



Immune cell diversity contributes to the pathogenesis of myocarditis

Xiumeng Hua¹ · Jiangping Song¹

Published online: 2 May 2019

© Springer Science+Business Media, LLC, part of Springer Nature 2019

Abstract

Myocarditis (MCD) is a type of inflammatory disease in which inflammatory cells infiltrate the myocardium, leading to cardiac dysfunction, myocardial necrosis, and fibrosis. Although it has been reported that MCD is mediated by T cells, the immune system is complex and includes many types of immune cells that interact with one another. Through investigations of the inflammatory responses in MCD including myocardial necrosis, fibrosis, and arrhythmia, we have gained further insight into the pathogenesis of MCD. This article aims to discuss the diversity and the roles of immune cells involved in the pathogenesis of MCD. Moreover, immunotherapy for the treatment of MCD remains controversial, and further investigation is required to identify accurate immunotherapies for special cell types.

Keywords Myocarditis · Immune cell diversity · Interaction · Pathogenesis · Immunotherapy

Introduction

Myocarditis (MCD) refers to the clinical and histological manifestations of a broad range of pathological immune processes in the heart, in which alterations in the number and function of lymphocyte subsets and macrophages and antibody-mediated injuries are present [1]. In most cases, MCD resolves spontaneously, but in susceptible individuals it may progress to a chronic stage, which leads to pathological cardiac remodeling that includes tissue fibrosis, hypertrophy, and apoptosis of cardiomyocytes. This remodeling results in a phenotype of dilated heart chambers with impaired cardiac function, in particular impaired contractility (inflammatory cardiomyopathy) [2]. Endocardial biopsy is the gold standard for diagnosing MCD, despite a lack of sensitivity, mainly due to sampling error [3, 4].

Many pathogenic factors contribute to the occurrence of MCD, including infectious factors (viruses, bacteria, and protozoa) and noninfectious factors (toxins, vaccines, and several drugs), as well as systemic autoimmune diseases that can also trigger heart-specific autoimmunity and inflammatory responses [5]. Inflammatory responses are mainly mediated by

immune cells and cytokines. In the resting or a diseased state, there are many types of immune cells that make up the whole cardiac cell library [6].

In the resting heart, the exact percentage of immune cells present is unknown, but presumed to be low, while a large number of immune cells infiltrate the space around cardiomyocytes in disease conditions, especially MCD [7, 8]. As has been previously demonstrated, a diverse set of immune cells contribute to inflammatory response processes, including innate and adaptive immune cells, so it is essential to determine the roles of specific immune cells in the pathogenesis of MCD. This review aims to summarize all the immune cells reported to be involved in MCD so far and to determine the related mechanisms in the process of MCD.

Immune anatomy in the resting heart

Innate and adaptive immune cells make up the cells of the immune system (Fig. 1). Like most tissues, the primary immune cells that reside in the heart are macrophages, which localize near endothelial cells or within the interstitial space, with very few monocytes found within cardiac tissue [9, 10]. Scarce numbers of dendritic cells (DCs) have been found within cardiac tissue and cardiac valves [11]. Mast cells, which are also found in resting cardiac tissue, are thought to be important early triggers of immune responses [12]. Neutrophils, which mostly take part in the pathophysiology of bacterial infection, are typically not found within noninfectious cardiac tissue [13–15]. As for adaptive immune cells, a

✉ Jiangping Song
fwsongjiangping@126.com

¹ State Key Laboratory of Cardiovascular Disease, Fuwai Hospital, National Center for Cardiovascular Diseases, Chinese Academy of Medical Sciences and Peking Union Medical College, 167A Beilishi Road, Xi Cheng District, Beijing 100037, China

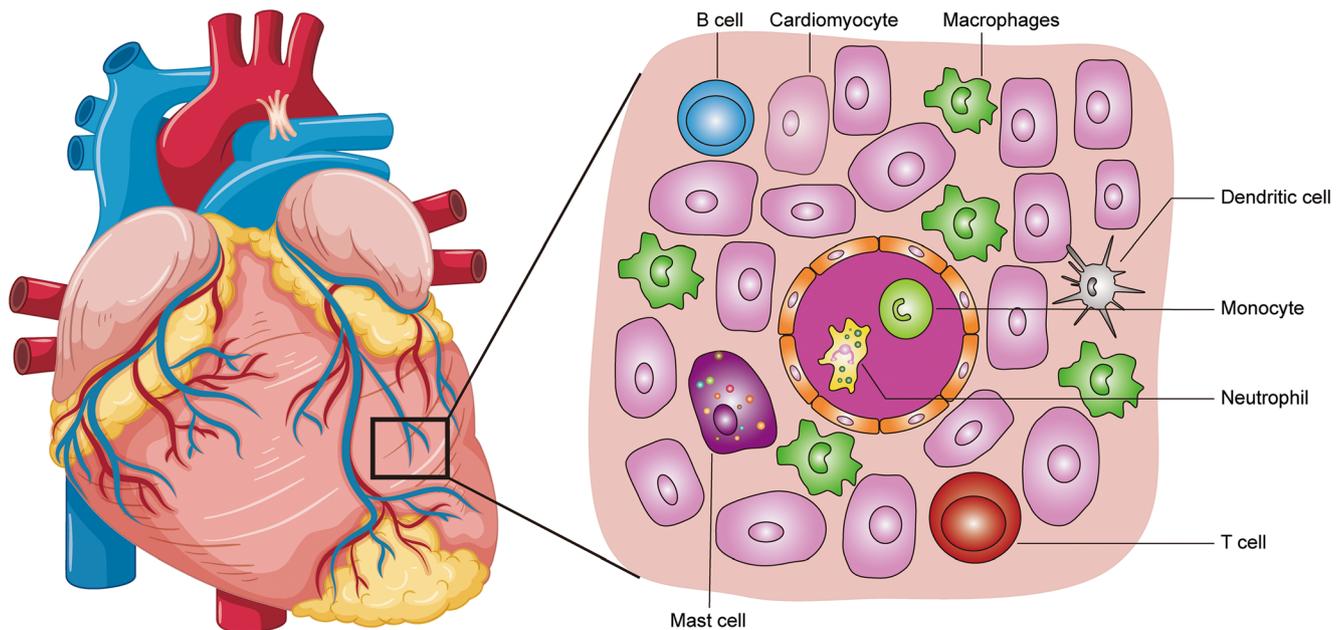


Fig. 1 Immune anatomy in the resting heart. Macrophages make up the majority of the immune cell population, which localize around the endothelial cells and myocardial interstitium. The mast cells and dendritic cells (DCs) can also be found in the resting heart. As has been

shown previously, neutrophils and monocytes are not found in the resting heart, except in the blood vessels. T cells and B cells were seldom found in normal condition

small number of regulatory T cell subsets and B cells are present in cardiac tissue during resting conditions [16, 17]. During resting conditions, innate immune cells make up a major proportion of all the immune cells present. Regardless of the inflammatory trigger, cardiac tissue initiates a dynamic cellular cascade that initially activates resident immune cells and over time evolves in a coordinated manner, leading to the recruitment of diverse leukocyte populations into the inflamed tissue, especially MCD (Table 1).

Macrophages protect against MCD by alleviating myocardial inflammation

Macrophages, which have characteristic CD68 and CD163 surface markers, make up a large portion of the cardiac immune cell population [18]. Recent studies using genetic fate mapping and adoptive transfer techniques have demonstrated that resident cardiac macrophage populations are defined by parabiosis. Rather than being a homogenous population, three discrete subsets with different origins and functions make up the resident cardiac macrophage population [19]. These three macrophage subsets can be well defined based on the expression of MHC class II and CC chemokine receptor 2 (CCR2). MHC class II^{hi} and MHC class II^{low} cardiac macrophages are both CCR2- and are numerically the dominant subsets in the heart [19]. These two subsets are primarily derived from embryonic progenitors, with a substantial number arising from

embryonic yolk sac precursors. They renew through in situ proliferation rather than through monocyte input. The third cardiac macrophage subset is made up of CCR2+ macrophages, which are derived from, and slowly replenished by, circulating blood monocytes. Previously, dichotomous expression of two classical myeloid cell markers (F4/80 and CD11b) was used to discern embryonic versus adult monocyte origin [43]. However, genetic fate mapping remains the most accurate method for lineage discrimination of cardiac macrophage because these cell surface markers alone failed to distinguish the different cardiac macrophage lineages [19, 44].

Interestingly, constructing a cardiac tissue injury model is the way to define the role of recruited monocytes and monocyte-derived macrophages. Nonselective depletion strategies have shown that in the absence of both monocytes and macrophages scar formation is impaired after cardiac ischemic injury, with decreased collagen production, decreased angiogenesis, and increased mortality due to myocardial rupture [10, 45]. These data suggest that recruited monocytes have a pathological role in the setting of sterile cardiac injury.

