



3D-QSAR pharmacophore modelling, virtual screening and docking studies for lead discovery of a novel scaffold for VEGFR 2 inhibitors: Design, synthesis and biological evaluation

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ABSTRACT

A series of novel 6,7-dihydro-5H-cyclopenta[d]pyrimidine derivatives was successfully designed, synthesized and evaluated as a new chemical scaffold with vascular endothelial growth factor receptor (VEGFR 2) inhibitory activity. Compounds **6c** and **6b** showed enzyme inhibition of 97% and 87% at 10 μ M, respectively, and exhibited potent dose-related VEGFR 2 inhibition with IC₅₀ values of 0.85 μ M and 2.26 μ M, respectively. The design of the 6,7-dihydro-5H-cyclopenta[d]pyrimidine scaffold was implemented via consecutive molecular modelling protocols prior to the synthesis and biological evaluation of the derivatives. First, sorafenib was docked in the binding site of VEGFR 2 to study its binding orientation and affinity, followed by the generation of a valid 3D QSAR pharmacophore model for use in the virtual screening of different 3D databases. Structures with promising pharmacophore-based virtual screening results were refined using molecular docking studies in the binding site of VEGFR 2. A novel scaffold was designed by incorporating the results of the pharmacophore model generation and molecular docking studies. The new scaffold showed hydrophobic interactions with the kinase front pocket that may be attributed to increasing residence time in VEGFR 2, which is a key success factor for ligand optimization in drug discovery. Different derivatives of the novel scaffold were validated using docking studies and pharmacophore mapping, where they exhibited promising results as VEGFR 2 inhibitors to be synthesized and biologically evaluated. 6,7-dihydro-5H-cyclopenta[d]pyrimidine is a new scaffold that can be further optimized for the synthesis of promising VEGFR 2 inhibitors.

1. Introduction

Vascular endothelial growth factor receptor (VEGFR) is a member of the receptor tyrosine kinase (RTK) family. It is essential for the growth of blood vessels, either, new (vasculogenesis) or pre-existing blood vessels (angiogenesis) [1]. VEGFR is activated by the binding of vascular endothelial growth factors (VEGFs) to its extracellular domain, causing dimerization of the receptor [2]. This conformational change makes the ATP binding site of the receptor available for ATP binding, causing phosphorylation of the tyrosine residue of the receptor and subsequent activation [2,3]. There are three subtypes of VEGFRs: VEGFR 1, 2 and 3. VEGFR 1 (Flt 1) is critical for the regulation of macrophage and monocyte migration, VEGFR 2 (KDR) is responsible for vascular endothelial cell's normal and pathological developments, and VEGFR 3 (Flt 4) is responsible for the development of lymphatic endothelial cells and the spreading of cancerous cells to lymph nodes [4]. Since VEGFR 2 is the subtype responsible for angiogenesis and vasculogenesis [3], the inhibition of VEGFR 2 will affect the blood supply to tumour cells, inhibiting their growth, proliferation and metastasis

[5,6]. Inhibition of the VEGFR signalling pathway is a crucial therapeutic target for tumour inhibition. Targeting the neovascularization process of cancer cells will promote their starvation and death in a short time due to their rapid growth rate.

VEGFR small molecule inhibitors are categorized into two types according to the conformation of the receptor. Type I inhibitors compete with ATP molecules at the ATP binding site to bind to the receptor in its active form, adopting the Asp-Phe-Gly (DFG)-in conformation and consequently preventing the phosphorylation and activation of the receptor. In contrast, Type II inhibitors are non-ATP competitors that bind and stabilize the inactive form of the receptor, the DFG-out conformer, which is achieved through flipping of the DFG motif of the receptor to reveal an extra hydrophobic pocket [6,7]. Several molecules have been approved by the FDA for the treatment of different types of cancer [9]; for example, sorafenib is an oral multikinase dual-acting inhibitor that affects tumour cells directly through inhibiting the rapidly accelerated fibrosarcoma (RAF)/mitogen-activated protein kinase (MEK)/extracellular signal-regulated kinase (ERK) signalling pathway or targeting the tumour vasculature by inhibiting platelet-derived growth factor

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receptor-B (PDGFR-B) and VEGFR [9]. Sorafenib is a type II kinase inhibitor with a reported IC_{50} value of $0.057 \mu\text{M}$ for PDGFR-B and $0.09 \mu\text{M}$ for VEGFR 2 [10].

Several drug design studies were carried out with the aim of improving the potency and pharmacokinetic properties of existing VEGFR inhibitors by structural modification or finding new candidates for VEGFR inhibition. In this study, a novel 6,7-dihydro-5H-cyclopenta[d]pyrimidine scaffold was designed using consecutive computational protocols. The potency and effectiveness of the proposed derivatives as VEGFR 2 inhibitors were validated using molecular docking and pharmacophore mapping protocols. Then, the proposed derivatives were synthesized. Their VEGFR 2 inhibitory activity was evaluated, and their dose-related IC_{50} values were measured and compared to that of sorafenib as a reference compound.

2. Rationale

2.1. Molecular modelling studies

Computer-aided drug design (CADD) adopts different computational techniques to optimize commercially available drugs into new ligands with potential biological activities against certain biological targets. CADD is based on quantum mechanics and molecular modelling techniques and is used to reduce the expenses and time consumed by rational drug design.

In the current study, a novel 6,7-dihydro-5H-cyclopenta[d]pyrimidine scaffold was designed using different molecular modelling protocols: the molecular docking of sorafenib as a potent VEGFR 2 inhibitor to understand binding affinity and orientation inside the binding site of VEGFR 2, 3D QSAR (Three Dimensional Quantitative Structure-Activity Relationship) pharmacophore model generation and virtual screening of 3D databases using Accelrys Discovery Studio software version 4.5 (DS 4.5, Accelrys Ltd., UK). The 3D QSAR pharmacophore model generation protocol builds a pharmacophore model using a set of ligands with known activity to identify the essential functional features contributing to their high potency and biological activity. The generated pharmacophore model was used for the virtual screening of three 3D databases: the DruglikeDiverse, MiniMaybridge, and scPDB databases. The selected hits of the virtual screening were analysed and subjected to molecular docking into the crystal structure of VEGFR 2 with protein data bank code 3WZE in complex with sorafenib as a reference inhibitor. The good correlation between docking scores and pharmacophore fit values provided a reliable basis for designing new VEGFR 2 inhibitors.

2.1.1. Molecular docking of sorafenib

A molecular docking protocol was implemented using the CDOCKER protocol under the receptor ligand interaction protocol of Accelrys Discovery Studio 4.5. CDOCKER is a grid-based molecular docking method that uses the CHARMM force field-based molecular dynamics (MD) search algorithm to dock ligands into the binding site of the receptor. Different conformations of the ligand are generated with different scoring functions, such as -CDOCKER_ENERGY (CHARMM energy of the interaction energy and ligand strain) and -CDOCKER_INTERACTION_ENERGY (interaction energy only).

