



## Ureidobenzenesulfonamides as efficient inhibitors of carbonic anhydrase II

Immo Serbian<sup>a</sup>, Philipp Schwarzenberger<sup>a</sup>, Anne Loesche<sup>a</sup>, Sophie Hoenke<sup>a</sup>, Ahmed Al-Harrasi<sup>b</sup>, René Csuk<sup>a,\*</sup><sup>a</sup> Martin-Luther-University Halle-Wittenberg, Organic Chemistry, Kurt-Mothes-Str. 2, D-06120 Halle (Saale), Germany<sup>b</sup> University of Nizwa, Chair of Oman's Medicinal Plants and Marine Natural Products, PO Box 33, Birkat Al-Mauz, Nizwa, Oman

## ARTICLE INFO

## Keywords:

Carbonic anhydrase  
Inhibitor  
Urea  
Ureidobenzenesulfonamide  
Glaucoma

## ABSTRACT

Sulfonamides represent an important class of drugs because of their inhibitory effect on carbonic anhydrases (CAs). We therefore synthesized several ureidobenzenesulfonamides and evaluated their bCA II inhibition for their potential use as anti-glaucoma agents. Since these compounds must not show cytotoxic effects, their cytotoxic potential against several human tumor cell lines and non-malignant fibroblasts was investigated. Several fluorophenyl substituted sulfonamides were efficient inhibitors of bCA II. Only one benzylphenyl substituted sulfonamide, however, showed a remarkable selectivity for HT29 colorectal carcinoma cells while being significantly less cytotoxic to non-malignant fibroblasts.

## 1. Introduction

Carbonic anhydrases (CAs; EC 4.2.1.1) are essential for virtually all forms of life since they are involved in the conversion of carbon dioxide and water to bicarbonate and protons. They thereby maintain in tissues and blood an acid-base balance being crucial for the respiratory system. To date 15 different human CAs are known; they are widely distributed in various tissues [1–3]. Therefore carbonic anhydrases represent an interesting drug target to treat pathologies like diuretics [4], epilepsy and glaucoma. For the latter, CA inhibitors (CAIs) are already in use, and their application in treating obesity, bacterial or fungal infections and cancer is presently subject of research [1,2,5–10]. Especially in the last years the development of CAIs was of major interest [6,11–13] as it has been shown that CAIs might be supportive in anti-cancer therapy, and also inhibition of hCA IX and XII which are associated with hypoxic cancers offers therapeutic potential [1,6,14–17]. CA II was found as a novel biomarker for *Pseudomyxoma peritonei* [18], and its altered expression is associated with human carcinogenesis [18]. Quite recently it was shown that the content of CA II activity is related to processes of aging and neuro-degeneration [19]. The main application for CA II inhibitors, however, concerns persons suffering from glaucoma. Glaucoma is among the leading causes of global irreversible blindness. The number of persons suffering from glaucoma will increase to 111.8 million in 2040 [20]. This gives a renewed impetus to the search for and development of CAII inhibitors.

Sulfonamides represent a class of CAIs that are mainly associated with diuretics and anti-glaucoma agents where the sulfonamide moiety

is binding to zinc in CAs [5,21]. Several sulfonamides have already been synthesized in this context as they seem to be very promising agents to treat hypoxic tumors. With regard to SLC-0111 - a sulfonamide inhibitor already in phase I clinical trials - the presence of an ureido moiety seems very promising [2,6,22,23]. The cytotoxic of the latter has already been shown several times, and the bioactivity of several sulfonamides is comparable to that of doxorubicin [24]. CAIs intended for use as antiglaucoma agents must not show any cytotoxicity, and we became interested in developing CAII inhibitors based on sulfonamides being devoid of any cytotoxicity.

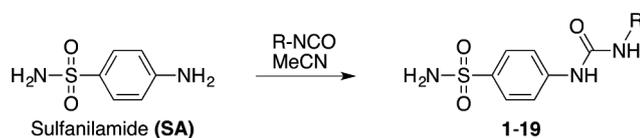
## 2. Results and discussion

## 2.1. Chemistry

The ureidobenzenesulfonamides 1–19 were accessed from the reaction of sulfanilamide with isocyanates [10]. Thereby our focus was the synthesis of benzylphenyl and naphthyl derivatives since preceding docking studies revealed high binding affinities for these compounds and CAII. Thus, compounds 1–19 were obtained in good yields. For comparison, SLC-0111 [25–36] (8, being already in phase I clinical trials) was synthesized, too. [23,37] Work-up of these reactions was easy, since most compounds were obtained in analytical pure form (≥99%) by washing the crude product with acetonitrile and ethanol followed by re-crystallization. Interestingly, 19 could not be precipitated in acetonitrile; even the addition of chloroform (all ureidobenzenesulfonamides are insoluble in this solvent) did not lead to a

\* Corresponding author at: Martin-Luther-University Halle-Wittenberg, Organic Chemistry, Kurt-Mothes-Str. 2, D-06120 Halle (Saale), Germany.

E-mail address: [rene.csuk@chemie.uni-halle.de](mailto:rene.csuk@chemie.uni-halle.de) (R. Csuk).



**Scheme 1.** 1 Naphthalen-1-yl, 2 4-benzylphenyl, 3 2-benzylphenyl, 4 4-methoxy-[1,1'-biphenyl]-3-yl, 5 4-methoxyphenyl, 6 3-methoxyphenyl, 7 2-methoxyphenyl, 8 4-fluorophenyl, 9 3-fluorophenyl, 10 2-fluorophenyl, 11 ethyl, 12 isopropyl, 13 butyl, 14 hexyl, 15 phenyl, 16 benzoyl, 17 ethyl 4-benzoate, 18 4-benzoic acid, 19 5-chloro-2-phenoxyphenyl.

precipitation of **19**. Therefore, **19** was purified by column chromatography (see Scheme 1).

## 2.2. Carbonic anhydrase II inhibition

The synthesized compounds were assayed employing bovine cytosolic carbonic anhydrase bCA II. The results of these assays are compiled in Table 1 and compared to acetazolamide (AAZ) and sulfanilamide (SA). All compounds showed significantly increased inhibition capabilities as compared to parent sulfanilamide. The inhibition rate of compounds **1** (85%), **3** (84%), **10** (80%) and **17** (75%) can even compete to the inhibition rate of AAZ (83%). Interestingly, compound **4** did not show any inhibition at all despite having a 2-substituted methoxy group, which might be considered to be an ideal functional group to inhibit CA II (as can be seen from compound **7**). The biphenyl moiety seems to be too rigid to fit into the cavity - besides having a good calculated binding energy of  $-8.2$  kcal/mol. The lowest  $IC_{50}$  values were measured for compounds **1** ( $0.47$   $\mu$ M), **3** ( $0.50$   $\mu$ M), **7** ( $0.45$   $\mu$ M), **10** ( $0.45$   $\mu$ M) and **17** ( $0.53$   $\mu$ M), and their  $K_i$  values were determined, too. Unfortunately, compound **2** (despite showing the best results in the docking studies) was falling off - compared to AAZ and compounds **1** and **3**. Especially alkyl substituted ureidobenzenesulfonamides **11-13** failed to be good inhibitors. All sulfonamides were mixed type inhibitors. Our results are - at least in part - in agreement with the results from previous studies: Thus, for a 2-methyl-4-methoxy analog of **5** a  $K_i = 3300$  nm [34], for a 3-isopropoxy analog of **6** a  $K_i = 3.9$  nm [38], for **7** a  $K_i = 4700$  nm [9,34], for **8** a  $K_i = 96$  nm [9,34], for **10** a  $K_i = 33$  nm [6], and for **15** a  $K_i = 240$  nm [39] have been reported. All other compounds of this study are either unknown so far (**2-4**, **14**, **16**, **19**) or have not been tested for their ability to inhibit bCAII (**5**, **6**, **9**, **11-13**, **17**, **18**).

**Table 1**

Inhibition of bCA II with compounds **1-19** and comparison to the standard inhibitor acetazolamide (AAZ) and the starting material sulfanilamide (SA). All experiments were performed at least in triplicate; reported are means  $\pm$  SD.