Macrophages play an important role in the pathogenesis of MCD. In CVB3-induced MCD when macrophages were depleted by liposome-encapsulated clodronate treatment, the mice presented higher viral titers but reduced acute myocarditis and chronic fibrosis [20]. In further studies, interleukin (IL)-33 upregulation significantly alleviated the severity of viral MCD with an increased cardiac contractive function and higher survival rate. Mechanistic

Table 1 Summary of immune characteristics in MCD

Type of immune cells	Markers	Functional roles	Association with MCD	Reference
Innate immune cells				
Macrophage (M)	CD68/CD163	Antigen presenting, innate immune response, electrical conduction facilitation	Macrophage M2 polarization protects mice from viral MCD by alleviating inflammation	[18]
MHC II ^{hi} M	MHC II ^{hi} /CCR2 ⁻			[19–21]
MHC II ^{low} M	MHC II ^{low} /CCR2 ⁻			
CCR2 ⁺ M	MHC II/CCR2 ⁺			
Dendritic cell (DCs)				[22]
Plasmacytoid DCs (pDCs)	CD123 CD303	Type 1 interferon production activating CTL	Limiting MCD	[23, 24]
Conventional or classical DCs (cDCs)	CD209, CD103 and CD141 MHC II	Antigen presenting		[24–26]
Natural killer cells (NK)	CD56 and CD94 (NKG2A)	Killing cells and regulating cardiac inflammation	Protect against MCD	[27–29]
Mast cells	Tryptases, chymases, and carboxypeptidase A3	The first line of defense and the release of granules to regulate the inflammatory response	The promotion of MCD and fibrosis need more investigation	[30, 31]
Adaptive immune cells				
CD4 ⁺ T cells	CD3, CD4			[2, 32]
Th1 cells	IL-12, IFN- γ , TNF- α	Regulating the inflammatory response	Limiting MCD	[33, 34]
Th2 cells	IL-4, IL-5, IL-10, IL-23		Promoting MCD	[33, 35]
Th17 cells	IL-17, IL-6, IL-21, IL-22		Promoting the MCD transition to DCM	[36, 37]
Tregs	CD4, CD25, FOXP3	Immunosuppressive activity against a broad and diverse array of antigens within different microenvironments by produce inhibitory cytokines	Limiting MCD	[32, 38]
CD8 ⁺ T cells	CD8	Cytotoxic T lymphocytes (CTLs), memory T cells and regulatory T cells	Promoting MCD, but this needs to be further investigated	[39–41]
B cells	CD19, CD20	Antibody production	Promoting MCD	[16, 42]

studies demonstrated that IL-33 could stimulate ST2L+ F4/80 + macrophages and ST2L+ CD4 + T cells in cardiac tissue to express IL-4, which was a potent inducer of macrophage M2 polarization by promoting the activation of JAK/STAT6, CEBP- β , PI3K, and other signaling pathways to increase the M2 gene expression of arginase-1, IL-10, macrophage mannose receptor (MMR), and macrophage galactose type C-type lectin (MGL). Additionally, M2 polarization resulted in the release of IL-10, which was an inducer of regulatory T cell-mediated immune responses (Fig. 2). Mice with adoptively transferred M2 macrophages exhibited less cardiac inflammation and attenuated myocarditis, suggesting a protective role for M2 macrophage in viral myocarditis [21]. Furthermore, male and female mice had different susceptibilities to MCD due to differences in M2 macrophage polarization and strikingly, the transfer of M2 macrophages into susceptible male mice remarkably alleviated myocardial inflammation by modulating local cytokine profiles and promoting peripheral regulatory T cell

differentiation [46]. Taken together, these data indicate that M2 macrophage polarization can protect mice from MCD by alleviating myocardial inflammation.

Macrophages play an essential role in not only the inflammatory response, but they also facilitate electrical conduction through the distal atrioventricular node where conducting cells are densely interspersed with elongated macrophages expressing connexin (CX)-43 [47]. Studies showed that when coupled to spontaneously beating cardiomyocytes via CX-43-containing gap junctions, cardiac macrophages have a negative resting membrane potential and depolarize in synchrony with cardiomyocytes. Conversely, macrophages render the resting membrane potential of cardiomyocytes more positively charged and accelerate their repolarization, according to computational modeling. Mechanistic studies suggested that the photostimulation of channelrhodopsin-2-expressing macrophages improves atrioventricular conduction, whereas conditional deletion of CX-43 in macrophages and congenital lack of macrophages delay atrioventricular conduction [47]. Meanwhile, some reports have indicated that a portion of

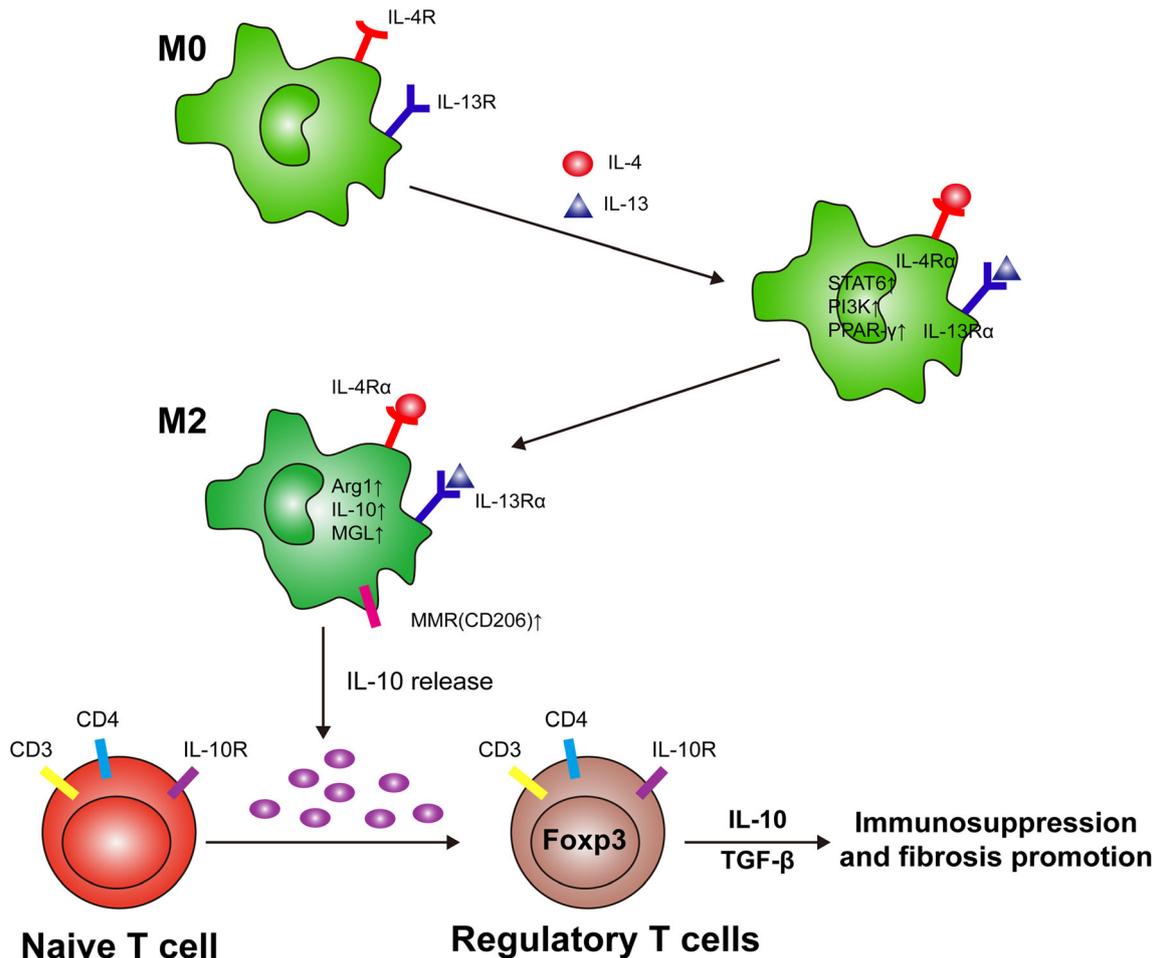


Fig. 2 M2 polarization function in MCD. IL-4/13 induces the transformation from M0 to M2 by activating the JAK/STAT6, PI3K, CEBP/β, and other pathways to increase the expression of M2 genes.

Furthermore, M2 cells release IL-10, which can help activate Treg, which in turn release IL-10 and TGF-β to induce inflammation and promote fibrosis

MCD patients have arrhythmias; for example, Uemura et al. reported that 3 of 50 (6%) patients with unexplained atrioventricular heart block had myocarditis [48]. However, whether macrophages contribute to atrioventricular heart block in MCD needs to be further investigated.