The determination of the binding interactions and orientation of VEGFR 2 kinase inhibitors was studied using the X-ray crystal structure of VEGFR 2 with sorafenib (PDB code 3WZE). The molecular docking protocol (CDOCKER) of Accelrys Discovery Studio 4.5 was used, and the target protein was prepared and the active binding site identified based on the positional coordinates of the co-crystallized inhibitor. The results of the CDOCKER protocol were validated by re-docking of the co-crystallized structure of the reference sorafenib inside the active site of VEGFR 2 (PDB code 3WZE). The root mean square deviation (RMSD) between the re-docked conformer and the co-crystallized conformer of sorafenib is 0.257, (Fig. 1), which confirms the validity of the docking

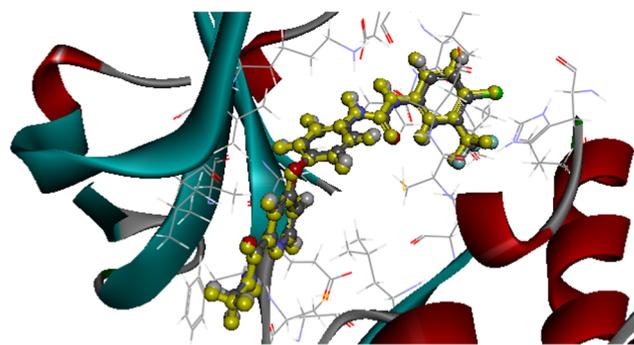


Fig. 1. 3D image of the superimposition of the re-docked conformer of sorafenib over the co-crystallized conformer (colored yellow) with RMSD value of 0.257. (For interpretation using colors, the reader is referred to the web version of this article).

protocol.

Molecular docking simulation provides insight into the binding interaction and affinity between the compound and the receptor. Promising biological activity is indicated by lower CDOCKER energy and similar binding interactions to that of the reference co-crystallized inhibitor. A docking study of sorafenib resulted in a binding energy score of -48.92 with a total of ten hydrogen bonds. The essential amino acid Cys919 located in the hinge region binds to the nitrogen of the pyridine ring, the NH and the methyl group of the amide moiety. In addition, Glu917 in the hinge region binds to the CH proton at the 6-position of the pyridine ring; the urea moiety binds to the receptor through various hydrogen bonding interactions where both urea NH groups interact with Glu885 in the C-helix; and the carbonyl group interacts with Asp1046 in the DFG motif and with Cys1045. The fluoride atoms of the external trifluoromethyl moiety interact with His1026 and Cys1045. Furthermore, the pyridine moiety forms hydrophobic interactions with a hydrophobic pocket formed by amino acid residues Cys919, Leu1035, Ala866, Val848 and Leu840, and the phenyl ring binds to another hydrophobic pocket through interactions with Lys868, Cys1045, Val916, Phe1047 and Val848. In addition, the terminal phenyl ring forms hydrophobic bond with Leu889, the meta trifluoromethyl substitution forms hydrophobic interactions with Val898, Ile1044, His1026 and Leu1019, and the para chloro substituent exhibits hydrophobic interactions with Leu1019 and Ile888 (Table 4). The resulting interactions resemble the reported binding mode of sorafenib in the X-ray crystal structure of VEGFR 2 with PDB code 3WZE [8], (Fig. 2).

2.1.2. 3DQSAR pharmacophore model generation and validation

2.1.2.1. Assortment of anticancer library. A set of 25 VEGFR 2 inhibitors of known activity with IC_{50} values ranging from 0.027 to $9.4 \mu\text{M}$ were collected from the literature [11], (Fig. 3). Library ligands were prepared using the prepare ligands protocol of Accelrys Discovery Studio 4.5. Ligand preparation is essential for energy minimization, the correction of incorrect valences and the protonation of ligands.

2.1.2.2. 3D QSAR pharmacophore model generation. 3D QSAR pharmacophore is an advanced common feature pharmacophore ligand-based protocol of Discovery Studio 4.5 that is used to explore essential pharmacophore features from a set of ligands with known activity against the biological target of interest. Pharmacophore model generation is implemented using the HypoGen algorithm [8,9], which considers only features that are common to active compounds, while common features among inactive compounds are excluded [13]. For the generation of potential pharmacophore models, a feature-mapping protocol was initiated to identify important chemical features imbedded within the training set compounds (T1-15). Subsequently, a pharmacophore model was generated using twenty-five compounds

