. *Anticancer Research*, 35(5), 2709e13.
  87. Park, D. J., Yoon, C., Thomas, N., Ku, G. Y., Janjigian, Y. Y., Kelsen, D. P., et al. (2014). Prognostic significance of targetable angiogenic and growth factors in patients undergoing resection for gastric and gastroesophageal junction cancers. *Annals of Surgical Oncology*, 21(4), 1130e7.
  88. Takahashi, N., Furuta, K., Taniguchi, H., Sasaki, Y., Shoji, H., Honma, Y., et al. (2016). Serum level of hepatocyte growth factor is a novel marker of predicting the outcome and resistance to the treatment with trastuzumab in HER2-positive patients with metastatic gastric cancer. *Oncotarget*, 7(4), 4925e38.

89. Lim, J. B., Kim, D. K., & Chung, H. W. (2014). Clinical significance of serum thymus and activation-regulated chemokine in gastric cancer: potential as a serum biomarker. *Cancer Science*, *105*(10), 1327e33.
90. Jang, J. H., Shin, K. H., & Park, J. G. (2001). Mutations in fibroblast growth factor receptor 2 and fibroblast growth factor receptor 3 genes associated with human gastric and colorectal cancers. *Cancer Research*, *61*, 3541–3543.
91. Murase, H., Inokuchi, M., Takagi, Y., et al. (2014). Prognostic significance of the co- overexpression of fibroblast growth factor receptors 1, 2 and 4 in gastric cancer. *Molecular and Clinical Oncology*, *2*, 509–517.
92. Brooks, A. N., Kilgour, E., & Smith, P. D. (2012). Molecular pathways: fibroblast growth factor signaling: a new therapeutic opportunity in cancer. *Clinical Cancer Research*, *18*, 1855–1862.
93. Van Cutsem, E., Bang, Y. J., Mansoor, W., Petty, R. D., Chao, Y., Cunningham, D., Ferry, D. R., Smith, N. R., Frewer, P., Ratnayake, J., Stockman, P. K., Kilgour, E., & Landers, D. (2017). A randomized, open-label study of the efficacy and safety of AZD4547 monotherapy versus paclitaxel for the treatment of advanced gastric adenocarcinoma with FGFR2 polysomy or gene amplification. *Annals of Oncology*, *28*(6), 1316–1324. <https://doi.org/10.1093/annonc/mdx107>.
94. Metzger, R., Leichman, C. G., Danenberg, K. D., Danenberg, P. V., Lenz, H. J., Hayashi, K., Groshen, S., Salonga, D., Cohen, H., Laine, L., Crookes, P., Silberman, H., Baranda, J., Konda, B., & Leichman, L. (1998). ERCC1 mRNA levels complement thymidylate synthase mRNA levels in predicting response and survival for gastric cancer patients receiving combination cisplatin and fluorouracil chemotherapy. *Journal of Clinical Oncology*, *16*, 309–316.
95. Yamada, Y., Boku, N., Nishina, T., Yamaguchi, K., Denda, T., Tsuji, A., Hamamoto, Y., Konishi, K., Tsuji, Y., Amagai, K., Ohkawa, S., Fujita, Y., Nishisaki, H., Kawai, H., Takashima, A., Mizusawa, J., Nakamura, K., & Ohtsu, A. (2013). Impact of excision repair cross-complementing gene 1 (ERCC1) on the outcomes of patients with advanced gastric cancer: correlative study in Japan Clinical Oncology Group Trial JCOG9912. *Annals of Oncology*, *24*(10), 2560–2565. <https://doi.org/10.1093/annonc/mdt238> Epub 2013 Jul 24.
96. Deng, M., Tang, H. L., Lu, X. H., Liu, M. Y., Lu, X. M., Gu, Y. X., Liu, J. F., & He, Z. M. (2013). miR-26a suppresses tumor growth and metastasis by targeting FGF9 in gastric cancer. *PLoS One*, *8*(8), e72662. <https://doi.org/10.1371/journal.pone.0072662>.
97. Ji, M., Xu, B., Jiang, J. T., Wu, J., Li, X. D., Zhao, W. Q., Zhang, H. Y., Zhou, W. J., & Wu, C. P. (2013). Relationship between glutathione S-transferase P1 (GSTP1), X-ray repair cross complementing group 1 (XRCC1) and 5,10-methylenetetrahydrofolate reductase (5,10-MTHFR) gene polymorphisms and response to chemotherapy in advanced gastric cancer. *Onkologie*, *36*(6), 335–340. <https://doi.org/10.1159/000351260>.
98. Sato, F., & Meltzer, S. J. (2006). CpG island hypermethylation in progression of esophageal and gastric cancer. *Cancer*, *106*, 483–493.
99. Kang, C., Song, J. J., Lee, J., & Kim, M. Y. (2014). Epigenetics: an emerging player in gastric cancer. *World journal of gastroenterology: WJG*, *20*, 6433–6447.