Dendritic cells induce autoimmune myocarditis by increasing Th1 polarization

Ralph Steinman, who discovered DCs and their role in initiating the adaptive immune response, was awarded as the 2011 Nobel Prize in Physiology or Medicine. DCs are specialized sentinel cells that act as antigen presenting cells (APCs) and bridge the innate and adaptive immune systems without directly engaging in effectors activities, such as pathogen killing [22]. Through pattern recognition receptors, such as toll-like receptors (TLRs), DCs recognize pathogens and migrate to the

T cell area of lymphoid organs to present pathogen-derived antigens to antigen-specific T cells.

DCs comprise two major classes: plasmacytoid DCs (pDCs) and conventional or classical DCs (cDCs). The pDCs are characterized by the surface markers CD123 and CD303 [23], while cDCs are characterized by the surface markers CD209, CD103, and CD141 [25, 26]. The pDCs rapidly produce type 1 interferon (IFN) following activation through nucleic acid-sensing TLRs, such as TLR7 and TLR9. cDCs are dedicated APCs that have a characteristic dendritic morphology and express high levels of MHC class II molecules. DCs play an important role in many inflammatory processes such as immune tolerance and autoimmunity. It has been reported that adoptive transfer of DCs loaded with cardiac antigen can induce the infiltration of CD4⁺ T cells into cardiac tissue and the development of experimental autoimmune myocarditis (EAM), which indicates that DCs are sufficient to induce MCD [24]. CD40 is essential in triggering self-

peptide-loaded DCs, and TLR simulation together with antigen-specific DCs has been shown to be required for disease induction [24]. Recently, Kania et al. found that IFN- γ , TLR, and nitric oxide (NO) signaling cooperated together to limit disease severity in EAM [49]. These data suggested that NO-mediated negative regulation of autoreactive T cells by tumor necrosis factor- α (TNF- α) and inducible NO synthase producing dendritic cells (TipDCs) occurs during autoimmune myocarditis. Together, factors-related infection or injury exposes the self-peptide and TLRs, which can then be taken up by and activate cDCs. Activated cDCs can induce Th1 polarization by releasing IL-12 to induce MCD (Fig. 3). Together, DCs play a role not only in antigen presentation, but also in the regulation of MCD. However, whether DCs are strictly necessary for the development of autoimmune myocarditis or just have a role in antigen mimicry-associated cardiac inflammation in MCD remains to be determined.

Natural killer cells can protect against myocarditis

Natural killer (NK) cells are a type of innate immune cell that characterized by the surface markers CD56 and CD94 (also called NKG2A) [27, 28]. NK cells are a versatile subpopulation of lymphocytes that make essential contributions to human innate and adaptive immune responses

to infection, especially viral infection [27]. In addition to their role as killer cells, NK cells can also regulate the cardiac inflammatory environment by inhibiting viral replication, autoreactive T cells, and ILC-derived Th2 cytokines, promoting eosinophil apoptosis and monocyte maturation, and inhibiting activated cardiac fibroblasts (Fig. 4).

In viral MCD, NK cells and macrophages are plentiful in the early infiltration of the heart after CVB3 infection [51]. They efficiently clear the virus by releasing perforin to kill infected cells and by stimulating adaptive immunity by producing IFN- γ [29]. Thus, NK cells were thought to suppress viral MCD by limiting viral replication by killing the virally infected cardiomyocytes; however, this process also results in the unavoidable release of autoantigens, which induce autoimmunity.

NK cells can also play an important role in regulating the inflammatory response by releasing cytokines to regulate other immune cells [52]. For example, NK cells can combine the cells surface receptors and cytokines to boost the maturation and activation of DCs, macrophages, and T cells. Conversely, NK cells can also kill immature DCs, activated CD4⁺T cells, and hyperactivated macrophages to decrease the inflammatory response. These regulatory functions of NK cells are kept in check by the recognition of constitutively expressed self-molecules by means of inhibitory receptors. It is still unknown whether there is any difference between NK cell killing of cardiomyocytes and NK cell regulation of other types of immune cells.

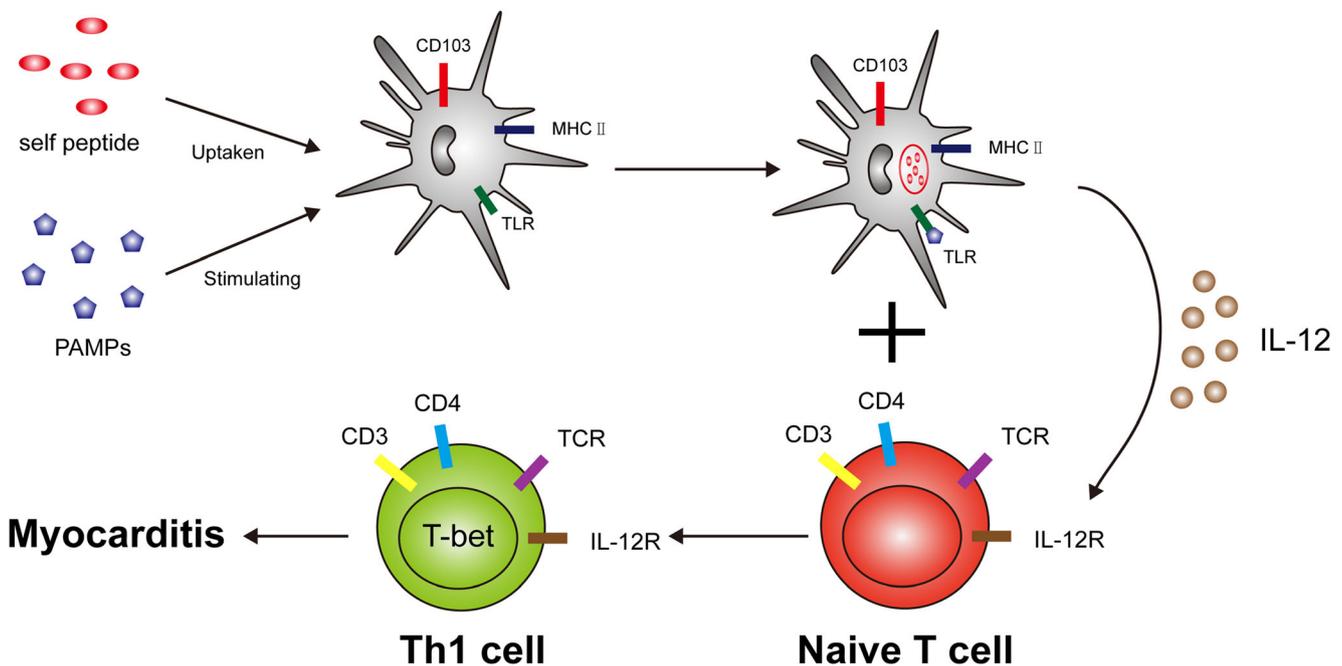


Fig. 3 cDCs can induce autoimmune myocarditis by inducing Th1 polarization. Self-peptide and PAMPs/DAMPs can induce the maturation of cDCs, which can combine with naïve T cells through

MHC II and TCR and can induce Th1 cells with the help of IL-12 released by cDCs. Th1 cells can induce myocarditis [50]