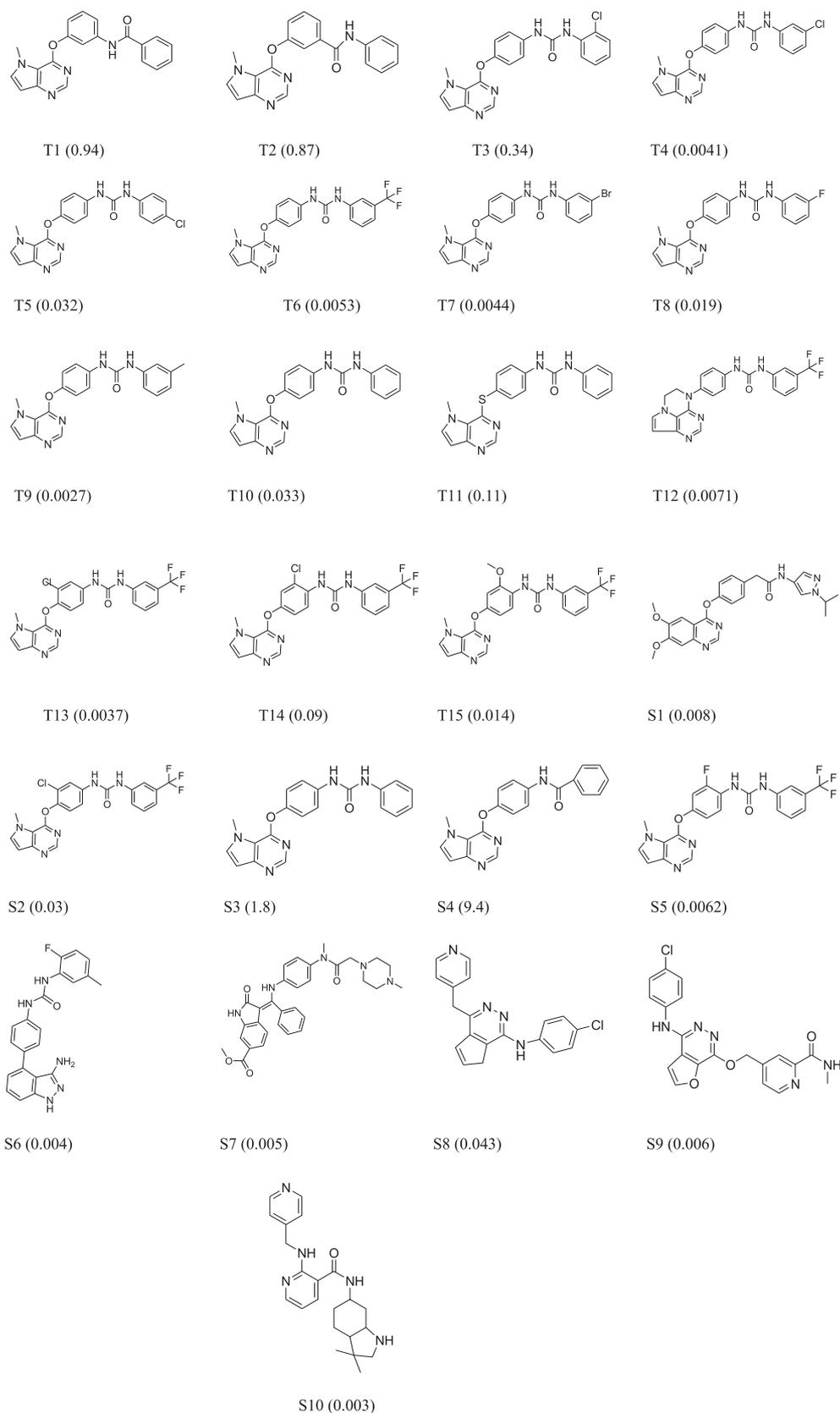


Fig. 3. The library of T1-15 training set and S15-25 test set of known active VEGFR 2 inhibitors used for pharmacophore model generation along with their IC₅₀ values (μM) indicated in parenthesis.

Val916, Val848 and Ala866 and the interaction of the phenyl ring attached to the thiazole ring with Lys868, Cys1045, Val899 and Val916. In addition, hydrophobic interactions between the terminal phenyl ring

and the Leu889 amino acid were observed (**Table 3**).

2r4b_GW7, with a fit value of 7.10 and a CDOCKER binding score of -35.08, binds to the receptor through interaction of the N1 of the

Table 1
Tabular column illustrating the details of the ten hypotheses generated using hypoGen algorithm.

Hypothesis	Total cost	Cost difference	RMS	Correlation coefficient	Features ^b	Maximum fit
1	59.35	140.98	1.03	0.97	HBA, HYP, HYP, HYP, RA	10.80
2	66.56	133.78	1.42	0.95	HBA, HYP, HYP, HYP	7.36
3	74.56	125.78	1.76	0.92	HBD, HYP, HYP, HYP	7.81
4	74.86	125.48	1.77	0.92	HBD, HYP, HYP, HYP	7.90
5	74.95	125.39	1.76	0.92	HYP, HYP, HYP, HYP, HYP	8.83
6	75.82	124.52	1.74	0.92	HBA, HBA, HYP, HYP, HYP	7.08
7	76.17	124.16	1.78	0.92	HBA, HBD, HYP, RA	6.23
8	76.54	123.80	1.80	0.92	HBD, HYP, HYP, HYP	6.39
9	78.23	122.10	1.89	0.91	HBA, HYP, HYP, RA	7.34
10	78.97	121.37	1.89	0.91	HBD, HYP, RA, RA	6.41

^b Abbreviations used for features; hydrogen bond acceptor (HBA), hydrophobic (HYP) and ring aromatic (RA).

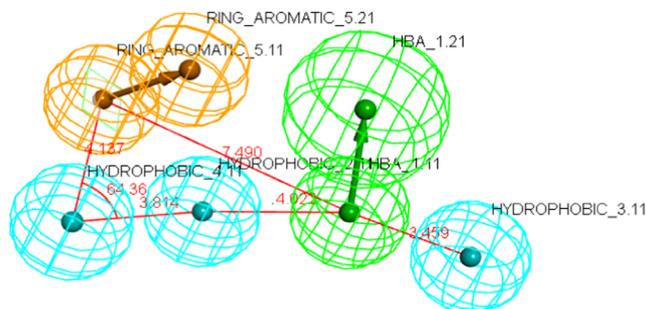


Fig. 4. The best generated Pharmacophore model with three features: hydrogen bond acceptor (HBA) colored green, ring aromatic (RA) colored orange and hydrophobic (HYP) colored cyan. With their inter-feature distances in angstroms and angles displayed.

Table 2
The experimental and predicted activity values of the training set compounds according to hypothesis 1 along with their fit values.

Training set	Experimental activity IC ₅₀ nM	Predicted activity IC ₅₀ nM	Fit value
T1	940	688.787	6.505
T2	870	1299	6.230
T3	340	349.804	6.800
T4	4.1	8.839	8.397
T5	32	13.729	8.206
T6	5.3	5.586	8.596
T7	4.4	8.998	8.389
T8	19	12.989	8.230
T9	2.7	3.086	8.854
T10	33	33.111	7.823
T11	110	113.757	7.287
T12	7.1	7.335	8.478
T13	30	14.713	8.176
T14	3.7	3.033	8.861
T15	14	17.234	8.107

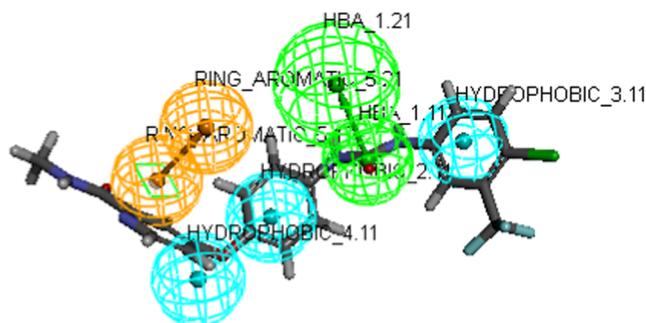


Fig. 5. Sorafenib as reference compound mapped into the best pharmacophore model with fit value of 6.19.

pyrimidine ring with Cys919 in the hinge region and binding of the CH proton at the 2-position of the pyrimidine ring with Glu917. Cys919 and Glu917 are key amino acids for the hydrogen bonding of the adenine ring of ATP to the hinge region of VEGFR 2 [15]. The hydrophobic interactions of **2r4b_GW7** involve the binding of the thienyl ring to the kinase front pocket formed by residues Phe918, Leu840, Leu1035 and Ala866 and the hydrophobic binding of the pyrimidine ring to residues Leu1035, Cys919 and Ala866. In addition, the phenyl ring binds to Val916, Cys1045, Val848, Phe1047 and Lys868, the 3-chloro substituent of the phenyl ring forms hydrophobic interactions with Lys868 and Val916, and the terminal phenyl ring forms hydrophobic interactions with Leu889 (Table 3). The three hits showed consistency in rendering the interactions of VEGFR 2 inhibitors.

2.2. Design strategy

In studying the structure-activity relationship of reported VEGFR 2 inhibitors and analysing their binding modes, three main features of type II inhibitors of VEGFR 2 were reported: most inhibitors include a flat heteroaromatic ring system that binds to the NH of Cys919 in the hinge region; a hydrogen bond donor and acceptor pair, which can be a urea or an amide group that binds with Glu885 and Asp1046 in the enzyme DFG motif, where two hydrogen bonds are formed between NH of the urea or amide group and Glu885 and another hydrogen bond is between the CO group and Asp1046 [16]; and finally, the binding of the terminal aryl group in the newly created allosteric hydrophobic pocket exposed upon flipping of the DFG loop, as shown in the inactive conformation or DFG-out conformation of the enzyme [17].