Compound	Inhibition [%]	$IC_{50}$ [ $\mu$ M]	$K_i$ [ $\mu$ M]	$K_i'$ [ $\mu$ M]	$AD_{4Zn}$ [kcal/mol]
AAZ	82.92 $\pm$ 0.37	0.42 $\pm$ 0.06	0.37 $\pm$ 0.01	> 0.40	- / -
SA	6.28 $\pm$ 0.15	9.33 $\pm$ 0.64	- / -	- / -	- / -
<b>1</b>	85.37 $\pm$ 0.65	0.47 $\pm$ 0.06	0.40 $\pm$ 0.04	> 0.30	-8.28
<b>2</b>	72.06 $\pm$ 0.59	0.75 $\pm$ 0.13	2.95 $\pm$ 0.10	> 6.00	-8.95
<b>3</b>	83.82 $\pm$ 0.69	0.50 $\pm$ 0.06	0.58 $\pm$ 0.03	> 0.40	-8.51
<b>4</b>	1.55 $\pm$ 0.56	- / -	- / -	- / -	-8.2
<b>5</b>	66.71 $\pm$ 0.37	0.56 $\pm$ 0.08	- / -	- / -	-7.38
<b>6</b>	67.76 $\pm$ 0.74	0.57 $\pm$ 0.09	0.80 $\pm$ 0.19	0.31 $\pm$ 0.14	-7.93
<b>7</b>	75.19 $\pm$ 0.25	0.45 $\pm$ 0.04	0.39 $\pm$ 0.01	> 2.50	-7.12
<b>8</b>	66.86 $\pm$ 0.03	0.53 $\pm$ 0.02	- / -	- / -	-7.44
<b>9</b>	75.13 $\pm$ 0.49	0.45 $\pm$ 0.07	0.54 $\pm$ 0.07	0.35 $\pm$ 0.07	-7.4
<b>10</b>	80.42 $\pm$ 0.56	0.45 $\pm$ 0.02	0.37 $\pm$ 0.01	> 3.00	-7.33
<b>11</b>	36.4 $\pm$ 1.14	1.48 $\pm$ 0.13	- / -	- / -	-6.88
<b>12</b>	41.70 $\pm$ 0.46	1.31 $\pm$ 0.11	- / -	- / -	-6.87
<b>13</b>	56.5 $\pm$ 1.27	0.78 $\pm$ 0.08	- / -	- / -	-7.5
<b>14</b>	74.90 $\pm$ 0.13	0.52 $\pm$ 0.05	0.53 $\pm$ 0.01	0.33 $\pm$ 0.01	-7.1
<b>15</b>	70.52 $\pm$ 0.40	0.52 $\pm$ 0.03	0.49 $\pm$ 0.03	> 0.50	-7.66
<b>16</b>	33.46 $\pm$ 0.13	1.22 $\pm$ 0.48	- / -	- / -	-7.51
<b>17</b>	75.36 $\pm$ 0.33	0.53 $\pm$ 0.04	0.49 $\pm$ 0.06	> 0.20	-7.89
<b>18</b>	58.4 $\pm$ 0.23	0.69 $\pm$ 0.03	- / -	- / -	-8.03
<b>19</b>	60.52 $\pm$ 0.80	0.72 $\pm$ 0.07	- / -	- / -	-7.69

## 2.3. Cytotoxicity activity

Since a low cytotoxicity is mandatory for anti-glaucoma active agents, compounds **1-19** were also screened in colorimetric sulforhodamine B (SRD) assays to examine their *in vitro* cytotoxicity. The  $EC_{50}$  values were determined for the nonmalignant mouse fibroblasts NIH 3T3 and six human tumor cell lines. The results are summarized in Table 2.

By and large, the ureidobenzenesulfonamides were not cytotoxic for the tested cell lines. Compound **19**, however, showed weak cytotoxicity for all evaluated cell lines. Rather outstanding results can be observed with compound **2** being a moderate bCA II inhibitor. Compound **2** showed a surprisingly high selectivity for the HT29 colorectal carcinoma cells with a selectivity factor  $S_{i(Ni3-HT3/HT29)} \geq 14$ . Although an  $EC_{50}$  value of  $2.1$   $\mu$ M is not as low as those measured for commercial chemotherapeutics (for example, paclitaxel or *cis*-platinum), our results show the potential of CA II inhibitors having been neglected for many years (see Fig. 1).

## 2.4. Molecular docking

As shown by Santos-Martins et al. [40], Autodock [41] seems well suited for docking of ligands with zinc-metallo-enzymes, and therefore we applied the Autodock4<sub>Zn</sub> force field using the X-ray crystallographic structure of 1V9E (from www.rcsb.org) as it matches the bovine carbonic anhydrase II used in the inhibition assays. All synthesized compounds docked with reasonable good scores (cf. Supplementary material). The sulfonamide moiety is fitted well next to the zinc ion making ionic interactions. Most of the docked compounds yielded similar top poses in the active site. Compound **1**, **3** and **4** are outliers compared to the other sulfonamides (compound **2** and **7** are depicted as examples). We herein identified benzylphenyl and biphenyl residues as interesting inhibitors. However, it is not possible to see a direct correlation of the Autodock4<sub>Zn</sub> scores to the measured inhibitory potential. This might be due either to the small sample size, the considerably large chemical difference in the chosen residues or intrinsic limitations of Autodock for calculations of metallo-enzymes.

## 3. Conclusion

Most of the synthesized ureidobenzenesulfonamides were good inhibitors for CAII but not cytotoxic for several human tumor cell lines

**Table 2**

Cytotoxicity of compounds 1–19. EC<sub>50</sub> values in  $\mu\text{M}$  from SRB assay after 96 h of treatment; nonmalignant mouse fibroblasts (NIH 3T3) and human cancer cell lines: A375 (melanoma), 518A2 (melanoma), HT29 (colorectal carcinoma), A549 (adenocarcinoma), FaDu (squamous carcinoma), SW1736 (thyroid carcinoma); independent experiments were at least performed in triplicate; standard errors are given).

Compound	NIH-3 T3	A375	518A2	HT29	A549	FaDu	SW1736
SA	> 30	> 30	> 30	> 30	> 30	> 30	> 30
1; 3–8	> 30	> 30	> 30	> 30	> 30	> 30	> 30
2	> 30	> 30	7.4 $\pm$ 0.8	2.1 $\pm$ 0.5	> 30	> 30	> 30
9	> 30	> 30	26.6 $\pm$ 2.1	> 30	> 30	> 30	> 30
10–18	> 30	> 30	> 30	> 30	> 30	> 30	> 30
19	14.6 $\pm$ 1.9	19.85 $\pm$ 1.4	11.1 $\pm$ 0.9	9.5 $\pm$ 0.8	16.8 $\pm$ 1.9	21.1 $\pm$ 2.9	19.5 $\pm$ 3.0

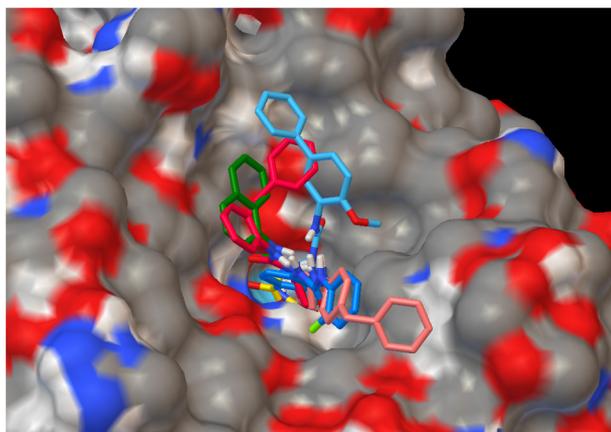


Fig. 1. Active site of hCAII 1v9e with compounds 1 (green), 2 (rose), 3 (red), 4 (light blue), 7 (dark blue).

and non-malignant fibroblasts (NIH 3T3). The benzylphenyl substituted ureidobenzenesulfonamide 2, however, seems to be an exception of the rule, inasmuch as this compound showed remarkable results in the SRB-assay as it was highly selective and cytotoxic for human colorectal adenoma carcinoma HT29 cells; this compound was only a very moderate inhibitor for bCAII. A comparison of the most similar benzylphenyl compounds 2 and 3 with 1 and 4 showed ureidobenzenesulfonamides as an interesting class of highly selective CAIs with promising therapeutic potential as anti-glaucoma agents.

## 4. Experimental section

### 4.1. General

Reagents were bought from commercial suppliers and used without further purification. The solvents were dried according to usual procedures. TLC was performed on silica gel (Macherey-Nagel, detection with UV absorption). Melting points are uncorrected (Büchi M-565). NMR spectra were recorded using the VARIAN spectrometers Gemini 2000 or Unity 500 at 27 °C ( $\delta$  given in ppm;  $J$  in Hz, typical experiments: <sup>13</sup>C-APT, HMBC, HSQC). ASAP-MS spectra were taken on an Advion expression CMS-L with an ASAP/APCI Ion source (capillary voltage 150 V, capillary temperature 220 °C, and voltage of the ion source: 15 V; APCI source temperature 300 °C with 5  $\mu\text{A}$ ). IR spectra were recorded on a Perkin-Elmer Spectrum Two (UATR Two Unit). The SRB assays were performed as previously reported.