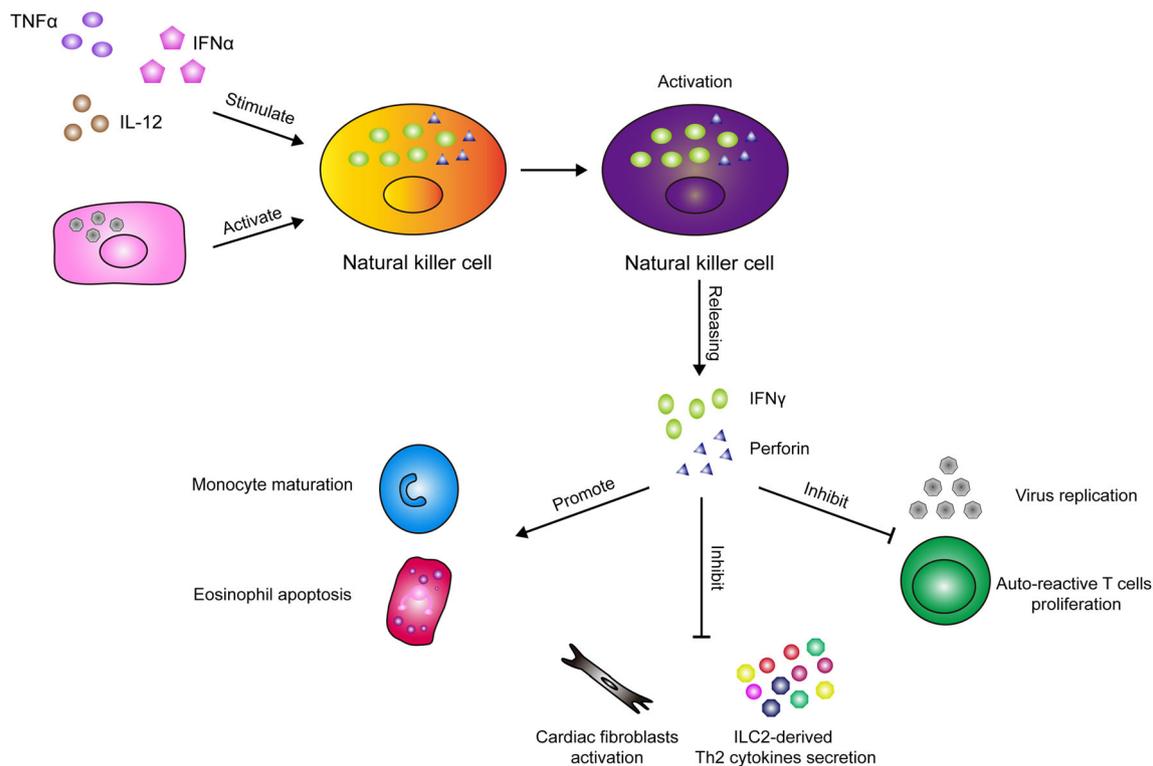


Fig. 4 Natural killer (NK) cells take part in MCD by releasing perforin and IFN- γ . NK cells can be activated by the TNF- α , IL-12, or IFN- α cytokines and infectious cells. Then, perforin and IFN- γ are released by activated NK cells to promote or inhibit several inflammatory responses

Mast cells play a critical role in the pathogenesis of myocarditis as inflammatory regulator

Mast cells are hematopoietic cells that arise from pluripotent precursors in the bone marrow. The distinguishing morphological feature of mast cells is their high content of electron-dense lysosome-like secretory granules (also known as secretory lysosomes) that occupy a major proportion of the cytoplasm of mature mast cells [30]. Mature mast cells are found in most tissues of the body and they are typically most abundant at sites close to host-environment interfaces, such as the skin and various other mucosal tissues. On account of the specific anatomical location, mast cells are thought as the first line of defense against external pathogens and attack by other environmental substances [53, 54]. Mast cells are characterized by proteases, including tryptases, chymases, and carboxypeptidase A3, which are different in humans and mice [30].

To investigate the role of mast cells in MCD (Fig. 5), Higuchi et al. used two strains of mast cell-deficient mice to verify that mast cells participate in the acute inflammatory reaction and the onset of ventricular remodeling associated with acute viral MCD [31]. These studies indicated that after constructing CVB3-induced MCD, the survival rate of mast cell-deficient mice was significantly higher than that of their

control littermates (77% versus 31%; $p = 0.03$; $n = 13$). Additionally, myocardial necrosis and cellular infiltration were significantly less pronounced in mast cell-deficient mice than in their control littermates. Furthermore, the expression levels of mast cell proteases were upregulated in the acute phase of viral MCD and rose further in the subacute phase of heart failure. The activation coincided with the development of myocardial necrosis and fibrosis and correlated with the upregulation of the expression of matrix metalloproteinase (MMP)-9 [31]. Thus, granule compounds released by mast cells can affect processes that are relevant to extracellular matrix (ECM) remodeling by activating ECM-remodeling enzymes.

Mast cells can also contribute to the immune regulation of Th2 polarization. In recent studies, these results showed that after adoptive transfer of immune cells, the mast cell-deficient mice showed stronger Th2 responses in the intestine than control mice. Mechanistic studies showed that mast cell-derived mouse mast cell protease-6 increased the expression of Bcl-6 in Th2 cells. Bcl-6 inhibited the expression of GATA-3 in Th2 cells, and subsequently, forkhead box P3 (Foxp3) was increased and Th2 cytokines were reduced. Thus, the cells showed immune regulatory properties similar to regulatory T cells [55]. Based on this research, we can speculate that mast cells may contribute to the pathogenesis of MCD in conditions in which MCD is characterized by a type 2 immune response

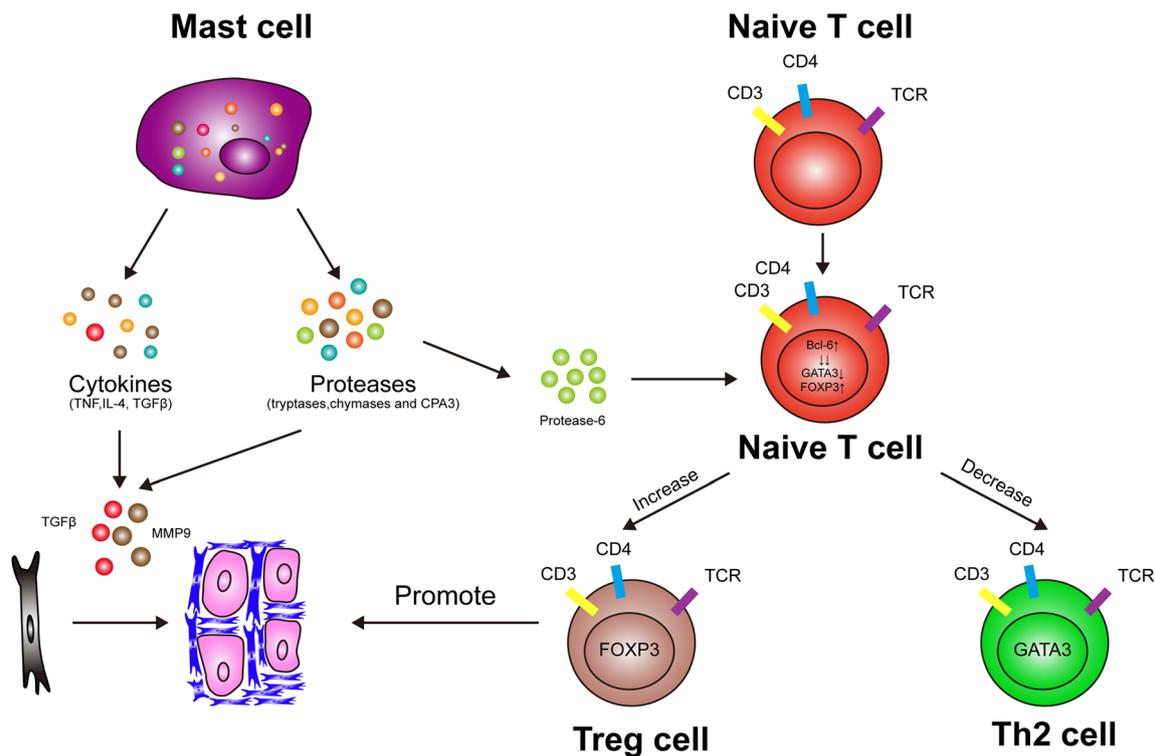


Fig. 5 Mast cells contribute to the process of EAM in fibrosis and reduce Th2 polarization. Mast cells are characterized by granule constituents including mast cell-specific proteases and cytokines. MMP9 and TGF-

β contribute to the progression of fibrosis. Protease-6 can change the transformation of naïve T cells to Treg cells than to Th2 cells. Treg cells further promote fibrosis development

[33, 56]. However, this hypothesis needs to be investigated further. For the role of mast cells in ECM remodeling leading to fibrosis, which is the most characteristic feature of the process from MCD to cardiomyopathy [57, 58], it is necessary to further investigate the role of mast cells in the chronic myopathy phase.

Role of CD4⁺ T cells in myocarditis as major immune cell effectors

As many studies have reported, autoreactive and most often infection-triggered CD4⁺ T cells are confirmed to be critical for MCD induction [2, 59]. According to the surface markers on T cells, there are two major subsets of T cells, CD4⁺T cells, and CD8⁺T cells (Fig. 6) [59]. Most of the T cells that are positive for the CD4⁺ surface marker are defined as the main drivers of heart-specific autoimmunity in MCD [60–62]. Expansion of heart-specific CD4⁺T cell effectors is facilitated in human and mice due to a high frequency of circulating naïve α -MyHC-specific CD4⁺T cells. CD4⁺T cells are also known as T helper (Th) cells and include Th1, Th2, Th17, and other T helper cells, aside from the CD4⁺ T cell subsets that include regulatory T cells (Tregs) [63]. Many studies, most of which are based on mouse models, indicate an

exclusive role for CD4⁺T cells in MCD development and progression. Unfortunately, the regulatory roles of the different CD4⁺T cell subtypes in MCD depend highly on the stage of disease and on a complex and not yet understood interaction between different inflammatory heart infiltrating and heart resident cell types.