Compound **2r4b_GW7** has a fit value of 7.10 and exhibits the 5 features (1HBA, 3HYP and 1RA) of the validated pharmacophore model, while sorafenib has a fit value of 6.19 and exhibits only 4 features (1HBA, 3HYP), (Table 4). Molecular docking studies of the hit compound **2r4b_GW7** revealed a hydrogen bond between N1 of the thienyl pyrimidine moiety and Cys919 and a hydrogen bond between the CH proton at the 2-position of the pyrimidine ring and Glu917. Both Cys919 and Glu917 are key amino acids for the hydrogen bonding of ATP to the hinge region of VEGFR 2 [15]. The thienyl pyrimidine moiety forms additional hydrophobic interactions at the front pocket of the kinase domain with residues Leu840, Ala866, Phe918 and Leu1035, (Table 4). Hydrophobic interaction with the kinase front pocket complements the hydrogen bonding with the hinge region, increasing the residence time [8], which is a key factor in improving potency *in vivo* and enhancing the correlation between the *in vitro* and *in vivo* efficacy of drugs [18].

Utilizing the aforementioned essential structural features, bioisosteric modification strategies and the results of the 3D QSAR pharmacophore model generation and molecular modelling protocols, a proposed scaffold of 6,7-dihydro-5H-cyclopenta[d]pyrimidine derivatives was designed and synthesized. The design of the new scaffold was based on the replacement of the *N*-methylpicolinamide moiety of sorafenib with a 6,7-dihydro-5H-cyclopenta[d]pyrimidine moiety. Bioisosteric

Table 4
Docking results of sorafenib and the three top hits KM10259, CDI665171 and 2r4b_GW7 in the active site of VEGFR 2 (3WZE) showing hydrogen bonds and hydrophobic interactions.

Name	Hydrogen bonds	Hydrogen bond length Å ⁰	Hydrophobic interactions
Sorafenib	O of Cys919 – H of CH ₃	2.5	Leu840
	O of Cys919 – H of NH	2.2	Val848
	NH of Cys919 – N1 of pyridine	2.3	Ala866
	O of Glu917 – C2 of pyridine	2.4	Cys919
	O of Glu885 – H of urea NH	2.0	Leu1035
	O of Glu885 – H of urea NH	2.1	Val848
	NH of Asp1046 – O of urea CO	1.8	Lys868
	H of Cys1045 – O of urea CO	2.7	Val916
	H of His1026 – F in terminal phenyl ring	2.7	Cys1045
	H of Cys1045 – F in terminal phenyl ring	2.5	Phe1047
			Leu889 – terminal phenyl ring
			Ile888
			Leu1019
		Val898	
		Leu1019	
		His1026	
		Ile1044	
KM10259	O of Glu885 – H of urea NH	2.1	Val848
	O of Glu885 – H of urea NH	2.1	Ala866
	NH of Asp1046 – O of urea CO	1.9	Cys919
	H of Cys1045 – O of urea CO	2.8	Leu1035
			Leu889
		Val898	
		Leu1019	
		Ile1044	
CDI665171	O of Glu885 – H of urea NH	1.9	Leu1035
	O of Glu885 – H of urea NH	3.0	Cys919
	NH of Asp1046 – O of urea CO	2.0	Phe918
	H of Cys1045 – O of urea CO	2.3	Leu840
			Leu1035
		Cys1045	
		Val916	
		Val848	
		Ala866	
		Lys868	
		Cys1045	
		Val899	
		Val916	
		Leu889 – terminal phenyl ring	
2r4b_GW7	NH of Cys919 – N1 of pyrimidine	2.5	Leu840
	O of Glu917 – C2 of pyrimidine	2.2	Ala866
			Phe918
			Leu1035
			Ala866
			Cys919
			Leu1035
			Val848
			Lys868
			Val916
			Cys1045
			Phe1047
			Lys8683
			Val916
			Leu889 – terminal phenyl ring

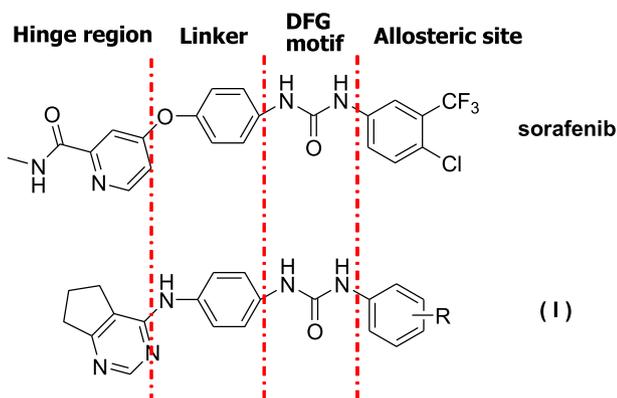


Fig. 6. Illustration of binding of sorafenib and the proposed scaffold (I) to the binding regions of VEGFR 2.

replacement was carried out by the substitution of the oxygen linker with a classical NH bioisostere, (Fig. 6). Introducing new substituents at different positions of an aromatic ring of a hit compound may optimize the binding affinity of the drug with the receptor in addition to conferring the physicochemical properties essential for drug distribution and metabolism. Different derivatives of (I) were proposed with alternative lipophilic substituents at the terminal aromatic ring to occupy the hydrophobic allosteric pocket. Substituents were either electron withdrawing, such as chloro and trifluoromethyl groups, or electron donating, such as methyl and methoxy groups.

2.2.1. Validation of the proposed scaffold of VEGFR 2 inhibitors

2.2.1.1. Pharmacophore mapping of the proposed VEGFR 2 inhibitors. The activity of the proposed scaffold as an effective VEGFR 2 inhibitor was validated through mapping of the proposed derivatives into the validated pharmacophore model. The results showed that the proposed derivatives **6a**, **6b**, **6c**, **6d** and **6e** exhibited five features of the generated pharmacophore model with fit values ranging from 6.63 to 9.21 (Table 5), which are higher than that of the reference