### 4.2. Molecular docking

Crystal structure of the Carbonic anhydrase (PDB = 1V9E) was retrieved from the protein databank. The enzyme was prepared according to usual procedures. Hydrogen atoms were added and water molecules were removed. Zinc was assigned a positive charge 2 as reported for Autodock4<sub>Zn</sub>. Ligand minimizations were performed with MMFF94

force field in Datawarrior. Openbabel was used to create the pdbqt files. Ligands were prepared using MGLTools 1.5.6. Calculations were done with Lamarckian Genetic algorithm; the grid center was placed at the zinc ion with a box size of 20 Å.

### 4.3. Carbonic anhydrase II inhibition assay

#### 4.3.1. Spectrometer and chemicals

A TECAN SpectraFluorPlus working on the kinetic mode and measuring the absorbance at  $\lambda = 415 \text{ nm}$  was used for the enzymatic studies. Carbonic anhydrase II (bCAII, from bovine erythrocytes) as well as 4-nitrophenyl acetate (4-NA) and acetazolamide were purchased from Sigma.

#### 4.3.2. Standard solutions preparation

Preparation of 50 mM Tris-HCl buffer solutions: Tris(hydroxymethyl)-aminomethane (606 mg) was dissolved in bidistilled water (100 ml) and adjusted with HCl to a pH of  $8.0 \pm 0.1$ . Buffer was freshly prepared and stored in the refrigerator. bCA II solution: the enzyme ( $\geq 3000 \text{ W-A units/mg}$ , 4.37 mg) was dissolved in freshly prepared buffer pH 8.0 (20 ml). 4-NA solution 6 mM: 4-NA (21.6 mg) was dissolved in methanol (2.1 ml) and bi-distilled water (17.9 ml). All solutions were stored in Eppendorf caps in the refrigerator or freezer, if necessary. The pure compounds were initially dissolved in DMSO. The final concentrations for the enzymatic assay were obtained by diluting the stock solution with bi-distilled water. No inhibition was detected by residual DMSO.

#### 4.3.3. bCA II assays

A mixture of buffer solution pH 8.0 (125  $\mu\text{L}$ ), enzyme (25  $\mu\text{L}$ ) and compounds solutions (25  $\mu\text{L}$ ) was prepared and incubated at 37 °C for 20 min in well plates. 4-NA (25  $\mu\text{L}$ ) was added to start the enzymatic reaction. The relative inhibition was determined as the quotient of the slopes (compound divided by blank) of the linear ranges. The concentration of each compound was 1  $\mu\text{M}$ . The used substrate concentration was  $[4\text{-NA}] = 0.75 \text{ mM}$ . The absorbance data ( $\lambda = 415 \text{ nm}$ ) was recorded under a controlled temperature of 37 °C for 10 min at 1 min intervals. The IC<sub>50</sub> values were calculated using GraphPad Prism 5 software. The final inhibitor concentrations were as follows:  $[\text{inhibitor}] = 0.05 \mu\text{M}, 0.1 \mu\text{M}, 0.2 \mu\text{M}, 0.4 \mu\text{M}, 0.5 \mu\text{M}, 0.8 \mu\text{M}, 1.0 \mu\text{M}, 2.0 \mu\text{M}, 5.0 \mu\text{M}, 6.0 \mu\text{M}$ . A mixture of various compounds solutions (25  $\mu\text{L}$ , 3 different concentrations and once blank), buffer solution (125  $\mu\text{L}$ ) and enzyme solution (25  $\mu\text{L}$ ) was prepared and incubated at 37 °C for 20 min. The substrate (25  $\mu\text{L}$ , 4 different concentrations) was added to start the enzymatic reaction. The substrate concentrations in the test were as follows:  $[4\text{-NA}] = 0.75 \text{ mM}, 0.50 \text{ mM}, 0.25 \text{ mM}, 0.15 \text{ mM}$ . The mode of inhibition as well as  $K_i$  and  $K_i'$  were determined using Lineweaver-Burk plots, Dixon plots and Cornish Bowden plots, respectively.

#### 4.4. General procedure for the synthesis of compounds 1–18

Sulfanilamide (2.5 mmol, 431 mg) was dissolved in dry acetonitrile

(10 ml) and the corresponding isocyanate (1 eq.) was added. The reaction was stirred for 12 h at room temperature. The precipitate was filtered off and washed subsequently with acetonitrile (5 ml), ethanol (5 ml) and diethyl ether (5 ml) to yield compounds 1–18 each as colorless crystalline solid.

#### 4.5. Syntheses

##### 4.5.1. 4-(3-(Naphthalen-1-yl)ureido)benzenesulfonamide (1) [42]

73%;  $R_F = 0.53$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 254 °C; IR (ATR):  $\nu = 1686m, 1491m, 1300m, 1245m, 1212m, 1147s, 794m, 769m, 614s, 542s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 9.40$  (s, 1H, 8-H), 8.85 (s, 1H, 6-H), 8.10 (d,  $J = 8.4$  Hz, 1H, 10-H), 7.97 (d,  $J = 7.5$  Hz, 1H, 14-H), 7.93 (d,  $J = 7.9$  Hz, 1H, 17-H), 7.79–7.72 (m, 2H, 3-H), 7.69–7.62 (m, 2H, 4-H), 7.69–7.64 (m, 1H, 12-H), 7.60–7.55 (m, 1H, 16-H), 7.55–7.48 (m, 1H, 11-H), 7.47 (m, 1H, 15-H), 7.21 (s, 2H, 1-H) ppm; <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 153.2$  (C7), 143.3 (C5), 137.7 (C9), 134.3 (C2), 134.2 (C13), 128.9 (C10), 127.3 (C3), 126.6 (C14), 126.4 (C11), 126.3 (C15), 126.3 (C16), 123.9 (C18), 121.8 (C17), 118.5 (C12), 117.9 (C4) ppm; MS (APCI):  $m/z$  (%) = 342.2 ([M+H]<sup>+</sup>, 88); analysis calcd for C<sub>17</sub>H<sub>15</sub>N<sub>3</sub>SO<sub>3</sub> (341.39): C 59.81, H 4.43, N 12.31, S 9.39; found: C 59.68, H 4.61, N 12.17, S 9.17.

##### 4.5.2. 4-(3-(4-Benzylphenyl)ureido)benzenesulfonamide (2)

46%;  $R_F = 0.52$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 231 °C; IR (ATR):  $\nu = 3262w, 2362w, 1651m, 1590m, 1538s, 1388m, 1329s, 1303m, 1220m, 1151s, 1096m, 587s, 546s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 8.98$  (s, 1H, 6-H), 8.68 (s, 1H, 8-H), 7.73–7.66 (m, 2H, 3-H), 7.60–7.54 (m, 2H, 4-H), 7.38–7.33 (m, 2H, 10-H), 7.26 (t,  $J = 7.6$  Hz, 2H, 16-H), 7.22–7.18 (m, 2H, 15-H), 7.18–7.15 (m, 1H, 19-H), 7.16 (s, 2H, 1-H), 7.14–7.11 (m, 2H, 11-H), 3.86 (s, 2H, 13-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 152.7$  (C7), 143.3 (C5), 142.0 (C14), 137.7 (C9), 137.2 (C2), 135.6 (C12), 129.5 (C3), 129.1 (C16), 128.8 (C15), 127.25 (C12), 126.3 (C19), 119.1 (C10), 117.8 (C4), 40.9 (C13); MS (APCI):  $m/z$  (%) = 382.2 ([M+H]<sup>+</sup>, 40); analysis calcd for C<sub>20</sub>H<sub>19</sub>N<sub>3</sub>SO<sub>3</sub> (381.45): C 62.97, H 5.02, N 11.02, S 8.41; found: C 62.82, H 5.23, N 4.76, S 8.22.