With regard to Th cells, MCD is considered type 2 immunity so Th2 cells contribute to the pathogenesis of MCD [33, 35]. Song et al. found that besides tissue damage, a lower expression of VDR, a high frequency of Th2 cells, and an increase in Th2 cytokines were characteristics of MCD patient hearts during the end stages of heart failure. Spontaneous Th2-biased inflammation was observed in the hearts of VDR^{-/-} mice. CD4⁺ T cells from VDR^{-/-} mouse hearts were in a highly activated state. The naïve VDR^{-/-} CD4⁺ T cells and naïve CD4⁺ T cells from human hearts with MCD were prone to differentiate into Th2 cells. VDR formed complexes with GATA3, the IL-4 transcription factor, to suppress IL-4 gene transcription. Transplantation with VDR^{-/-} CD4⁺T cells induced Th2-biased inflammation in the hearts of Rag2^{-/-} mice. Reconstitution of VDR in CD4⁺ T cells inhibited Th2-biased inflammation in the heart.

MCD is a potentially lethal inflammatory heart disease that frequently leads to dilated cardiomyopathy (DCM); however, it is unclear which type of Th cells contributes to this

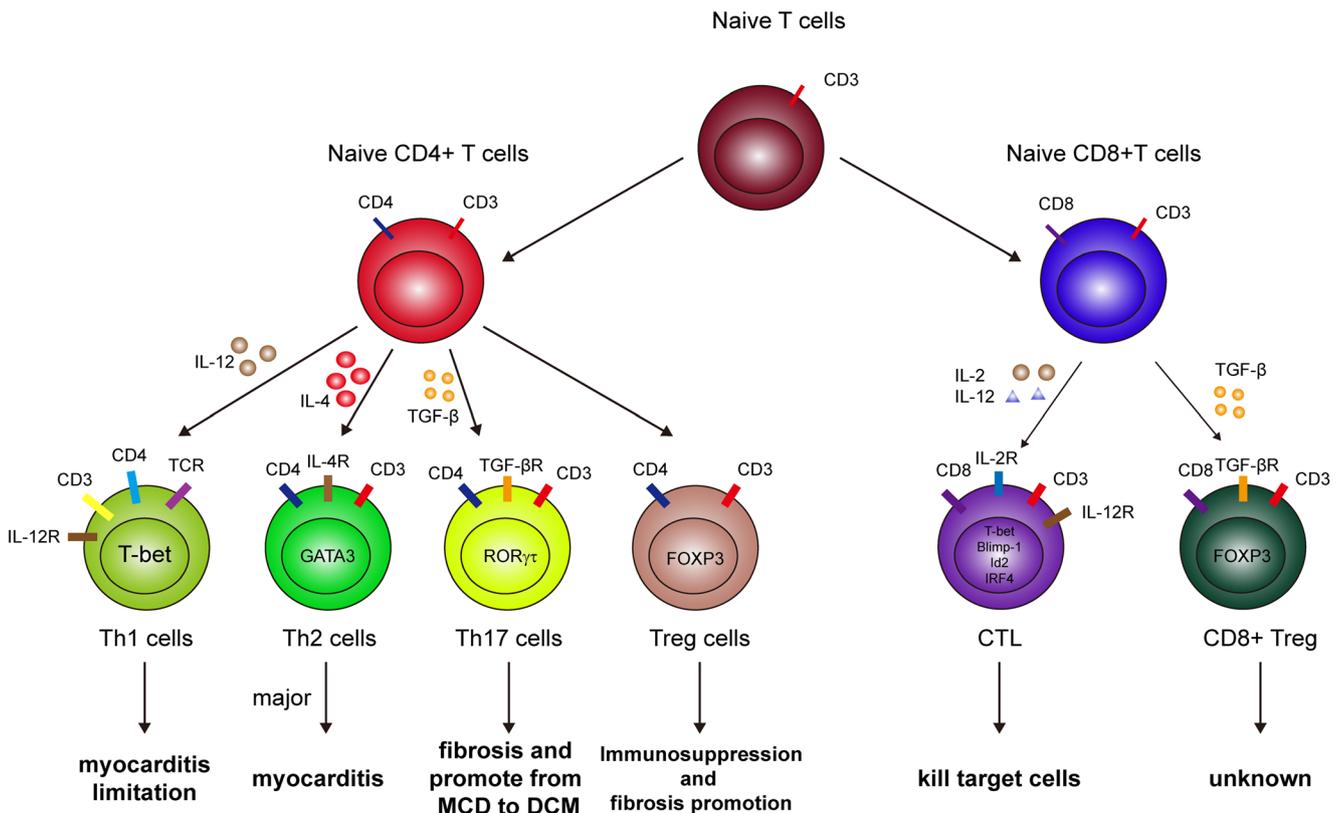


Fig. 6 Development of differentiated T cells and their roles in MCD. T cells are divided into two major types according to their surface markers (CD4 or CD8). Naive CD4⁺ T cells can be divided into four types of cells

during the accumulation of different cytokines from the innate cells, especially antigen presenting cells (APC). Naive CD8⁺ T cells can also be induced by cytokines from APC

progression. Veronika Nindl et al. found that cooperation of Th1 and Th17 cells determines the transition from autoimmune MCD to DCM [34], that IFN- γ was the major effector cytokine driving the initial inflammatory process, and that the cooperation of IFN- γ and IL-17A was essential for the development of progressive disease. Nevertheless, some contradictory findings have also been reported. For example, after immunizing IFN- γ - and IFN- γ R-deficient mice with α -MyHC/CFA, the mice developed more severe and persistent MCD, suggesting a protective regulatory role for IFN- γ in this disease model [36, 37]. However, which type of Th cells

contributes to the troublesome progression from MCD to DCM still needs to be investigated.

Treg cells are a specific subpopulation of T cells that comprise 5–10% of all peripheral CD4⁺T cells [32]. Treg cells are characterized by the surface markers CD4⁺, CD25⁺, and FOXP3⁺. FOXP3 is a specific marker of Treg cells that is essential for their maturation and function, so a loss of FOXP3 can result in disabled Treg cell function [2]. Treg cells can execute their immunosuppressive activity against a broad and diverse array of antigens within different microenvironments by producing inhibitory cytokines. Masahiro Ono et al.

Table 2 The methods of immunomodulation in myocarditis

Immunomodulatory drugs	Functional mechanism	Reference
High-dose intravenous immunoglobulin (IVIG)	Neutralization of autoantibodies; neutralization of chemokines and/or cytokines; Saturation and modulation of the expression of Fc γ receptors	[67, 68]
Peptide-ligands	Specify epitope-derived peptides as antibody-scavengers; direct targeting/suppression of aab-producing B cells and/or plasma-cells	[69, 70]
Immunoabsorption	IA of disease-causing aabs	[71–73]
Immunosuppression (azathioprine, steroids, cyclosporine A)	Widespread inhibitory effects on the immune system and action through a variety of signaling pathways; immunotoxicity	[74, 75]

found that GTR^{high} and FOXP3-expressing natural Tregs, made up of both CD25⁺ and CD25⁻ cell populations, contributed to the prevention of a variety of autoimmune/inflammatory diseases and that depletion of these cells allowed the activation of even weaker and rarer autoreactive T cells, causing widespread severe autoimmune disease [38]. However, diseases induced in this way include many which have been suspected of an autoimmune etiology in humans without much evidence.

Role of CD8⁺ T cells in myocarditis as cytotoxic lymphocytes

CD8⁺ T cells are referred to as cytotoxic T lymphocytes (CTLs), which were first reported in the 1960s to show specific cytotoxicity *in vitro* using mostly allogeneic combinations of cells [39]. Mechanistic studies suggest that CTLs most often kill target cells by inducing signaling through the same apoptotic pathways that are used for developmental cell death, including the perforin–granzyme pathway and the FAS pathway.

Additionally, various subgroups of CD8⁺ T lymphocytes demonstrate not only cytotoxic effects but also serve memory and regulatory roles in the body's immune response [40]. Massilamany et al. found that the CD8⁺ T cell response induced by Myhc338–348 was antigen-specific and transferred the disease to naïve animals, as evaluated by MHC class I/H-2Dd dextramer staining. Furthermore, the antigen-sensitized T cells predominantly produced interferon-gamma, the critical cytokine of effector cytotoxic T lymphocytes in EAM [41].

However, the role of regulatory CD8⁺ cells in MCD was unknown (Fig. 6).

B cells contribute to the autoimmunity process of myocarditis through autoantibody production

As mentioned intensively, autoimmunity contributes to the pathogenesis of MCD. In this condition, autoantibodies are key factors produced by B cells, which are characterized by CD19/CD20 [16]. Yoh Matsumoto et al. examined whether the cardiogenic epitopes that reside in the cardiac C-protein fragment2 (CC2) could be successful in inducing MCD and the development of DCM, compared with cardiac C-protein fragment 2 peptide 12 (CC2P12). They found the CC2-immunized rats exhibited marked B cell epitope spreading 4 weeks after immunization, whereas CC2P12-immunized rats raised antibodies against CC2P12 and CC2, but there was no difference in CC2- and CC2P12-immunized rat groups [42]. Then, they performed transfer experiments and demonstrated that both activation of T cells and anti-peptide antibody elevation were required for the initiation and subsequent progression of the disease. These findings strongly suggest that B cell epitope spreading is an essential step for the switch from MCD to DCM. The mechanism of how B cells identify the self-antigen to releasing autoantibody still needs further investigation to decrease the production of autoantibody.