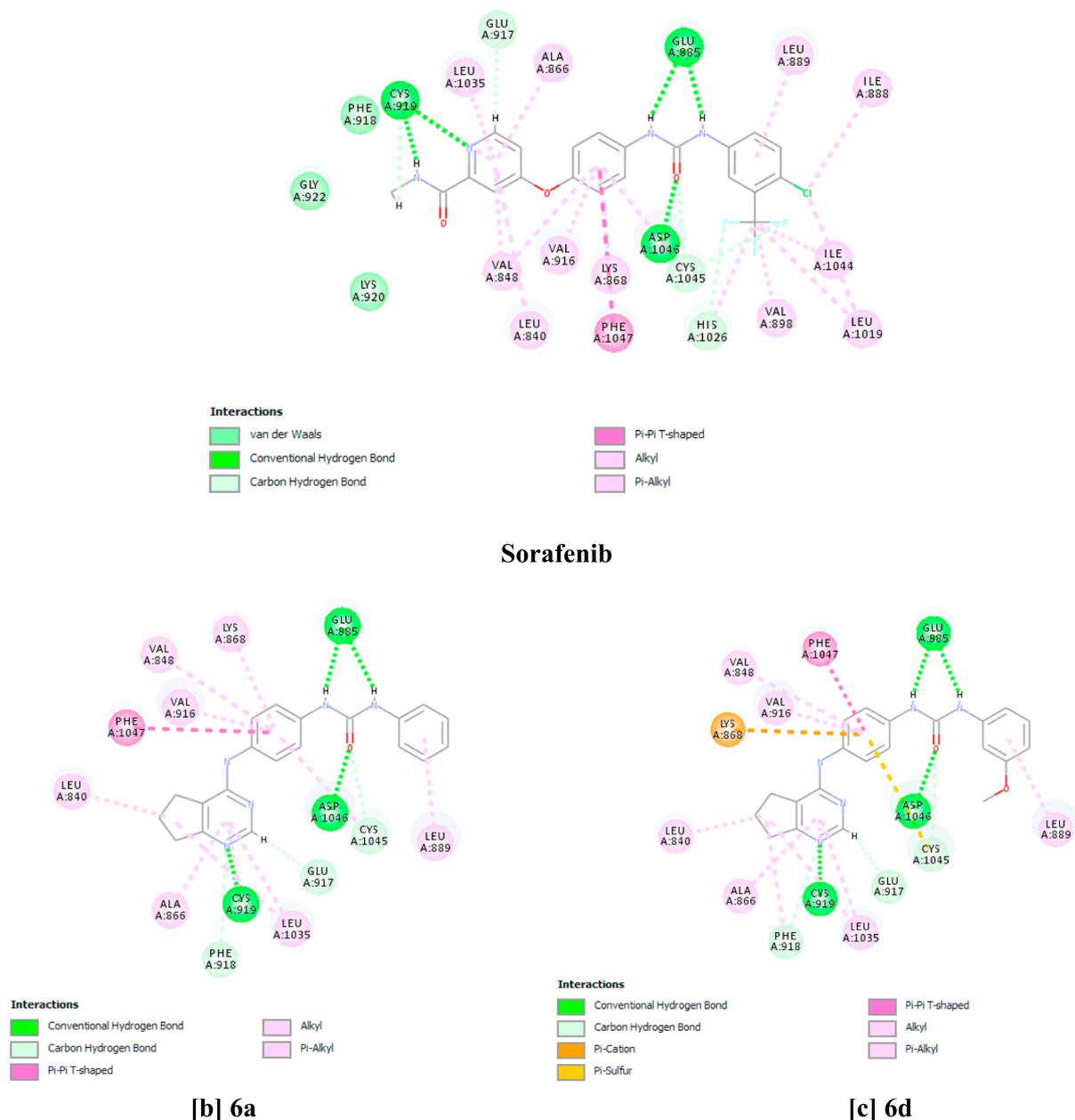


Fig. 7. 2D image of hydrogen bonds and hydrophobic interactions of sorafenib and proposed derivatives 6a and 6d with the kinase domain of VEGFR 2.

were synthesized and subjected to biological evaluation against the VEGFR 2 enzyme. Compounds **6b** and **6c** exhibited high percentage inhibition of 87% and 97%, respectively, at 10 μM . The compounds with the highest enzyme inhibitory effect were tested for VEGFR 2 inhibition and demonstrated IC_{50} values in the sub-micromolar range. The IC_{50} values of compounds **6c** and **6b** were 0.85 μM and 2.26 μM , respectively. Therefore, compounds **6c** and **6b** represent a new molecular scaffold that can be further optimized and used for the design of promising potent VEGFR 2 inhibitors.

5. Experimental

Starting material and reagents were purchased from Sigma Aldrich or Alfa-Aeser organics and were used without further purification. Melting points were recorded using BUCHI B-540 apparatus and were uncorrected. Reactions were monitored using thin layer chromatography (TLC) purchased from Merck and performed on 0.255 mm silica gel plates, with visualization under U.V. light (254 nm). The

hydrogenation process was carried out using hydrogenator (Parr Shaker) apparatus. FT-IR spectra were recorded on a Perkin Elmer FT-IR spectrophotometer and ^1H NMR spectra were recorded on BRUCKER 400 MHz spectrophotometer at Ain Shams University. Mass spectra were recorded on Thermo Scientific ISQ LT gas chromatograph mass spectrometer at the regional center for Mycology and Biotechnology Al-Azhar University and on thermos Q-exactive orbitrap instrument at laboratory for single cell mass spectrometry, quantitative biology center, Japan.

5.1. Chemistry

5.1.1. General procedure for the preparation of compounds (1a-e)

A solution of p-nitroaniline (1 g, 6 mmol: 1 equiv.) in dry dichloromethane (20 mL), was stirred with the appropriate isocyanate (6 mmol: 1 equiv.) at room temperature for 48 h. The resulting mixture was filtered and the filtered solid was crystallized from ethanol to yield derivatives (1a-e) [19].

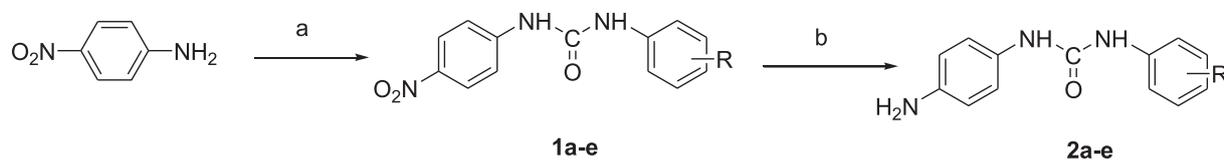
Table 6
Docking results of sorafenib and the three top hits KM10259, CDI665171 and 2r4b_GW7 in the active site of VEGFR 2 (3WZE) showing hydrogen bonds and hydrophobic interactions.