##### 4.5.3. 4-(3-(2-Benzylphenyl)ureido)benzenesulfonamide (3)

41%;  $R_F = 0.61$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 220 °C; IR (ATR):  $\nu = 3378m, 3347w, 3249w, 1654s, 1592m, 1533s, 1486s, 1339s, 1226m, 1156s, 1096w, 838w, 742m, 656m, 576m, 538s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 9.29$  (s, 1H, 6-H), 8.14 (s, 1H, 8), 7.75–7.72 (m, 1H, 12-H), 7.72–7.68 (m, 2H, 3-H), 7.61–7.54 (m, 2H, 4-H), 7.30–7.24 (m, 2H, 18-H), 7.21–7.18 (m, 1H, 14-H), 7.21–7.14 (m, 2H, 17-H), 7.17 (s, 2H, 1-H), 7.17–7.15 (m, 1H, 13-H), 7.06–7.03 (m, 1H, 11-H), 7.03–6.99 (m, 1H, 19-H), 3.97 (s, 2H, 15-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 153.1$  (C7), 143.4 (C5), 140.3 (C9), 137.2 (C2), 136.9 (C10), 132.7 (C16), 129.2 (C17), 128.9 (C18), 127.3 (C3), 127.0 (C14), 126.5 (C13), 124.3 (C11), 123.6 (C12), 117.8 (C4), 36.7 (C15); MS (APCI):  $m/z$  (%) = 382.3 ([M+H]<sup>+</sup>, 31); analysis calcd for C<sub>20</sub>H<sub>19</sub>N<sub>3</sub>SO<sub>3</sub> (381.45): C 62.97, H 5.02, N 11.02, S 8.41; found: C 62.70, H 5.25, N 10.81, S 8.21.

##### 4.5.4. 4-(3-(4-Methoxy-[1,1'-biphenyl]-3-yl)ureido)benzenesulfonamide (4)

37%;  $R_F = 0.53$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 270 °C; IR (ATR):  $\nu = 3370m, 3227w, 1709m, 1597m, 1540s, 1317s, 1247m, 1222m, 1151s, 820s, 755s, 697s, 617s, 531s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 9.71$  (s, 1H, 6-H), 8.49 (d,  $J = 2.3$  Hz, 1H, 14-H), 8.41 (s, 1H, 8-H), 7.76–7.69 (m, 2H, 3-H), 7.65–7.59 (m, 2H, 4-H), 7.59–7.54 (m, 2H, 17-H), 7.47–7.39 (m, 2H, 18-H), 7.34–7.28 (m, 1H, 19-H), 7.26 (dd,  $J = 8.4, 2.3$  Hz, 1H, 12-H), 7.18 (s, 2H, 13-H), 7.11 (d,  $J = 8.5$  Hz, 1H, 11-H), 3.92 (s, 3H, 15-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 152.7$  (C7), 147.9 (C10), 143.3 (C5), 140.7 (C16), 137.3 (C2),

133.2 (C13), 129.3 (C18), 129.1 (C9), 127.4 (C3), 127.3 (C19), 126.7 (C12), 120.8 (C9), 117.7 (C4), 117.3 (C14), 111.7 (C11), 56.5 (C15); MS (APCI):  $m/z$  (%) = 398.2 ([M+H]<sup>+</sup>, 20); analysis calcd for C<sub>20</sub>H<sub>19</sub>N<sub>3</sub>SO<sub>4</sub> (397.45): C 60.44, H 4.82, N 10.57, S 8.07; found: C 60.20, H 4.99, N 10.32, S 7.83.

##### 4.5.5. 4-(3-(4-Methoxyphenyl)ureido)benzenesulfonamide (5) [43]

68%;  $R_F = 0.43$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 229 °C; IR (ATR):  $\nu = 3258m, 1649s, 1592m, 1560s, 1305s, 1251s, 1161s, 828m, 594m, 544s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 8.94$  (s, 1H, 6-H), 8.56 (s, 1H, 8-H), 7.74–7.66 (m, 2H, 3-H), 7.61–7.54 (m, 2H, 4-H), 7.39–7.28 (m, 2H, 10-H), 7.16 (s, 2H, 1-H), 6.91–6.80 (m, 2H, 11-H), 3.70 (s, 3H, 13-H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 155.2$  (C12), 152.9 (C7), 143.5 (C5), 137.1 (C2), 132.7 (C9), 127.2 (C3), 120.8 (C10), 117.7 (C4), 114.5 (C11), 55.6 (C13); MS (APCI):  $m/z$  (%) = 322.2 ([M+H]<sup>+</sup>, 9); analysis calcd for C<sub>14</sub>H<sub>15</sub>N<sub>3</sub>SO<sub>4</sub> (321.35): C 52.33, H 4.70, N 13.08, S 9.98; found: C 52.11, H 4.93, N 12.94, S 9.72.

##### 4.5.6. 4-(3-(3-Methoxyphenyl)ureido)benzenesulfonamide (6) [44]

48%;  $R_F = 0.45$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 238 °C; IR (ATR):  $\nu = 3392w, 3280 w, 1694m, 15901m, 1528m, 1286m, 1152s, 771m, 549s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 9.01$  (s, 1H, 6-H), 8.76 (s, 1H, 8-H), 7.74–7.69 (m, 2H, 3-H), 7.62–7.55 (m, 2H, 4-H), 7.20–7.17 (m, 1H, 10-H), 7.18 (s, 2H, 1-H), 7.18–7.15 (m, 1H, 13-H), 6.93 (ddd,  $J = 8.1, 2.0, 0.9$  Hz, 1H, 14-H), 6.56 (ddd,  $J = 8.2, 2.5, 0.9$  Hz, 1H, 12-H), 3.72 (s, 3H, 15-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 160.2$  (C11), 152.6 (C7), 143.2 (C5), 141.0 (C9), 137.3 (C2), 130.1 (C13), 127.3 (C3), 117.9 (C4), 111.2 (C14), 108.1 (C12), 104.6 (C10), 55.4 (C15); MS (APCI):  $m/z$  (%) = 322.2 ([M+H]<sup>+</sup>, 3); analysis calcd for C<sub>14</sub>H<sub>15</sub>N<sub>3</sub>SO<sub>4</sub> (321.35): C 52.33, H 4.70, N 13.08, S 9.98; found: C 52.00, H 4.97, N 12.86, S 9.72.

##### 4.5.7. 4-(3-(2-Methoxyphenyl)ureido)benzenesulfonamide (7) [9,34,45]

52%;  $R_F = 0.53$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 225 °C; IR (ATR):  $\nu = 3321w, 1684m, 1591m, 1540s, 1309m, 1149s, 844m, 744s, 665m, 610m, 546s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 9.65$  (s, 1H, 6-H), 8.32 (s, 1H, 8-H), 8.10 (dd,  $J = 8.0, 1.7$  Hz, 1H, 14-H), 7.75–7.70 (m, 2H, 3-H), 7.63–7.57 (m, 2H, 4-H), 7.18 (s, 2H, 1-H), 7.01 (dd,  $J = 8.2, 1.5$  Hz, 1H, 11-H), 6.95 (td,  $J = 7.7, 1.7$  Hz, 1H, 12-H), 6.89 (td,  $J = 7.6, 1.5$  Hz, 1H, 13-H), 3.87 (s, 3H, 15-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 152.62$  (C7), 148.3 (C10), 143.4 (C5), 137.2 (C2), 128.7 (C9), 127.3 (C3), 122.7 (C12), 121.0 (C14), 118.9 (C13), 117.7 (C4), 111.3 (C11), 56.3 (C15); MS (APCI):  $m/z$  (%) = 322.2 ([M+H]<sup>+</sup>, 6); analysis calcd for C<sub>14</sub>H<sub>15</sub>N<sub>3</sub>SO<sub>4</sub> (321.35): C 52.33, H 4.70, N 13.08, S 9.98; found: C 52.12, H 4.97, N 12.85, S 9.77.

##### 4.5.8. 4-(3-(4-Fluorophenyl)ureido)benzenesulfonamide (8)

81%;  $R_F = 0.38$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 236 °C (lit.: 242–243 °C [34]); IR (ATR):  $\nu = 3412w, 3337w, 1673m, 1696m, 1501s, 1327m, 1210m, 1147s, 914m, 836m, 627m, 537s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 9.02$  (s, 1H, 6-H), 8.79 (s, 1H, 8-H), 7.74–7.68 (m, 2H, 3-H), 7.62–7.55 (m, 2H, 4-H), 7.45 (dd,  $J = 9.2, 4.9$  Hz, 2H, 10-H), 7.17 (s, 2H, 1-H), 7.12 (td,  $J = 8.9, 1.3$  Hz, 2H, 11-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 158.0$  (d,  $J = 238.9$  Hz, C12), 152.8 (C7), 143.3 (C5), 137.3 (C2), 136.1 (d,  $J = 2.4$  Hz, C9), 127.3 (C3), 120.7 (d,  $J = 8.1$  Hz, C10), 117.9 (C4), 115.8 (d,  $J = 22.4$  Hz, C11); MS (APCI):  $m/z$  (%) = 310.2 ([M+H]<sup>+</sup>, 99); analysis calcd for C<sub>13</sub>H<sub>12</sub>FN<sub>3</sub>SO<sub>3</sub> (309.32): C 50.48, H 3.91, N 13.58, S 10.37; found: C 50.21, H 4.11, N 13.39, S 10.20.