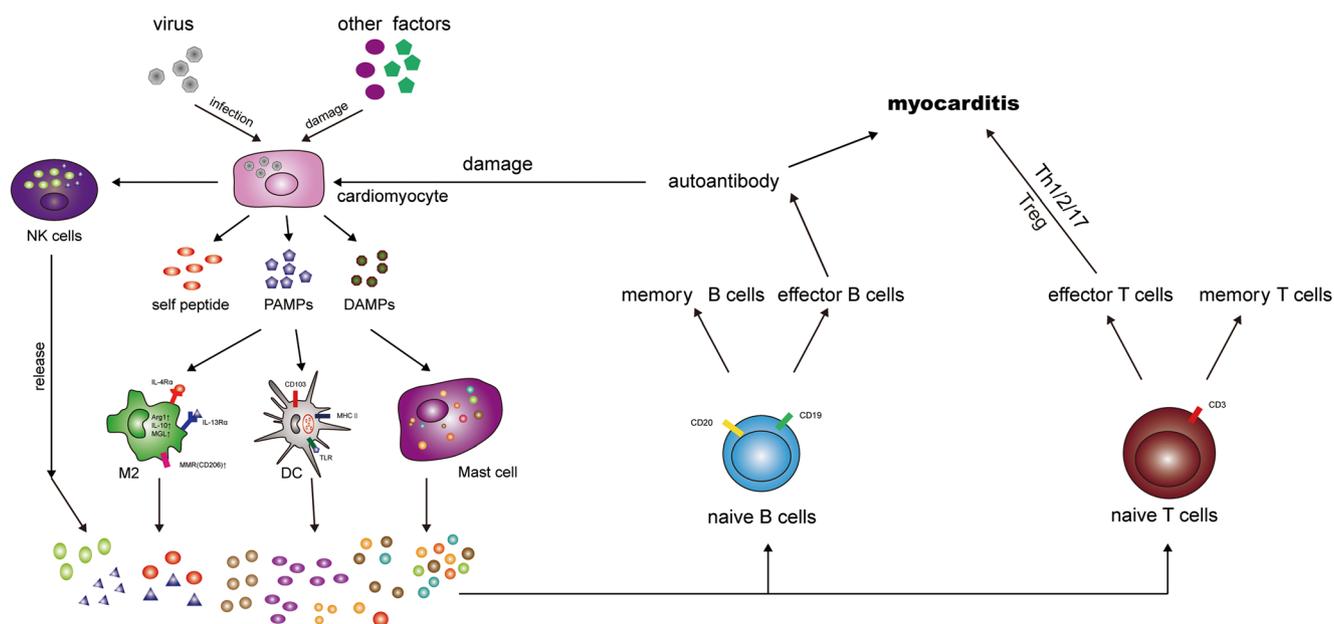


Fig. 7 The immune network in the pathogenesis of MCD

The prospects of immunotherapy in myocarditis

Immunotherapy has been advanced in cancer therapy and this has brought much hope for the patients who are suffering from cancer. In the field of cancer immunotherapy, adopting such methods regulates the immune system to inhibit the proliferation of tumors, such as enhancing T cell ability, because CD8⁺ T cells can clear up tumor cells [64]. Currently, almost all of the therapeutic strategies focus on promoting the ability of CTLs within a tumor, the type of CD8⁺ T cells, help in priming tumor-specific CTLs in the lymphoid organs and establishing efficient and durable antitumor immunity. With the advance of immunotherapy in tumors, immune checkpoint inhibition, such as the PD-1 antibody, can enhance the activity of antitumor T cells (majority CTLs) to clear up the tumor cells as therapy for cancer [65]. However, MCD is different from the cancer in the immune system, in that cancer may be the result of the exhaustion of T cells, while on the contrary, MCD may result from the abnormal activation of T cells to impair cardiomyocytes. Thus, the immunotherapy strategies used in cancer, which would have an opposite effect on MCD, are not appropriate for MCD. It has been reported the immune checkpoint inhibitors can induce MCD unexpectedly [66]. As there is abnormal activation of immune cells in MCD, some immunomodulation methods can be used for MCD (Table 2). For example, immunosuppressants have been applied to treat MCD with good outcomes in virus-negative cases without a known specificity of immune cells [76]. Additionally, it has been reported that TCR-based immunotherapy, which could be the future direction of MCD therapy, can be applicable to MCD, not only in animal but also in human MCD whose pathogenic mechanism is poorly understood [77]. As PD-1 can protect against inflammation and cardiomyocyte damage in T cell-mediated MCD [78], maybe increasing the expression of PD-1 induced by drugs or biosynthesis could help to treat MCD in the future.

Conclusions

Myocarditis is a type of inflammation of the heart involving innate and adaptive immune cells as shown in Table 1; it can lead to dilated cardiomyopathy. Different types of immune cells contribute to the development of MCD, and there is a complex network of inflammation (Fig. 7). Understanding the specific roles of these innate and adaptive cell populations at different stages of disease progression might provide the key to our insight into the pathogenesis of MCD as a base for developing potential therapeutic strategies. Immunotherapy has a huge potential role in the treatment of MCD, and further investigation is needed to accurately utilize immunotherapy on special cell types.

Funding This study was funded by CAMS Innovation Fund for Medical Sciences (CIFMS, 2016-I2M-1-015), PUMC Youth Fund (2016-XHQ03) and the National Natural Science Foundation of China (81670376).

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Sagar S, Liu PP, Cooper LJ (2012) Myocarditis. *Lancet* 379:738–747
- Vdovenko D, Eriksson U (2018) Regulatory role of CD4(+) T cells in myocarditis. *J Immunol Res* 2018:4396351
- Leone O, Veinot JP, Angelini A, Baandrup UT, Basso C, Berry G, Bruneval P, Burke M, Butany J, Calabrese F, D'Amati G, Edwards WD, Fallon JT, Fishbein MC, Gallagher PJ, Halushka MK, McManus B, Pucci A, Rodriguez ER, Saffitz JE, Sheppard MN, Steenbergen C, Stone JR, Tan C, Thiene G, van der Wal AC, Winters GL (2012) 2011 consensus statement on endomyocardial biopsy from the Association for European Cardiovascular Pathology and the Society for Cardiovascular Pathology. *Cardiovasc Pathol* 21:245–274
- Heymans S, Eriksson U, Lehtonen J, Cooper LJ (2016) The quest for new approaches in myocarditis and inflammatory cardiomyopathy. *J Am Coll Cardiol* 68:2348–2364
- Mahrholdt H, Wagner A, Deluigi CC, Kispert E, Hager S, Meinhardt G, Vogelsberg H, Fritz P, Dippon J, Bock CT, Klingel K, Kandolf R, Sechtem U (2006) Presentation, patterns of myocardial damage, and clinical course of viral myocarditis. *Circulation* 114:1581–1590
- Epelman S, Liu PP, Mann DL (2015) Role of innate and adaptive immune mechanisms in cardiac injury and repair. *Nat Rev Immunol* 15:117–129
- Hamid T, Prabhu SD (2017) Immunomodulation is the key to cardiac repair. *Circ Res* 120:1530–1532
- Fung G, Luo H, Qiu Y, Yang D, McManus B (2016) Myocarditis. *Circ Res* 118:496–514
- Pinto AR, Paolicelli R, Salimova E, Gospocic J, Slonimsky E, Bilbao-Cortes D, Godwin JW, Rosenthal NA (2012) An abundant tissue macrophage population in the adult murine heart with a distinct alternatively-activated macrophage profile. *PLoS One* 7: e36814
- Nahrendorf M, Swirski FK, Aikawa E, Stangenberg L, Wurdinger T, Figueiredo JL, Libby P, Weissleder R, Pittet MJ (2007) The healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary functions. *J Exp Med* 204: 3037–3047
- Choi JH, Do Y, Cheong C, Koh H, Boscardin SB, Oh YS, Bozzacco L, Trumpfeller C, Park CG, Steinman RM (2009) Identification of antigen-presenting dendritic cells in mouse aorta and cardiac valves. *J Exp Med* 206:497–505
- Frangogiannis NG, Lindsey ML, Michael LH, Youker KA, Bressler RB, Mendoza LH, Spengler RN, Smith CW, Entman ML (1998) Resident cardiac mast cells degranulate and release preformed TNF-alpha, initiating the cytokine cascade in experimental canine myocardial ischemia/reperfusion. *Circulation* 98:699–710
- Kolaczowska E, Kubes P (2013) Neutrophil recruitment and function in health and inflammation. *Nat Rev Immunol* 13:159–175