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	O of Glu885 – H of urea NH	2.1	Val848
	NH of Asp1046 – O of urea CO	1.8	Lys868
	H of Cys1045 – O of urea CO	2.7	Val916
	H of His1026 – F in terminal phenyl ring	2.7	Cys1045
	H of Cys1045 – F in terminal phenyl ring	2.5	Phe1047
			Leu889 – terminal phenyl ring
			Ile888 } Cl in terminal phenyl ring
			Leu1019
			Val898
		Leu1019 } CF ₃ in terminal phenyl ring	
		His1026	
		Ile1044	
6a	NH of Cys919 – N1 of pyridine	2.3	Leu1035
	H of Phe918 – N1 of pyridine	2.8	Leu840
	O of Glu917 – C2 of pyridine	2.2	Ala866
	O of Glu885 – H of urea NH	2.0	Leu1035
	O of Glu885 – H of urea NH	2.0	Cys919
	NH of Asp1046 – O of urea CO	1.8	Val848
	H of Cys1045 – O of urea CO	2.6	Lys868
			Val916 } phenyl ring
		Cys1045	
		Phe1047	
		Leu889 – terminal phenyl ring	
6b	NH of Cys919 – N1 of pyridine	2.3	Leu1035
	H of Phe918 – N1 of pyridine	2.8	Leu840
	O of Glu917 – C2 of pyridine	2.2	Phe918
	O of Glu885 – H of urea NH	2.0	Ala866
	O of Glu885 – H of urea NH	2.0	Leu1035
	NH of Asp1046 – O of urea CO	1.8	Cys919
	H of Cys1045 – O of urea CO	2.7	Val848
			Lys868
			Val916 } phenyl ring
			Cys1045
		Phe1047	
		Leu889 – terminal phenyl ring	
		Leu1019	
		His1026 } 3-CL	
		Ile1044	
		Val898	
		Ile888 } C of 4-CH ₃	
		Leu1019	
6c	NH of Cys919 – N1 of pyridine	2.3	Leu1035
	H of Phe918 – N1 of pyridine	2.8	Leu840
	O of Glu917 – C2 of pyridine	2.2	Phe918
	O of Glu885 – H of urea NH	2.1	Ala866
	O of Glu885 – H of urea NH	2.0	Leu1035
	NH of Asp1046 – O of urea CO	1.9	Cys919
	H of Cys1045 – O of urea CO	2.7	Val848
			Lys868
			Val916 } phenyl ring
			Cys1045
		Phe1047	
		Leu889 – terminal phenyl ring	
		Leu1019 – 4-CL	
6d	NH of Cys919 – N1 of pyridine	2.3	Leu1035
	H of Phe918 – N1 of pyridine	2.8	Leu840
	O of Glu917 – C2 of pyridine	2.2	Phe918
	O of Glu885 – H of urea NH	2.0	Ala866
	O of Glu885 – H of urea NH	2.0	Leu1035
	NH of Asp1046 – O of urea CO	1.8	Cys919
	H of Cys1045 – O of urea CO	2.6	Val848
			Lys868
			Val916 } phenyl ring
			Cys1045
		Phe1047	
		Leu889 – terminal phenyl ring	
6e	NH of Cys919 – N1 of pyridine	2.3	Leu1035
	H of Phe918 – N1 of pyridine	2.8	Leu840
	O of Glu917 – C2 of pyridine	2.2	Ala866
	O of Glu885 – H of urea NH	2.1	Leu1035
	O of Glu885 – H of urea NH	2.0	Cys919
	NH of Asp1046 – O of urea CO	1.8	Val848
	H of Cys1045 – O of urea CO	2.7	Lys868
	H of His1026 – F of 3-CF ₃	2.8	Val916
	H of Cys1045 – F of 3-CF ₃	2.6	Cys1045
			Phe1047
		Leu889 – terminal phenyl ring	
		Leu1019 } C of 3-CF ₃	
		His1026	

5.1.1.1. 1-(4-Nitrophenyl)-3-phenylurea (**1a**). Yield 40% as yellowish white crystals, m.p 226–228 °C, (as reported) [24].

5.1.1.2. 1-(3-Choloro-4-methylphenyl)-3-(4-nitrophenyl)urea (**1b**). Yield 50% as yellow orange crystals, m.p 182–185 °C, (as reported) [10].

5.1.1.3. 1-(3-trifluoromethyl-4-chlorophenyl)-3-(4-nitrophenyl)urea (**1c**). Yield 90% as pale yellow crystals, m.p 238–240 °C. FT-IR (ú max, cm⁻¹): 1596 (C=C), 1646 (C=N), 1717 (C=O), 3128 (CH aromatic), 3352 (NH).



1a, 2a ; R= H
1b, 2b ; R= 3-Cl, 4-CH₃
1c, 2c ; R= 4-Cl, 3-CF₃
1d, 2d ; R= 3-OCH₃
1e, 2e ; R= 3-CF₃

Scheme 1. Synthesis of 4-aminophenyl substituted phenyl ureas reagents and conditions: (a) Phenyl isocyanates, DCM, rt, 48 hrs; (b) H₂, Pd/C, MeOH, rt, 4 hrs.

5.1.1.4. *1-(3-Methoxyphenyl)-3-(4-nitrophenyl)urea (1d)*. Yield 47% as yellow crystals, m.p 210–212 °C, (as reported) [24].

5.1.1.5. *1-(4-Nitrophenyl)-3-(3-(trifluoromethyl)phenyl)urea (1e)*. Yield 60% as greenish yellow crystals, m.p 260 °C. FT-IR (ν max, cm⁻¹): 1575 (C=C), 1621 (C=N), 1720 (C=O), 3162 (CH aromatic), 3356 (NH).

5.1.2. General procedure for the preparation of compounds (2a–e)

Pd-C (0.1 g, 10%) was added to a solution of the appropriate nitro phenyl urea derivative (1a–e) (4 mmol) in methanol (100 mL), the mixture was stirred under H₂ at room temperature, at 60 bar for 4 h. The catalyst was removed by filtration over celite. Then the filtrate was concentrated and dried to afford crystals of compounds (2a–e) which were then recrystallized from methanol [20].

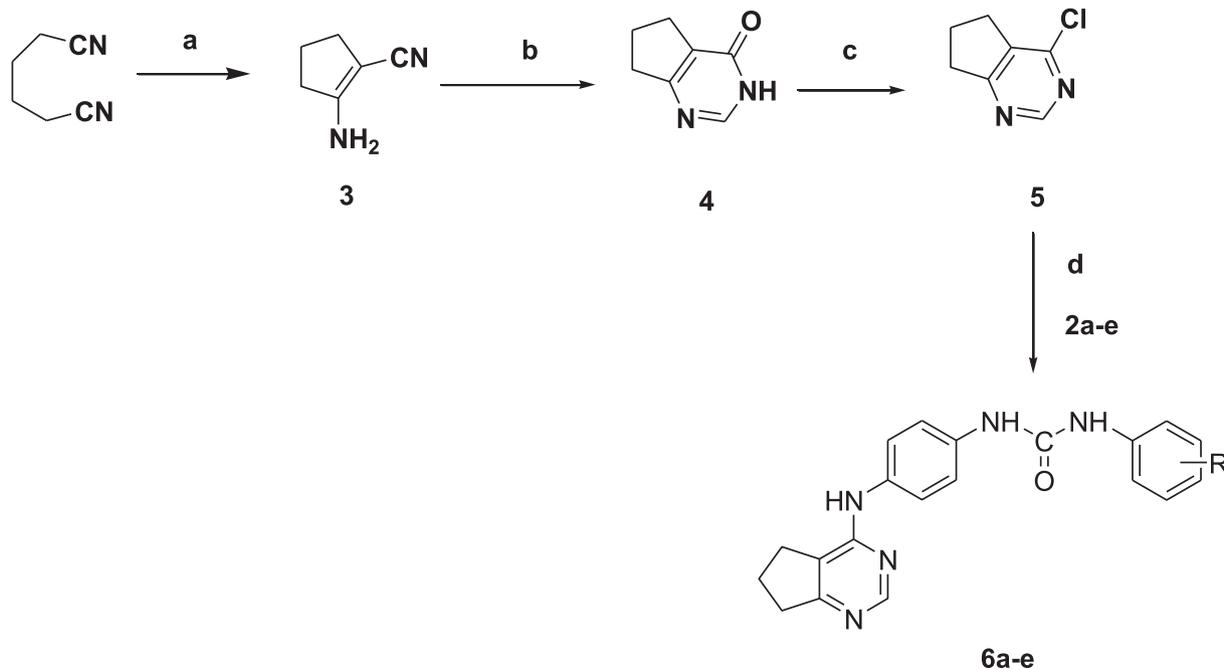
Table 7
VEGFR 2 enzyme inhibition of the synthesized compounds at 10 μ M.