##### 4.5.9. 4-(3-(3-Fluorophenyl)ureido)benzenesulfonamide (9) [44]

72%;  $R_F = 0.39$  (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 240 °C; IR (ATR):  $\nu = 3297w, 1701m, 1541m, 1312m, 1157s, 829m, 773m, 657m, 540m, 526s\text{ cm}^{-1}$ ; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta = 9.09$  (s, 1H, 6-H), 8.99 (s, 1H, 8-H), 7.76–7.69 (m, 2H, 3-H), 7.63–7.56 (m, 2H, 4-H), 7.47 (dt,  $J = 11.9, 2.3$  Hz, 1H, 10-H), 7.30 (td,  $J = 8.2, 6.9$  Hz, 1H, 12-H),

7.19 (s, 2H, 1-H), 7.15–7.10 (m, 1H, 14-H), 6.83–6.75 (m, 1H, 13-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 162.8 (d, *J* = 240.8 Hz, C11), 152.6 (C7), 143.01 (C5), 141.7 (d, *J* = 11.1 Hz, C9), 137.6, 130.8 (d, *J* = 9.6 Hz, C13), 127.3 (C3), 118.1 (C4), 114.6 (d, *J* = 2.7 Hz, C14), 109.0 (d, *J* = 21.1 Hz, C10), 105.6 (d, *J* = 26.4 Hz, C12); MS (APCI): *m/z* (%) = 310.2 ([M+H]<sup>+</sup>, 99); analysis calcd for C<sub>13</sub>H<sub>12</sub>FN<sub>3</sub>SO<sub>3</sub> (309.32): C 50.48, H 3.91, N 13.58, S 10.37; found: C 50.23, H 3.68, N 13.42, S 10.13.

#### 4.5.10. 4-(3-(2-Fluorophenyl)ureido)benzensulfonamide (10) [6]

67%; R<sub>F</sub> = 0.55 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 233 °C; IR (ATR):  $\nu$  = 3396w, 3307w, 3204w, 1703m, 1541m, 1305s, 1253m, 1149s, 1100m, 906m, 836m, 761s, 676m, 610m, 548s, cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 9.23 (s, 1H, 6-H), 8.70 (s, 1H, 8-H), 8.11 (td, *J* = 8.2, 1.7 Hz, 1H, 14-H), 7.76–7.70 (m, 2H, 3-H), 7.63–7.56 (m, 2H, 4-H), 7.27–7.20 (m, 1H, 11-H), 7.20–7.10 (m, 1H, 13-H), 7.19 (s, 2H, 1-H), 7.06–6.98 (m, 1H, 12-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 152.7 (d, *J* = 241.8 Hz, C10), 152.4 (C7), 143.0 (C5), 137.6 (C2), 127.6 (d, *J* = 10.2 Hz, C9), 127.4 (C3), 125.0 (d, *J* = 3.4 Hz, C14), 123.4 (d, *J* = 7.5 Hz, C12), 121.3 (d, *J* = 1.6 Hz, C13), 117.9 (C4), 115.5 (d, *J* = 19.0 Hz, C11); MS (APCI): *m/z* (%) = 310.2 ([M+H]<sup>+</sup>, 46); analysis calcd for C<sub>13</sub>H<sub>12</sub>FN<sub>3</sub>SO<sub>3</sub> (309.32): C 50.48, H 3.91, N 13.58, S 10.37; found: C 50.15, H 4.19, N 13.51, S 10.04.

#### 4.5.11. 4-(3-Ethylureido)benzensulfonamide (11) [44]

63%; R<sub>F</sub> = 0.35 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 232 °C; IR (ATR):  $\nu$  = 3295w, 1661m, 1593m, 1535m, 1317m, 1233m, 1159s, 1097m, 832m, 741m, 589m, 537s cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 8.79 (s, 1H, 6-H), 7.68–7.60 (m, 2H, 3-H), 7.55–7.47 (m, 2H, 4-H), 7.11 (s, 2H, 1-H), 6.22 (t, *J* = 5.6 Hz, 1H, 8-H), 3.10 (qd, *J* = 7.1, 5.5 Hz, 2H, 9-H), 1.04 (t, *J* = 7.2 Hz, 3H, 10-H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 155.16 (C7), 144.18 (C5), 136.36 (C2), 127.16 (C3), 117.20 (C4), 34.44 (C9), 15.77 (C10); MS (APCI): *m/z* (%) = 244.1 ([M+H]<sup>+</sup>, 29); analysis calcd for C<sub>9</sub>H<sub>13</sub>N<sub>3</sub>SO<sub>3</sub> (243.28): C 44.43, H 5.39, N 17.27, S 13.18; found: C 44.30, H 17.47, N 17.00, S 12.86.

#### 4.5.12. 4-(3-Isopropylureido)benzensulfonamide (12) [44]

49%; R<sub>F</sub> = 0.7 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 223 °C; IR (ATR):  $\nu$  = 3325w, 1634s, 1133m, 1247m, 1149s, 1099m, 739m, 587m, 539m cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 8.65 (s, 1H, 6-H), 7.68–7.60 (m, 2H, 3-H), 7.53–7.46 (m, 2H, 4-H), 7.11 (s, 2H, 1-H), 6.12 (d, *J* = 7.5 Hz, 1H, 8-H), 3.82–3.66 (m, 1H, 9-H), 1.08 (d, *J* = 6.6 Hz, 5H, 10-H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 154.5 (C7), 144.1 (C5), 136.4 (C2), 127.2 (C3), 117.1 (C4), 41.5 (C9), 23.3 (C10); MS (APCI): *m/z* (%) = 256.2 ([M+H]<sup>+</sup>, 99); analysis calcd for C<sub>10</sub>H<sub>15</sub>N<sub>3</sub>SO<sub>3</sub> (257.31): C 46.68, H 5.88, N 16.33, S 12.46; found: C 46.42, H 6.03, N 16.02, S 12.18.

#### 4.5.13. 4-(3-Butylureido)benzensulfonamide (13) [46–49]

64%; R<sub>F</sub> = 0.41 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 195 °C (lit.: 196–199 °C [48]); IR (ATR):  $\nu$  = 3403w, 3296m, 3088w, 2963w, 1687s, 1593m, 1533m, 1324m, 1228m, 1154s, 830s, 598m, 542s cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 8.76 (s, 1H, 6-H), 7.67–7.61 (m, 2H, 3-H), 7.54–7.47 (m, 2H, 4-H), 7.11 (s, 2H, 1-H), 6.23 (t, *J* = 5.6 Hz, 1H, 8-H), 3.07 (q, *J* = 6.9 Hz, 2H, 9-H), 1.45–1.35 (m, 2H, 10-H), 1.35–1.24 (m, 2H, 11-H), 0.88 (t, *J* = 7.3 Hz, 3H, 12-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 155.2 (C7), 144.2 (C5), 136.4 (C2), 127.2 (C3), 117.2 (C4), 39.2 (C9), 32.2 (C10), 20.0 (C11), 14.1 (C12); MS (APCI): *m/z* (%) = 272.2 ([M+H]<sup>+</sup>, 23); analysis calcd for C<sub>11</sub>H<sub>17</sub>N<sub>3</sub>SO<sub>3</sub> (271.34): C 48.69, H 6.32, N 15.49, S 11.82; found: C 48.70, H 6.51, N 15.24, S 11.69.

#### 4.5.14. 4-(3-Hexylureido)benzensulfonamide (14)

72%; R<sub>F</sub> = 0.49 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 201 °C; IR (ATR):  $\nu$  = 3410m, 3352m, 3201w, 2938m, 2850m, 1682s, 1596m, 1307s, 1237m, 1151s, 825m, 590m, 541m, 513m cm<sup>-1</sup>; <sup>1</sup>H NMR

(400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 8.77 (s, 1H, 6-H), 7.68–7.60 (m, 2H, 3-H), 7.54–7.46 (m, 2H, 4-H), 7.12 (s, 2H, 1-H), 6.24 (t, *J* = 5.7 Hz, 1H, 8-H), 3.06 (td, *J* = 6.9, 5.7 Hz, 2H, 9-H), 1.40 (tt, *J* = 7.2, 3.8 Hz, 2H, 10-H), 1.30–1.21 (m, 4H, 12-H + 13-H), 1.26–1.25 (m, 2H, 13-H), 0.89–0.81 (m, 3H, 14-H); <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 155.2 (C7), 144.2 (C5), 136.3 (C2), 127.2 (C3), 117.2 (C4), 39.5 (C9), 31.4 (C12), 30.0 (C11), 26.5 (C11), 22.5 (C13), 14.4 (C14); MS (APCI): *m/z* (%) = 300.2 ([M+H]<sup>+</sup>, 32); analysis calcd for C<sub>13</sub>H<sub>21</sub>N<sub>3</sub>SO<sub>3</sub> (299.39): C 52.15, H 7.07, N 14.04, S 10.71; found: C 52.00, H 7.23, N 13.77, S 10.51.