100. Hamilton, J. P., Sato, F., Jin, Z., Greenwald, B. D., Ito, T., Mori, Y., Paun, B. C., Kan, T., Cheng, Y., Wang, S., Yang, J., Abraham, J. M., & Meltzer, S. J. (2006). Reprimo methylation is a potential biomarker of Barrett's-associated esophageal neoplastic progression. *Clinical Cancer Research*, *12*, 6637–6642.
101. Eads, C. A., Lord, R. V., Wickramasinghe, K., Long, T. I., Kurumboor, S. K., Bernstein, L., Peters, J. H., DeMeester, S., DeMeester, T., Skinner, K. A., & Laird, P. W. (2001). Epigenetic patterns in the progression of esophageal adenocarcinoma. *Cancer Research*, *61*, 3410–3418.
102. Wang, J. S., Guo, M., Montgomery, E. A., Thompson, R. E., Cosby, H., Hicks, L., Wang, S., Herman, J. G., & Canto, M. I. (2009). DNA promoter hypermethylation of p16 and APC predicts neoplastic progression in Barrett's esophagus. *The American Journal of Gastroenterology*, *104*, 2153–2160.
103. Oka, D., Yamashita, S., Tomioka, T., Nakanishi, Y., Kato, H., Kaminishi, M., & Ushijima, T. (2009). The presence of aberrant DNA methylation in noncancerous esophageal mucosae in association with smoking history: a target for risk diagnosis and prevention of esophageal cancers. *Cancer*, *115*, 3412–3426.
104. Schneider, B. J., Shah, M. A., Klute, K., Ocean, A., Popa, E., Altorki, N., Lieberman, M., Schreiner, A., Yantiss, R., Christos, P. J., Palmer, R., You, D., Viale, A., Kermani, P., & Scandura, J. M. (2017). Phase I Study of epigenetic priming with azacitidine prior to standard neoadjuvant chemotherapy for patients with resectable gastric and esophageal adenocarcinoma: evidence of tumor hypomethylation as an indicator of major histopathologic response. *Clinical Cancer Research*, *23*(11), 2673–2680. <https://doi.org/10.1158/1078-0432.CCR-16-1896>.
105. Marin, J. J., Al-Abdulla, R., Lozano, E., Briz, O., Bujanda, L., Banales, J. M., & Macias, R. I. (2016). Mechanisms of resistance to chemotherapy in gastric cancer. *Anti-Cancer Agents in Medicinal Chemistry*, *16*(3), 318–334 **Review**.
106. Khare, P. D., Shao-Xi, L., Kuroki, M., Hirose, Y., Arakawa, F., Nakamura, K., Tomita, Y., & Kuroki, M. (2001). Specifically targeted killing of carcinoembryonic antigen (CEA)-expressing cells by a retroviral vector displaying single-chain variable fragmented antibody to CEA and carrying the gene for inducible nitric oxide synthase. *Cancer Research*, *61*(1), 370–375.
107. Shaw, D. M., Embleton, M. J., Westwater, C., Ryan, M. G., Myers, K. A., Kingsman, S. M., Carroll, M. W., & Stern, P. L. (2000). Isolation of a high affinity scFv from a monoclonal antibody recognising the oncofoetal antigen 5T4. *Biochimica et Biophysica Acta*, *1524*(2–3), 238–246.
108. Biagioni, A., Chillà, A., Andreucci, E., Laurenzana, A., Margheri, F., Peppicelli, S., Del Rosso, M., & Fibbi, G. (2017). Type II CRISPR/Cas9 approach in the oncological therapy. *Journal of Experimental & Clinical Cancer Research*, *36*(1), 80. <https://doi.org/10.1186/s13046-017-0550-0> **Review**.
109. Ohashi, M., Kanai, F., Ueno, H., Tanaka, T., Tateishi, K., Kawakami, T., Koike, Y., Ikenoue, T., Shiratori, Y., Hamada, H., & Omata, M. (1999). Adenovirus mediated p53 tumour suppressor gene therapy for human gastric cancer cells in vitro and in vivo. *Gut*, *44*(3), 366–371.
110. Okino, T., Onda, M., Matsukura, N., Inada, K. I., Tatematsu, M., Suzuki, S., & Shimada, T. (2001). Sequential histopathological changes in vivo after suicide gene therapy of gastric cancer induced by N-methyl-N'-nitro-N-nitrosoguanidine in rats. *Japanese Journal of Cancer Research*, *92*(6), 673–679.
111. Shimizu, T., Shimada, H., Ochiai, T., & Hamada, H. (2001). Enhanced growth suppression in esophageal carcinoma cells using adenovirus-mediated fusion gene transfer (uracil phosphoribosyl transferase and herpes simplex virus thymidine kinase). *Cancer Gene Therapy*, *8*, 512–521.
112. Joung, J., Konermann, S., Gootenberg, J. S., Abudayyeh, O. O., Platt, R. J., Brigham, M. D., Sanjana, N. E., & Zhang, F. (2017). Genome-scale CRISPR-Cas9 knockout and transcriptional activation screening. *Nature Protocols*, *12*(4), 828–863. <https://doi.org/10.1038/nprot.2017.016>.

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.