Compounds	R	% inhibition
6a	H	5
6b	3-Cl-4-CH ₃	87
6c	4-Cl-3-CF ₃	97
6d [†]	3-OCH ₃	0
6e	3-CF ₃	1

* These compounds are sparingly dissolved at 10 mM in 100% DMSO.

5.1.2.1. *1-(4-Aminophenyl)-3-phenylurea (2a)*. Yield 70% as white crystals, m.p 224–227 °C, (as reported) [25].

5.1.2.2. *1-(4-Aminophenyl)-3-(3-Chloro-4-methylphenyl)urea (2b)*. Yield 80% as grey crystals, m.p 203–205 °C, (as reported) [10].



6a ; R= H
6b ; R= 3-Cl, 4-CH₃
6c ; R= 4-Cl, 3-CF₃
6d ; R= 3-OCH₃
6e ; R= 3-CF₃

Scheme 2. Synthesis of 6,7-dihydro-5H-cyclopenta[d]pyrimidine derivatives reagents and conditions: (a) NaH, toluene, 15 hrs; (b) HCOOH, acetic anhydride, reflux, 48 hrs; (c) POCl₃, reflux, 3 hrs; (d) Ethanol, TEA, reflux, 18–48 hrs.

Table 8
The IC₅₀ value of Synthesized compounds with high VEGFR 2 percent inhibition.

Compounds	VEGFR 2 IC ₅₀ (μM)
6b	2.26
6c	0.85
Staurosporine	0.09
Sorafenib	0.09

5.1.2.3. *1-(4-Aminophenyl)-3-(3-trifluoromethyl-4-chlorophenyl)urea (2c)*. Yield 80% as grayish white crystals, m.p 187–190 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 6.52–6.54 (d, *J* = 8 Hz, 1H, H_{2'}), 7.07–7.09 (d, *J* = 8 Hz, 1H, H_{6'}), 7.58–7.60 (d, *J* = 8 Hz, 1H, H_{5''}), 7.67–7.71 (m, 2H, H_{3'}, H_{5'}), 8.10–8.12 (d, *J* = 8 Hz, 1H, H_{6''}), 8.19–8.31 (m, 1H, H_{2''}), 8.99 (s, 1H, NH, exchangeable by D₂O), 9.37 (s, 1H, NH, exchangeable by D₂O), 9.44 (s, 1H, NH, exchangeable by D₂O), 9.64 (s, 1H, NH, exchangeable by D₂O). Ms: (Mwt.: 329); *m/z*, 331 [M⁺ + 2, (30%)], 329 [M⁺, (100%)], 108 (77%), 80 (40%), 53 (24%).

5.1.2.4. *1-(4-Aminophenyl)-3-(3-Methoxyphenyl)urea (2d)*. Yield 70% as white crystals, m.p 165–168 °C, (as reported) [10].

5.1.2.5. *1-(4-Aminophenyl)-3-(3-(trifluoromethyl)phenyl)urea (2e)*. Yield 80% as grayish white crystals, m.p 147–149 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 4.80 (s, 2H, NH, exchangeable by D₂O), 6.51–6.53 (d, *J* = 8 Hz, 2H, H_{2'}, H_{6'}), 7.07–7.1 (d, *J* = 12 Hz, 2H, H_{3'}, H_{5'}), 7.25–7.27 (d, *J* = 8 Hz, 1H, H_{4''}), 7.46–7.54 (m, 1H, H_{5''}), 7.48–7.50 (d, *J* = 8 Hz, 1H, H_{6''}), 8.01 (s, 1H, H_{2''}), 8.27 (s, 1H, NH, exchangeable by D₂O), 8.88 (s, 1H, NH, exchangeable by D₂O). Ms: (Mwt.: 295); *m/z*, 296 [M⁺ + 1, (18%)], 295 [M⁺, (93%)], 108 (100%), 80 (55%), 53 (22%).

5.1.3. 2-aminocyclopent-1-ene carbonitrile (3)

A solution of adipodinitrile (10.8 g, 100 mmol) in toluene (100 mL) is added dropwise to a slurry of sodium hydride (60% suspension in oil, 4.20 g, 105 mmol) in anhydrous toluene (30 mL). The mixture was heated under reflux for 15 h. The product was extracted with ethyl acetate (3x50 ml). Then recrystallized by dissolving in hot toluene (25 mL) and then adding petroleum ether (150 mL). The product was obtained as yellowish white solid (6.60 g, 61%), m.p 148 °C [21].

5.1.4. 6,7-dihydro-3H-cyclopenta[d]pyrimidin-4(5H)-one (4)

Acetic anhydride (46 mL, 492.38 mmol: 23.8 equiv.) was added portion wise to a stirred formic acid (23 mL, 614.92 mmol: 29.8 equiv.) at 0 °C over 30 min. This was followed by addition of compound (3) (2 g, 20 mmol: 1 equiv.) and stirring under reflux at 130 °C for 48 h. The solvent was evaporated under vacuum and the resultant solid was washed with diethyl ether, dried and recrystallized from methanol. The product was separated as buff crystals (1.2 g, 60%), m.p 220 °C [26].

5.1.5. 4-chloro-6,7-dihydro-5H-cyclopenta[d]pyrimidine (5)

A mixture of compound (4) (2 g, 0.01 mmol: 1 equiv.) and phosphorous oxychloride (29 mL, 278 mmol: 18.2 equiv.) was heated under reflux for 3 h. The mixture was then slowly poured on ice/water, neutralized using ammonia solution (33%, 50 mL), then extracted with ethyl acetate (2x50 mL). The combined organic layer was separated and the solvent was evaporated under vacuum to afford brown solid that was washed with diethyl ether to give the titled compound (5) as brown crystals (1.5 g, 76%) which was used directly in the next reaction [27].

5.1.6. General procedure for the preparation of compounds (6a–e)

The appropriate 4-aminophenyl substituted phenyl urea (2a–e) (1 mmol: 1 equiv.) and TEA (0.3 mL, 2 mmol: 2 equiv.) were added to a solution of the 4-chloro cyclopenta[d]pyrimidine derivative 5 (0.25 g,

1 mmol: 1 equiv.) in ethanol (15 mL). The mixture was heated under reflux for 18–24 h and the reaction was followed up using TLC. The resultant solid was collected by filtration, washed with hot ethanol, allowed to dry and recrystallized from ethanol to give the titled compounds (6a–e).