#### 4.5.15. 4-(3-Phenylureido)benzensulfonamide (15) [39,45,50–58]

68%; R<sub>F</sub> = 0.4 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 229 °C (lit.: 233–235 °C [34]); IR (ATR):  $\nu$  = 3306w, 1699s, 1593s, 1526m, 1316m, 1147s, 746m, 647m, 534s cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 9.02 (s, 1H, 6-H), 8.75 (s, 1H, 8-H), 7.75–7.69 (m, 2H, 3-H), 7.63–7.56 (m, 2H, 4-H), 7.48–7.41 (m, 2H, 10-H), 7.32–7.24 (m, 2H, 11-H), 7.18 (s, 2H, 1H), 6.98 (tt, *J* = 7.3, 1.1 Hz, 1H, 12-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 152.7 (C7), 143.3 (C5), 139.8 (C9), 137.3 (C2), 129.3 (C11), 127.3 (C3), 122.7 (C12), 118.9 (C10), 117.9 (C4); MS (APCI): *m/z* (%) = 292.2 ([M+H]<sup>+</sup>, 84); analysis calcd for C<sub>13</sub>H<sub>13</sub>N<sub>3</sub>SO<sub>3</sub> (291.33): C 53.60, H 4.50, N 14.42, S 11.01; found: C 53.42, H 4.77, N 14.05, S 10.78.

#### 4.5.16. N-((4-Sulfamoylphenyl)carbamoyl)benzamide (16)

44%; R<sub>F</sub> = 0.7 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 263 °C; IR (ATR):  $\nu$  = 3339w, 3245w, 1700s, 1593m, 1160m, 1097m, 750m, 704s, 544s cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 11.12 (s, 1H, 8-H), 11.03 (s, 1H, 6-H), 8.04–7.98 (m, 2H, 11-H), 7.82–7.73 (m, 4H, 3-H + 4-H), 7.69–7.62 (m, 1H), 7.57–7.50 (m, 2H, 4-H), 7.27 (s, 2H, 1-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 169.2 (C9), 151.6 (C7), 141.1 (C4), 139.3 (C2), 133.6 (C10), 132.6 (C13), 129.1 (C4), 128.8 (C12), 127.3 (C11), 119.9 (C4); MS (APCI): *m/z* (%) = 320.2 ([M+H]<sup>+</sup>, 6); analysis calcd for C<sub>14</sub>H<sub>13</sub>N<sub>3</sub>SO<sub>4</sub> (319.34): C 52.66, H 4.10, N 13.16, S 10.04; found: C 52.41, H 4.28, N 13.02, S 9.76.

#### 4.5.17. Ethyl 4-(3-(4-sulfamoylphenyl)ureido)benzoate (17) [44]

84%; R<sub>F</sub> = 0.4 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 263 °C; IR (ATR):  $\nu$  = 3298w, 1698s, 1591s, 1411m, 1370m, 1150m, 1100m, 1016m, 902w, 768m, 642m, 572m, 538s cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 9.18 (s, 1H, 8-H), 9.14 (s, 1H, 6-H), 7.91–7.86 (m, 2H, 11-H), 7.77–7.70 (m, 2H, 3-H), 7.64–7.60 (m, 2H, 4-H), 7.60–7.56 (m, 2H, 10-H), 7.19 (s, 2H, 1-H), 4.26 (q, *J* = 7.0 Hz, 2H, 14-H), 1.29 (t, *J* = 7.1 Hz, 3H, 15-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 165.8 (C13), 152.4 (C7), 144.4 (C9), 142.9 (C5), 137.7 (C2), 130.8 (C11), 127.3 (C3), 123.56 (C12), 118.2 (C4), 118.0 (C10), 60.8 (C14), 14.7 (C15); MS (APCI): *m/z* (%) = 364.2 ([M+H]<sup>+</sup>, 12); analysis calcd for C<sub>16</sub>H<sub>17</sub>N<sub>3</sub>SO<sub>5</sub> (363.39): C 52.88, H 4.72, N 11.56, S 8.82; found: C 52.51, H 4.99, N 11.31, S 8.62.

#### 4.5.18. 4-(3-(4-Sulfamoylphenyl)ureido)benzoic acid (18)

Compound **12** (200 mg, 0.55 mmol) was suspended in methanol (10 ml) and potassium hydroxide (310 mg, 5.5 mmol) was added. The mixture was heated under reflux for 4 h. After cooling to room temperature the white solid was filtered off and washed with methanol to give compound **18** as a colorless crystalline solid (196 mg, 0.54 mmol, 98%); R<sub>F</sub> = 0.11 (SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH/DMF, 16:1:1); m.p.: 299–303 °C; IR (ATR):  $\nu$  = 1655s, 1590s, 1535s, 1329m, 1218m, 1151s, 764s, 640m, 541s cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 12.60 (s, 1H, 14-H), 9.15 (s, 1H, 8-H), 9.15 (s, 1H, 6-H), 7.92–7.84 (m, 2H, 11-H), 7.77–7.69 (m, 2H, 3-H), 7.64–7.59 (m, 2H, 4-H), 7.58–7.54 (m, 2H, 10-H), 7.19 (s, 2H, 1-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 167.4 (C13), 152.5 (C7), 144.1 (C9), 142.93 (C5), 137.65 (C2), 131.0 (C10), 127.3 (C3), 124.5 (C12), 118.2 (C4), 117.9 (C11); MS (APCI): *m/z* (%) = 336.2 ([M+H]<sup>+</sup>, 99) analysis calcd for C<sub>14</sub>H<sub>13</sub>N<sub>3</sub>SO<sub>5</sub> (335.34): C 50.14, H 3.91, N 12.53, S 9.56; found: C 49.87, H 4.17, N 12.32, S 9.37.

#### 4.5.19. 4-(3-(5-chloro-2-phenoxyphenyl)ureido)benzenesulfonamide (19)

Sulfanilamide (206 mg, 1.2 mmol) was dissolved in acetonitrile (10 ml) and 5-chloro-2-phenoxyphenylisocyanate (294 mg, 1.2 mmol) was added. The reaction was stirred for 24 h at room temperature. After removing the solvent under reduced pressure, the residue was purified by column chromatography (SiO<sub>2</sub>, hexane/ethyl acetate, 2:1) to give compound **19** as an off-white crystalline solid (72%, 360 mg, 0.86 mmol); R<sub>f</sub> = 0.35 (SiO<sub>2</sub>, hexane/ethyl acetate, 2:1); m.p.: 209 °C; IR (ATR):  $\nu$  = 3361*m*, 1713*m*, 1590*s*, 1529*s*, 1473*m*, 1310*m*, 1218*s*, 1146*m*, 838*m*, 612*m*, 535*s*, 492*s* cm<sup>-1</sup>; <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  = 9.68 (s, 1H, 6-H), 8.74 (s, 1H, 8-H), 8.36 (d, *J* = 2.6 Hz, 1H, 10-H), 7.75–7.69 (*m*, 2H, 3-H), 7.61–7.54 (*m*, 2H, 4-H), 7.46–7.38 (*m*, 2H, 17-H), 7.19 (s, 2H, 1-H), 7.20–7.16 (*m*, 1H, H18), 7.11–7.04 (*m*, 2H, 16-H), 7.00 (dd, *J* = 8.7, 2.6 Hz, 1H, 12-H), 6.82 (d, *J* = 8.7 Hz, 1H, 13-H); <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  156.7 (C15), 152.4 (C7), 144.5 (C14), 142.8 (C5), 137.7 (C2), 132.6 (C28), 130.7 (C17), 128.0 (C11), 127.4 (C3), 124.6 (C15), 122.4 (C12), 119.8 (C13), 119.2 (C10), 119.1 (C16), 118.0 (C4); MS (APCI): *m/z* (%) = 418.1 ([M+H]<sup>+</sup>, 13%) analysis calcd for C<sub>19</sub>H<sub>16</sub>ClN<sub>3</sub>SO<sub>4</sub> (417.87): C 54.61, H 3.86, N 10.06, S 7.67; found: C 54.41, H 4.03, N 9.75, S 7.50.