14. Amulic B, Cazalet C, Hayes GL, Metzler KD, Zychlinsky A (2012) Neutrophil function: from mechanisms to disease. *Annu Rev Immunol* 30:459–489
15. Shah AD, Denaxas S, Nicholas O, Hingorani AD, Hemingway H (2017) Neutrophil counts and initial presentation of 12 cardiovascular diseases: a CALIBER cohort study. *J Am Coll Cardiol* 69:1160–1169
16. Zouggari Y, Ait-Oufella H, Bonnin P, Simon T, Sage AP, Guerin C, Vilar J, Caligiuri G, Tsiantoulas D, Laurans L, Dumeau E, Kotti S, Bruneval P, Charo IF, Binder CJ, Danchin N, Tedgui A, Tedder TF, Silvestre JS, Mallat Z (2013) B lymphocytes trigger monocyte mobilization and impair heart function after acute myocardial infarction. *Nat Med* 19:1273–1280
17. Saxena A, Dobaczewski M, Rai V, Haque Z, Chen W, Li N, Frangogiannis NG (2014) Regulatory T cells are recruited in the infarcted mouse myocardium and may modulate fibroblast phenotype and function. *Am J Physiol Heart Circ Physiol* 307:H1233–H1242
18. Tan KL, Scott DW, Hong F, Kahl BS, Fisher RI, Bartlett NL, Advani RH, Buckstein R, Rimsza LM, Connors JM, Steidl C, Gordon LL, Horning SJ, Gascoyne RD (2012) Tumor-associated macrophages predict inferior outcomes in classic Hodgkin lymphoma: a correlative study from the E2496 intergroup trial. *Blood* 120:3280–3287
19. Epelman S, Lavine KJ, Beaudin AE, Sojka DK, Carrero JA, Calderon B, Brija T, Gautier EL, Ivanov S, Satpathy AT, Schilling JD, Schwendener R, Sergin I, Razani B, Forsberg EC, Yokoyama WM, Unanue ER, Colonna M, Randolph GJ, Mann DL (2014) Embryonic and adult-derived resident cardiac macrophages are maintained through distinct mechanisms at steady state and during inflammation. *Immunity* 40:91–104
20. Jaquenod DGC, Ure AE, Rivadeneyra L, Schattner M, Gomez RM (2015) Macrophages and galectin 3 play critical roles in CVB3-induced murine acute myocarditis and chronic fibrosis. *J Mol Cell Cardiol* 85:58–70
21. Wang C, Dong C, Xiong S (2017) IL-33 enhances macrophage M2 polarization and protects mice from CVB3-induced viral myocarditis. *J Mol Cell Cardiol* 103:22–30
22. Ganguly D, Haak S, Sisirak V, Reizis B (2013) The role of dendritic cells in autoimmunity. *Nat Rev Immunol* 13:566–577
23. Sapienza MR, Fuligni F, Agostinelli C, Tripodo C, Righi S, Laginestra MA, Pileri AJ, Mancini M, Rossi M, Ricci F, Gazzola A, Melle F, Mannu C, Ulbar F, Arpinati M, Paulli M, Maeda T, Gibellini D, Pagano L, Pimpinelli N, Santucci M, Cerroni L, Croce CM, Facchetti F, Piccaluga PP, Pileri SA (2014) Molecular profiling of blastic plasmacytoid dendritic cell neoplasm reveals a unique pattern and suggests selective sensitivity to NF- κ B pathway inhibition. *Leukemia* 28:1606–1616
24. Eriksson U, Ricci R, Hunziker L, Kurrer MO, Oudit GY, Watts TH, Sonderegger I, Bachmaier K, Kopf M, Penninger JM (2003) Dendritic cell-induced autoimmune heart failure requires cooperation between adaptive and innate immunity. *Nat Med* 9:1484–1490
25. Kretzschmar D, Betge S, Windisch A, Pistulli R, Rohm I, Fritzenwanger M, Jung C, Schubert K, Theis B, Petersen I, Drobnik S, Mall G, Figulla HR, Yilmaz A (2012) Recruitment of circulating dendritic cell precursors into the infarcted myocardium and pro-inflammatory response in acute myocardial infarction. *Clin Sci (Lond)* 123:387–398
26. Collin M, McGovern N, Haniffa M (2013) Human dendritic cell subsets. *Immunology* 140:22–30
27. Parham P, Guethlein LA (2018) Genetics of natural killer cells in human health, disease, and survival. *Annu Rev Immunol* 36:519–548
28. Mace EM, Gunesch JT, Dixon A, Orange JS (2016) Human NK cell development requires CD56-mediated motility and formation of the developmental synapse. *Nat Commun* 7:12171
29. Jost S, Altfeld M (2013) Control of human viral infections by natural killer cells. *Annu Rev Immunol* 31:163–194
30. Wernersson S, Pejler G (2014) Mast cell secretory granules: armed for battle. *Nat Rev Immunol* 14:478–494
31. Higuchi H, Hara M, Yamamoto K, Miyamoto T, Kinoshita M, Yamada T, Uchiyama K, Matsumori A (2008) Mast cells play a critical role in the pathogenesis of viral myocarditis. *Circulation* 118:363–372
32. Meng X, Yang J, Dong M, Zhang K, Tu E, Gao Q, Chen W, Zhang C, Zhang Y (2016) Regulatory T cells in cardiovascular diseases. *Nat Rev Cardiol* 13:167–179
33. Song J, Chen X, Cheng L, Rao M, Chen K, Zhang N, Meng J, Li M, Liu ZQ, Yang PC (2018) Vitamin D receptor restricts Th2-biased inflammation in the heart. *Cardiovasc Res* 114:870–879
34. Nindl V, Maier R, Ratering D, De Giuli R, Zust R, Thiel V, Scandella E, Di Padova F, Kopf M, Rudin M, Rulicke T, Ludewig B (2012) Cooperation of Th1 and Th17 cells determines transition from autoimmune myocarditis to dilated cardiomyopathy. *Eur J Immunol* 42:2311–2321
35. Walker JA, McKenzie A (2018) TH2 cell development and function. *Nat Rev Immunol* 18:121–133
36. Eriksson U, Kurrer MO, Sebald W, Brombacher F, Kopf M (2001) Dual role of the IL-12/IFN- γ axis in the development of autoimmune myocarditis: induction by IL-12 and protection by IFN- γ . *J Immunol* 167:5464–5469
37. Eriksson U, Kurrer MO, Bingisser R, Eugster HP, Saremaslani P, Follath F, Marsch S, Widmer U (2001) Lethal autoimmune myocarditis in interferon- γ receptor-deficient mice: enhanced disease severity by impaired inducible nitric oxide synthase induction. *Circulation* 103:18–21
38. Ono M, Shimizu J, Miyachi Y, Sakaguchi S (2006) Control of autoimmune myocarditis and multiorgan inflammation by glucocorticoid-induced TNF receptor family-related protein (high), Foxp3-expressing CD25+ and CD25- regulatory T cells. *J Immunol* 176:4748–4756
39. Golstein P, Griffiths GM (2018) An early history of T cell-mediated cytotoxicity. *Nat Rev Immunol* 18:527–535
40. Yu Y, Ma X, Gong R, Zhu J, Wei L, Yao J (2018) Recent advances in CD8(+) regulatory T cell research. *Oncol Lett* 15:8187–8194
41. Massilamany C, Gangapla A, Basavalingappa RH, Rajasekaran RA, Khalilzad-Sharghi V, Han Z, Othman S, Steffen D, Reddy J (2016) Localization of CD8 T cell epitope within cardiac myosin heavy chain- α 334-352 that induces autoimmune myocarditis in a/J mice. *Int J Cardiol* 202:311–321
42. Matsumoto Y, Park IK, Kohyama K (2007) B-cell epitope spreading is a critical step for the switch from C-protein-induced myocarditis to dilated cardiomyopathy. *Am J Pathol* 170:43–51
43. Schulz C, Gomez PE, Choro L, Szabo-Rogers H, Cagnard N, Kierdorf K, Prinz M, Wu B, Jacobsen SE, Pollard JW, Frampton J, Liu KJ, Geissmann F (2012) A lineage of myeloid cells independent of Myb and hematopoietic stem cells. *Science* 336:86–90
44. Molawi K, Wolf Y, Kandalla PK, Favret J, Hagemeyer N, Frenzel K, Pinto AR, Klapproth K, Henri S, Malissen B, Rodewald HR, Rosenthal NA, Bajenoff M, Prinz M, Jung S, Sieweke MH (2014) Progressive replacement of embryo-derived cardiac macrophages with age. *J Exp Med* 211:2151–2158
45. van Amerongen MJ, Harmsen MC, van Rooijen N, Petersen AH, van Luyn MJ (2007) Macrophage depletion impairs wound healing and increases left ventricular remodeling after myocardial injury in mice. *Am J Pathol* 170:818–829
46. Li K, Xu W, Guo Q, Jiang Z, Wang P, Yue Y, Xiong S (2009) Differential macrophage polarization in male and female BALB/c mice infected with coxsackievirus B3 defines susceptibility to viral myocarditis. *Circ Res* 105:353–364
47. Hulsmans M, Clauss S, Xiao L, Aguirre AD, King KR, Hanley A, Hucker WJ, Wulfers EM, Seemann G, Courties G, Iwamoto Y, Sun