5.1.6.1. *1-(4-(6,7-dihydro-5H-cyclopenta[d]pyrimidin-4-ylamino)phenyl)-3-phenylurea (6a)*. The product was separated as yellowish brown powder (0.2 g, 33%), m.p > 300 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.16–1.20 (t, 2H, cyclopentenyl H), 1.94–1.99 (m, 1H, cyclopentenyl H), 2.63–2.65 (m, 1H, cyclopentenyl H), 2.72–2.77 (m, 1H, cyclopentenyl H), 3.07–3.11 (m, 1H, cyclopentenyl H), 6.51–6.53 (d, *J* = 8 Hz, 2H, H_{2'}, H_{6'}), 6.90–7.01 (m, 1H, H_{4''}), 7.08–7.10 (d, *J* = 8 Hz, 2H, H_{3'}, H_{5'}), 7.22–7.47 (m, 4H, H_{2''}, H_{3''}, H_{5''}, H_{6''}), 8.32 (s, 1H, pyrimidine H), 8.41 (s, 1H, NH, exchangeable by D₂O), 8.66 (s, 1H, NH, exchangeable by D₂O), 8.90 (s, 1H, NH, exchangeable by D₂O). HRMS (ESI-MS) calcd: 346.16339 for C₂₀H₁₉N₅O [M + H⁺]. Found: 346.16672.

5.1.6.2. *1-(3-chloro-4-methylphenyl)-3-(4-(6,7-dihydro-5H-cyclopenta[d]pyrimidin-4-ylamino)phenyl)urea (6b)*. The product was separated as yellowish brown powder (0.102 g, 17%), m.p 233–235 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.09–1.13 (m, 1H, cyclopentenyl H), 1.86–1.91 (m, 1H, cyclopentenyl H), 2.04–2.12 (m, 1H, cyclopentenyl H), 2.26 (s, 3H, Ph-CH₃), 2.76–2.83 (m, 3H, cyclopentenyl H), 6.55–6.56 (d, 2H, H_{2'}, H_{6'}), 7.00–7.58 (m, 5H, ArH), 8.37 (s, 1H, pyrimidine H), 8.68 (s, 1H, NH, exchangeable by D₂O), 8.70 (s, 1H, NH, exchangeable by D₂O), 8.81 (s, 1H, NH, exchangeable by D₂O). HRMS (ESI-MS) calcd: 394.14333 for C₂₁H₂₀N₅OCl [M + H⁺]. Found: 394.14331.

5.1.6.3. *1-(4-chloro-3-(trifluoromethyl)phenyl)-3-(4-(6,7-dihydro-5H-cyclopenta[d]pyrimidin-4-ylamino)phenyl)urea (6c)*. The product was separated as yellow powder (0.11 g, 19%), m.p 288–290 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.22 (s, 1H, cyclopentenyl H), 1.98–2.06 (m, 2H, cyclopentenyl H), 2.76–2.79 (m, 3H, cyclopentenyl H), 7.29 (s, 1H, ArH), 7.40–7.42 (d, *J* = 8 Hz, 1H, H_{5''}), 7.57–7.61 (m, 4H, ArH), 7.64–7.66 (d, *J* = 8 Hz, 1H, H_{6''}), 8.15 (s, 1H, pyrimidine H), 8.71 (s, 1H, NH, exchangeable by D₂O), 9.37 (s, 1H, NH, exchangeable by D₂O), 9.94 (s, 1H, NH, exchangeable by D₂O). HRMS (ESI-MS) calcd: 448.11507 for C₂₁H₁₇N₅OClF₃ [M + H⁺]. Found: 448.11562.

5.1.6.4. *1-(4-(6,7-dihydro-5H-cyclopenta[d]pyrimidin-4-ylamino)phenyl)-3-(3-methoxyphenyl)urea (6d)*. The product was separated as grayish white powder (0.13 g, 22%), m.p > 300 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.24 (s, 4H, cyclopentenyl H), 1.91 (s, 2H, cyclopentenyl H), 3.50 (s, 3H, OCH₃, below water of DMSO), 6.86–6.88 (d, *J* = 8 Hz, 1H, H_{2'}), 6.96–6.97 (d, *J* = 4 Hz, 1H, H_{6'}), 7.04–7.06 (d, *J* = 8 Hz, 1H, H_{4''}), 7.21–7.23 (d, *J* = 8 Hz, 1H, H_{3'}), 7.47–7.50 (d, *J* = 12 Hz, 1H, H_{6''}), 7.62–7.65 (d, *J* = 12 Hz, 1H, H_{5''}), 7.82–7.86 (d, 1H, H_{2''}), 7.93–7.94 (d, *J* = 4 Hz, 1H, H_{5'}), 8.15 (s, 1H, pyrimidine H), 8.65 (s, 1H, NH, exchangeable by D₂O), 8.67 (s, 1H, NH, exchangeable by D₂O), 10.56 (s, 1H, NH, exchangeable by D₂O); HRMS (ESI-MS) calcd: 376.17721 for C₂₁H₂₂N₅O₂ [M + H⁺]. Found: 376.17709.

5.1.6.5. *1-(4-(6,7-dihydro-5H-cyclopenta[d]pyrimidin-4-ylamino)phenyl)-3-(3-(trifluoromethyl)phenyl)urea (6e)*. The product was separated as grayish white powder (0.13 g, 22%), m.p 283–285 °C. ¹H NMR (400 MHz, DMSO-*d*₆) δ 1.14 (s, 4H, cyclopentenyl H), 1.24 (s, 2H, cyclopentenyl H), 6.50–6.52 (d, *J* = 8 Hz, 2H, H_{2'}, H_{6'}), 7.08–7.10 (d, *J* = 8 Hz, 2H, H_{4''}, H_{5''}), 7.22–7.24 (d, *J* = 8 Hz, 2H, H_{3'}, H_{5'}), 7.46–7.48 (d, *J* = 8 Hz, 1H, H_{6''}), 7.57 (s, 1H, H_{2''}), 7.99 (s, 1H, pyrimidine H), 9.01 (s, 1H, NH, exchangeable by D₂O), 8.84 (s, 1H, NH, exchangeable by D₂O), 10.56 (s, 1H, NH, exchangeable by D₂O). HRMS (ESI-MS) calcd: 414.15404 for C₂₁H₁₉N₅O₃ [M + H⁺]. Found: 294.08449 and 370.08269.

5.2. Biology

The structures of the synthesized compounds were submitted to BPS Bioscience, San Diego, CA, USA (www.bpsbioscience.com) for a VEGFR 2 tyrosine kinase assay. The assay was carried out using a Kinase-Glo Plus Luminescence Kinase Assay Kit (Promega). Kinase activity is measured by quantitating the amount of ATP remaining in the solution after a kinase reaction. The luminescent signal from the assay is correlated with the amount of ATP present and is inversely correlated with the amount of kinase activity. Compounds **6a**, **6b**, **6c**, **6d**, and **6e** were diluted in 10% DMSO, and 5 μ l of the dilution was added to a 50 μ l reaction so that the final concentration of DMSO was 1% in all reactions.

All of the enzymatic reactions were conducted at 30 °C for 45 min. The 50 μ l reaction mixture contained 40 mM Tris, pH 7.4, 10 mM MgCl₂, 0.1 mg/ml BSA, 1 mM DTT, 0.2 mg/ml Poly (Glu, Tyr) substrate, 10 μ M ATP and enzyme. After the enzymatic reaction, 50 μ l of Kinase-Glo Plus Luminescence Kinase Assay Solution (Promega) was added to each reaction, and the plate was incubated for 15 min at room temperature. The luminescence signal was measured using a BioTek Synergy 2 microplate reader.

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