#### Acknowledgements

We like to thank Dr. D. Ströhl and his team for the NMR spectra, and Dr. R. Kluge for measuring the MS spectra. We are indebted to Ms V. Simon, BSc., for measuring the IR spectra. The cell lines were kindly provided by Dr. Th. Müller (Dep. of Haematology/Oncology, Martin-Luther-Universität Halle-Wittenberg). Financial support by the Oman Research Council ORG/HSS/14/004 to A. A.-H.) is gratefully acknowledged.

#### Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.bioorg.2019.103123>.

#### References

- N. Chiaramonte, M.N. Romanelli, E. Teodori, C.T. Supuran, Amino acids as building blocks for carbonic anhydrase inhibitors, *Metabolites* 8 (2018) 36.
- C. Congiu, V. Onnis, A. Deplano, G. Balboni, N. Dedeoglu, C.T. Supuran, Synthesis of sulfonamides incorporating piperazinyl-ureido moieties and their carbonic anhydrase I, II, IX and XII inhibitory activity, *Bioorg. Med. Chem. Lett.* 25 (2015) 3850–3853.
- E. Kupriyanova, N. Pronina, D. Los, Carbonic anhydrase - a universal enzyme of the carbon-based life, *Photosynthetica* 55 (2017) 3–19.
- C.T. Supuran, Diuretics: from classical carbonic anhydrase inhibitors to novel applications of the sulfonamides, *Curr. Pharm. Des.* 14 (2008) 641–648.
- M. Ahmed, M.A. Qadir, A. Hameed, M.N. Arshad, A.M. Asiri, M. Muddassar, Sulfonamides containing curcumin scaffold: synthesis, characterization, carbonic anhydrase inhibition and molecular docking studies, *Bioorg. Chem.* 76 (2018) 218–227.
- F. Carta, D. Vullo, S.M. Osman, Z. AlOthman, C.T. Supuran, Synthesis and carbonic anhydrase inhibition of a series of SLC-011.1 analogs, *Bioorgan. Med. Chem.* 25 (2017) 2569–2576.
- A. Casini, A. Scozzafava, A. Mastrolorenzo, C.T. Supuran, Sulfonamides and sulfonylated derivatives as anticancer agents, *Curr. Cancer Drug Tar.* 2 (2002) 55–75.
- H.I. Gul, C. Yamali, H. Sakagami, A. Angeli, J. Leitans, A. Kazaks, K. Tars, D.O. Ozgun, C.T. Supuran, New anticancer drug candidates sulfonamides as selective hCA IX or hCA XII inhibitors, *Bioorg. Chem.* 77 (2018) 411–419.
- C.T. Supuran, Carbonic anhydrase inhibitors as emerging agents for the treatment and imaging of hypoxic tumors, *Exp. Opin. Invest. Drugs* 27 (2018) 963–970.
- C.T. Supuran, Special Issue: Sulfonamides, *Molecules*, vol. 22, 2017.
- G. De Simone, V. Alterio, C.T. Supuran, Exploiting the hydrophobic and hydrophilic binding sites for designing carbonic anhydrase inhibitors, *Exp. Opin. Drug Dis.* 8 (2013) 793–810.
- M.D. Altıntop, B. Sever, A. Ozdemir, K. Kucukoglu, H. Onem, H. Nadaroglu, Z.A. Kaplanckli, Potential inhibitors of human carbonic anhydrase isozymes I and II: design, synthesis and docking studies of new 1,3,4-thiadiazole derivatives, *Bioorgan. Med. Chem.* 25 (2017) 3547–3554.
- M.N. Peerzada, P. Khan, K. Ahmad, M.I. Hassan, A. Azam, Synthesis, characterization and biological evaluation of tertiary sulfonamide derivatives of pyridyl-indole based heteroaryl chalcone as potential carbonic anhydrase IX inhibitors and anticancer agents, *Eur. J. Med. Chem.* 155 (2018) 13–23.
- P.K. Chrysanthopoulos, P. Mujumdar, L.A. Woods, O. Dolezal, B. Ren, T.S. Peat, S.A. Poulsen, Identification of a new zinc binding chemotype by fragment screening, *J. Med. Chem.* 60 (2017) 7333–7349.
- M.G. El-Gazzar, N.H. Nafie, A. Nocentini, M.M. Ghorab, H.I. Heiba, C.T. Supuran, Carbonic anhydrase inhibition with a series of novel benzenesulfonamide-triazole conjugates, *J. Enzym. Inhib. Med. Chem.* 33 (2018) 1565–1574.
- S. Singh, C.L. Lomelino, M.Y. Mboge, S.C. Frost, R. McKenna, Cancer drug development of carbonic anhydrase inhibitors beyond the active site, *Molecules* 23 (2018) E1045.
- J.Y. Winum, M. Rami, A. Scozzafava, J.L. Montero, C. Supuran, Carbonic anhydrase IX: a new druggable target for the design of antitumor agents, *Med. Res. Rev.* 28 (2008) 445–463.
- P. Jarvinen, A.J. Kivela, P. Nummela, A. Lepisto, A. Ristimaki, S. Parkkila, Carbonic anhydrase II: a novel biomarker for pseudomyxoma peritonei, *APMIS* 125 (2017) 207–212.
- A. Pollard, F. Shephard, J. Freed, S. Liddell, L. Chakrabarti, Mitochondrial proteomic profiling reveals increased carbonic anhydrase II in aging and neurodegeneration, *Aging-Us* 8 (2016) 2425–2436.
- Y.-C. Tham, X. Li, T.Y. Wong, H.A. Quigley, T. Aung, C.-Y. Cheng, Global prevalence of glaucoma and projections of glaucoma burden through 2014, *Ophthalmology* 121 (2014) 2081–2090.
- C.T. Supuran, Carbonic anhydrases - an overview, *Curr. Pharm. Des.* 14 (2008) 603–614.
- W.M. Eldehna, M. Fares, M. Ceruso, H.A. Ghabbour, S.M. Abou-Seri, H.A. Abdel-Aziz, D.A. Abou El Ella, C.T. Supuran, Amido/ureidobenzene-sulfonamides-isatin conjugates as low nanomolar/subnanomolar inhibitors of the tumor-associated carbonic anhydrase isoform XII, *Eur. J. Med. Chem.* 110 (2016) 259–266.
- M.Y. Mboge, B.P. Mahon, N. Lamas, L. Socorro, F. Carta, C.T. Supuran, S.C. Frost, R. McKenna, Structure activity study of carbonic anhydrase IX: Selective inhibition with ureido-substituted benzenesulfonamides, *Eur. J. Med. Chem.* 132 (2017) 184–191.
- M.M. Ghorab, M. Ceruso, M.S. Alsaïd, Y.M. Nissan, R.K. Arafa, C.T. Supuran, Novel sulfonamides bearing pyrrole and pyrrolopyrimidine moieties as carbonic anhydrase inhibitors: synthesis, cytotoxic activity and molecular modeling, *Eur. J. Med. Chem.* 87 (2014) 186–196.
- N.M. Abdel Gawad, N.H. Amin, M.T. Elsaadi, F.M.M. Mohamed, A. Angeli, V. De Luca, C. Capasso, C.T. Supuran, Synthesis of 4-(thiazol-2-ylamino)-benzenesulfonamides with carbonic anhydrase I, II and IX inhibitory activity and cytotoxic effects against breast cancer cell lines, *Bioorg. Med. Chem.* 24 (2016) 3043–3051.
- E. Andreucci, S. Peppicelli, F. Carta, G. Brisotto, E. Biscontin, J. Ruzzolini, F. Bianchini, A. Biagioni, C.T. Supuran, L. Calorini, Carbonic anhydrase IX inhibition affects viability of cancer cells adapted to extracellular acidosis, *J. Mol. Med.* 95 (2017) 1341–1353.
- E. Andreucci, J. Ruzzolini, S. Peppicelli, F. Bianchini, A. Laurenzana, F. Carta, C.T. Supuran, L. Calorini, The carbonic anhydrase IX inhibitor SLC-0111 sensitizes cancer cells to conventional chemotherapy, *J. Enzyme Inhib. Med. Chem.* 34 (2019) 117–123.
- A. Angeli, D. Tanini, T.S. Peat, L. Di Cesare Mannelli, G. Bartolucci, A. Capperucci, C. Ghelardini, C.T. Supuran, F. Carta, Discovery of new selenoureido analogues of 4-(4-fluorophenylureido)benzenesulfonamide as carbonic anhydrase inhibitors, *ACS Med. Chem. Lett.* 8 (2017) 963–968.
- M. Bozdag, F. Carta, M. Ceruso, M. Ferraroni, P.C. McDonald, S. Dedhar, C.T. Supuran, Discovery of 4-hydroxy-3-(3-(phenylureido)benzenesulfonamides as SLC-0111 analogues for the treatment of hypoxic tumors overexpressing carbonic anhydrase IX, *J. Med. Chem.* 61 (2018) 6328–6338.
- F. Carta, D. Vullo, S.M. Osman, Z. Al Othman, C.T. Supuran, Synthesis and carbonic anhydrase inhibition of a series of SLC-0111 analogs, *Bioorg. Med. Chem.* 25 (2017) 2569–2576.
- F.E. Lock, P.C. McDonald, Y. Lou, I. Serrano, S.C. Chafe, C. Ostlund, S. Aparicio, J.Y. Winum, C.T. Supuran, S. Dedhar, Targeting carbonic anhydrase IX depletes breast cancer stem cells within the hypoxic niche, *Oncogene* 32 (2013) 5210–5219.
- C.L. Lomelino, B.P. Mahon, R. McKenna, F. Carta, C.T. Supuran, Kinetic and X-ray crystallographic investigations on carbonic anhydrase isoforms I, II, IX and XII of a thioureido analog of SLC-0111, *Bioorg. Med. Chem.* 24 (2016) 976–981.
- F. Pacchiano, M. Aggarwal, B.S. Avvaru, A.H. Robbins, A. Scozzafava, R. McKenna, C.T. Supuran, Selective hydrophobic pocket binding observed within the carbonic anhydrase II active site accommodate different 4-substituted-ureido-benzenesulfonamides and correlate to inhibitor potency, *Chem Commun (Cambridge, UK)* 46 (2010) 8371–8373.
- F. Pacchiano, F. Carta, P.C. McDonald, Y. Lou, D. Vullo, A. Scozzafava, S. Dedhar, C.T. Supuran, Ureido-substituted benzenesulfonamides potently inhibit carbonic anhydrase IX and show antimetastatic activity in a model of breast cancer metastasis, *J. Med. Chem.* 54 (2011) 1896–1902.
- F. Pacchiano, F. Carta, D. Vullo, A. Scozzafava, C.T. Supuran, Inhibition of  $\beta$ -carbonic anhydrases with ureido-substituted benzenesulfonamides, *Bioorg. Med. Chem. Lett.* 21 (2011) 102–105.
- D. Vullo, C.T. Supuran, A. Scozzafava, G. De Simone, S.M. Monti, V. Alterio, F. Carta, Kinetic and X-ray crystallographic investigations of substituted 2-thio-6-oxo-1,6-dihydropyrimidine-benzenesulfonamides acting as carbonic anhydrase inhibitors, *Bioorg. Med. Chem.* 24 (2016) 3643–3648.
- A. Angeli, F. Carta, G. Bartolucci, C.T. Supuran, Synthesis of novel acyl selenoureido benzenesulfonamides as carbonic anhydrase I, II, VII and IX inhibitors, *Bioorgan. Med. Chem.* 25 (2017) 3567–3573.
- Q. Shi, T.M. Kaiser, Z.W. Dentmon, M. Ceruso, D. Vullo, C.T. Supuran, J.P. Snyder, Design and validation of FRESH, a drug discovery paradigm resting on robust