- Y, Savol AJ, Sager HB, Lavine KJ, Fishbein GA, Capen DE, Da SN, Miquero L, Wakimoto H, Seidman CE, Seidman JG, Sadreyev RI, Naxerova K, Mitchell RN, Brown D, Libby P, Weissleder R, Swirski FK, Kohl P, Vinegoni C, Milan DJ, Ellinor PT, Nahrendorf M (2017) Macrophages facilitate electrical conduction in the heart. *Cell* 169:510–522.e20
48. Uemura A, Morimoto S, Hiramitsu S, Hishida H (2001) Endomyocardial biopsy findings in 50 patients with idiopathic atrioventricular block: presence of myocarditis. *Jpn Heart J* 42: 691–700
 49. Kania G, Siegert S, Behnke S, Prados-Rosales R, Casadevall A, Luscher TF, Luther SA, Kopf M, Eriksson U, Blyszczuk P (2013) Innate signaling promotes formation of regulatory nitric oxide-producing dendritic cells limiting T-cell expansion in experimental autoimmune myocarditis. *Circulation* 127:2285–2294
 50. Griffin GK, Lichtman AH (2013) Two sides to every proinflammatory coin: new insights into the role of dendritic cells in the regulation of T-cell driven autoimmune myocarditis. *Circulation* 127: 2257–2260
 51. Fairweather D, Kaya Z, Shellam GR, Lawson CM, Rose NR (2001) From infection to autoimmunity. *J Autoimmun* 16(3):175–186
 52. Vivier E, Tomasello E, Baratin M, Walzer T, Ugolini S (2008) Functions of natural killer cells. *Nat Immunol* 9:503–510
 53. Marshall JS (2004) Mast-cell responses to pathogens. *Nat Rev Immunol* 4:787–799
 54. St JA, Abraham SN (2013) Innate immunity and its regulation by mast cells. *J Immunol* 190:4458–4463
 55. Liu ZQ, Song JP, Liu X, Jiang J, Chen X, Yang L, Hu T, Zheng PY, Liu ZG, Yang PC (2014) Mast cell-derived serine proteinase regulates T helper 2 polarization. *Sci Rep* 4:4649
 56. Gieseck RR, Wilson MS, Wynn TA (2017) Type 2 immunity in tissue repair and fibrosis. *Nat Rev Immunol* 18:62–76
 57. Segura AM, Frazier OH, Buja LM (2014) Fibrosis and heart failure. *Heart Fail Rev* 19:173–185
 58. Blyszczuk P, Muller-Edenborn B, Valenta T, Osto E, Stellato M, Behnke S, Glatz K, Basler K, Luscher TF, Distler O, Eriksson U, Kania G (2017) Transforming growth factor-beta-dependent Wnt secretion controls myofibroblast formation and myocardial fibrosis progression in experimental autoimmune myocarditis. *Eur Heart J* 38:1413–1425
 59. Opavsky MA, Penninger J, Aitken K, Wen WH, Dawood F, Mak T, Liu P (1999) Susceptibility to myocarditis is dependent on the response of alphabeta T lymphocytes to coxsackieviral infection. *Circ Res* 85:551–558
 60. Smith SC, Allen PM (1991) Myosin-induced acute myocarditis is a T cell-mediated disease. *J Immunol* 147:2141–2147
 61. Penninger JM, Pummerer C, Liu P, Neu N, Bachmaier K (1997) Cellular and molecular mechanisms of murine autoimmune myocarditis. *APMIS* 105:1–13
 62. Chen P, Baldeviano GC, Ligons DL, Talor MV, Barin JG, Rose NR, Cihakova D (2012) Susceptibility to autoimmune myocarditis is associated with intrinsic differences in CD4(+) T cells. *Clin Exp Immunol* 169:79–88
 63. Bonelli M, Shih HY, Hirahara K, Singelton K, Laurence A, Poholek A, Hand T, Mikami Y, Vahedi G, Kanno Y, O'Shea JJ (2014) Helper T cell plasticity: impact of extrinsic and intrinsic signals on transcriptomes and epigenomes. *Curr Top Microbiol Immunol* 381:279–326
 64. Borst J, Ahrends T, Babala N, Melief C, Kastenmuller W (2018) CD4(+) T cell help in cancer immunology and immunotherapy. *Nat Rev Immunol* 18:635–647
 65. Ribas A, Wolchok JD (2018) Cancer immunotherapy using checkpoint blockade. *Science* 359:1350–1355
 66. Johnson DB, Balko JM, Compton ML, Chalkias S, Gorham J, Xu Y, Hicks M, Puzanov I, Alexander MR, Bloomer TL, Becker JR, Slosky DA, Phillips EJ, Pilkinton MA, Craig-Owens L, Kola N, Plautz G, Reshef DS, Deutsch JS, Deering RP, Olenchock BA, Lichtman AH, Roden DM, Seidman CE, Koralknik IJ, Seidman JG, Hoffman RD, Taube JM, Diaz LJ, Anders RA, Sosman JA, Moslehi JJ (2016) Fulminant myocarditis with combination immune checkpoint blockade. *N Engl J Med* 375:1749–1755
 67. Chaigne B, Mouthon L (2017) Mechanisms of action of intravenous immunoglobulin. *Transfus Apher Sci* 56(1):45–49
 68. Orange J, Hossny E, Weiler C, Ballow M, Berger M, Bonilla F, Buckley R, Chinen J, Elgamal Y, Mazer B (2006) Use of intravenous immunoglobulin in human disease: a review of evidence by members of the primary immunodeficiency Committee of the American Academy of allergy, asthma and immunology. *J Allergy Clin Immunol* 117(4):S525–S553
 69. Neubert K, Meister S, Moser K, Weisel F, Maseda D, Amann K, Wiethe C, Winkler TH, Kalden JR, Manz RA, Voll RE (2008) The proteasome inhibitor bortezomib depletes plasma cells and protects mice with lupus-like disease from nephritis. *Nat Med* 14(7):748–755
 70. Nikolaev VO, Boivin V, Störk S, Angermann CE, Ertl G, Lohse MJ, Jahns R (2007) A novel fluorescence method for the rapid detection of functional β_1 -adrenergic receptor autoantibodies in heart failure. *J Am Coll Cardiol* 50(5):423–431
 71. Mobini R, Staudt A, Felix SB, Baumann G, Wallukat G, Deinum J, Svensson H, Hjalmarson Å, Michael F (2003) Hemodynamic improvement and removal of autoantibodies against β -adrenergic receptor by immunoabsorption therapy in dilated cardiomyopathy. *J Autoimmun* 20(4):345–350
 72. Trimper C, Herda LR, Eckerle LG, Pohle S, Müller C, Landsberger M, Felix SB, Staudt A (2010) Immunoabsorption in dilated cardiomyopathy: long-term reduction of cardiodepressant antibodies. *Eur J Clin Investig* 40(8):685–691
 73. Felix SB, Staudt A, Landsberger M, Grosse Y, Stangl V, Spielhagen T, Wallukat G, Wernecke KD, Baumann G, Stangl K (2002) Removal of cardiodepressant antibodies in dilated cardiomyopathy by immunoabsorption. *J Am Coll Cardiol* 39(4):646–652
 74. Benvenuto LJ, Anderson MR, Arcasoy SM (2018) New frontiers in immunosuppression. *J Thorac Dis* 10(5):3141–3155
 75. Schmeits PCJ, Schaap MM, Luijten M, van Someren E, Boorsma A, van Loveren H, Peijnenburg AACM, Hendriksen PJM (2015) Detection of the mechanism of immunotoxicity of cyclosporine a in murine in vitro and in vivo models. *Arch Toxicol* 89(12):2325–2337
 76. Winter MP, Sulzgruber P, Koller L, Bartko P, Goliasch G, Niessner A (2018) Immunomodulatory treatment for lymphocytic myocarditis—a systematic review and meta-analysis. *Heart Fail Rev* 23:573–581
 77. Matsumoto Y, Jee Y, Sugisaki M (2000) Successful TCR-based immunotherapy for autoimmune myocarditis with DNA vaccines after rapid identification of pathogenic TCR. *J Immunol* 164:2248–2254
 78. Tarrío ML, Grabie N, Bu DX, Sharpe AH, Lichtman AH (2012) PD-1 protects against inflammation and myocyte damage in T cell-mediated myocarditis. *J Immunol* 188:4876–4884

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.