- chemical synthesis, ACS Med. Chem. Lett. 6 (2015) 518–522.
- [39] D. Vullo, M. Franchi, E. Gallori, J. Antel, A. Scozzafava, C.T. Supuran, Carbonic anhydrase inhibitors. Inhibition of mitochondrial isozyme V with aromatic and heterocyclic sulfonamides, J. Med. Chem. 47 (2004) 1272–1279.
- [40] D. Santos-Martins, S. Forli, M.J. Ramos, A.J. Olson, AutoDock4(Zn): an improved auto dock force field for small-molecule docking to zinc metalloproteins, J. Chem. Inf. Model. 54 (2014) 2371–2379.
- [41] G.M. Morris, R. Huey, W. Lindstrom, M.F. Sanner, R.K. Belew, D.S. Goodsell, A.J. Olson, AutoDock4 and AutoDockTools4: automated docking with selective receptor flexibility, J. Comput. Chem. 30 (2009) 2785–2791.
- [42] W.H. Brooks, D.E. McCloskey, K.G. Daniel, S.E. Ealick, J.A. Secrist III, W.R. Waud, A.E. Pegg, W.C. Guida, In silico chemical library screening and experimental validation of a novel 9-aminoacridine based lead-inhibitor of human S-adenosylmethionine decarboxylase, J. Chem. Inf. Model. 47 (2007) 1897–1905.
- [43] T. Minami, K. Hamada, A. Sekine, Thermal recording sheet containing amino-benzenesulfonamides as developer and fluoran leuco dye, JP08282117A; CAPLUS, 1997, 61098.
- [44] D.S. Goldfarb, Method using lifespan-altering compounds for altering the lifespan of eukaryotic organisms, and screening for such compounds, US20090163545A1; CAPLUS, 2009, 846110.
- [45] S. Singh, C.T. Supuran, Chemometric modeling of breast cancer associated carbonic anhydrase IX inhibitors belonging to the ureido-substituted benzene sulfonamide class, J. Enzyme Inhib. Med. Chem. 29 (2014) 877–883.
- [46] R. Behnisch, F. Hoffmeister, H. Horstmann, E. Schraufstaetter, W. Wirth, Sulfonamides with anticonvulsive effect, Med. Chem., Abhandl. Med.-Chem. Forschungsstaetten Farbwerke Hoechst A.-G. 7 (1963) 296–314.
- [47] B. Hokfelt, A. Joansson, Hypoglycemic activity in relation to chemical structure of potential oral antidiabetic substances. I. 1-Sulfonyl3-alkylureas, J. Med. Pharm. Chem. 5 (1962) 231–239.
- [48] W. Logemann, L. Caprio, D. Artini, Hypoglycemizing sulfonamides, Farmaco, Ed. Sci. 12 (1957) 586–593.
- [49] P.A. Petyunin, V.P. Chernykh, I.P. Bannyi, Z.S. Spesivtseva, Arenesulfonylureas and arenesulfonyloxamides with sugar-reducing activity. XX. Oxalic acid amides and ureides, Khim.-Farm. Zh. 6 (1972) 9–13.
- [50] M. Ceruso, S. Antel, A. Scozzafava, C.T. Supuran, Synthesis and inhibition potency of novel ureido benzenesulfonamides incorporating GABA as tumor-associated carbonic anhydrase IX and XII inhibitors, J. Enzyme Inhib. Med. Chem. 31 (2016) 205–211.
- [51] H.M. Faidallah, K.A. Khan, Synthesis and biological evaluation of new barbituric and thiobarbituric acid fluoro analogs of benzenesulfonamides as antidiabetic and antibacterial agents, J. Fluor. Chem. 142 (2012) 96–104.
- [52] M. Jalali-Heravi, A. Kyani, Application of genetic algorithm-kernel partial least square as a novel nonlinear feature selection method: activity of carbonic anhydrase II inhibitors, Eur. J. Med. Chem. 42 (2007) 649–659.
- [53] M. Pitea, A. Marie, V. Ariesan, C. Margineanu, Studies on some p-substituted benzenesulfonamide derivatives, Arch. Pharm. (Weinheim, Ger.) 309 (1976) 586–591.
- [54] J.S. Roth, E.F. Degering, Preparation of sulfanilamide derivatives containing a urea or thiourea grouping, J. Am. Chem. Soc. 67 (1945) 126–128.
- [55] J. Singh, B. Shaik, S. Singh, V.K. Agrawal, P.V. Khadikar, O. Deeb, C.T. Supuran, Comparative QSAR study on para-substituted aromatic sulphonamides as CAII inhibitors: information versus topological (distance-based and connectivity) indices, Chem. Biol. Drug Des. 71 (2008) 244–259.
- [56] B. Skowronska-Serafin, T. Urbanski, Preparation of amidino urea derivatives and their reactions, Tetrahedron 10 (1960) 12–25.
- [57] C.T. Supuran, A. Scozzafava, B.C. Jurca, M.A. Ilies, Carbonic anhydrase inhibitors. Part 49: synthesis of substituted ureido and thioureido derivatives of aromatic/heterocyclic sulfonamides with increased affinities for isoenzyme I, Eur. J. Med. Chem. 33 (1998) 83–93.
- [58] D. Vullo, W. Leewattanapasuk, F.A. Muhlschlegel, A. Mastrolorenzo, C. Capasso, C.T. Supuran, Carbonic anhydrase inhibitors: Inhibition of the  $\beta$ -class enzyme from the pathogenic yeast *Candida glabrata* with sulfonamides, sulfamates and sulfamides, Bioorg. Med. Chem. Lett. 23 (2013) 2647–2652.