



Outcome and molecular characteristics of non-invasive encapsulated follicular variant of papillary thyroid carcinoma with oncocytic features

Bin Xu¹ · Ed Reznik² · R. Michael Tuttle³ · Jeffrey Knauf³ · James A. Fagin³ · Nora Katabi⁴ · Snjezana Dogan⁴ · Nathaniel Aleynick⁴ · Venkatraman Seshan² · Sumit Middha⁴ · Danny Enepekides⁵ · Gian Piero Casadei⁶ · Erica Solaroli⁷ · Giovanni Tallini⁸ · Ronald Ghossein⁴ · Ian Ganly⁹

Received: 10 December 2018 / Accepted: 17 January 2019 / Published online: 28 January 2019
© Springer Science+Business Media, LLC, part of Springer Nature 2019

Abstract

Purpose In 2016, non-invasive encapsulated follicular variant of papillary thyroid carcinoma (NI-EFVPTC) was renamed as noninvasive thyroid follicular neoplasm with papillary-like nuclear features (NIFTP). However, as the study cohort did not mention tumors with oncocytic features, such lesions are still labeled by some as FVPTC. It is therefore crucial to evaluate the outcome and molecular profile of oncocytic NI-EFVPTC.

Methods A multi-institutional clinico-pathologic review was conducted to select 61 patients having oncocytic NI-EFVPTC. A detailed molecular profile was carried out in 15 patients.

Results Oncocytic NI-EFVPTCs predominantly affected women in their 50s. There was no distant metastasis, lymph node metastases, or structural recurrence in the entire cohort. Among patients with ≥5 years of FU, all 33 individuals did not recur with a median FU of 10.2 years. Oncocytic NI-EFVPTC commonly had *RAS* (33%) mutations, a high frequency of mitochondrial DNA mutations (67%) and multiple chromosomal gains/losses (53%). No fusion genes were detected.

Conclusions Oncocytic NI-EFVPTC, when stringently selected for, lacks metastasis at presentation and follows an extremely indolent clinical course, even when treated conservatively with lobectomy alone without RAI therapy. These tumors share a similar mutational profile as NIFTP, FVPTC, and follicular neoplasm and are predominantly *RAS*-related. Like Hurthle cell neoplasms, they harbor a high frequency of mitochondrial DNA mutations, which contribute to the oncocytic cytomorphology. However, they lack the widespread chromosomal alterations observed in Hurthle cell carcinoma. Consideration should be given to include oncocytic NI-EFVPTCs as NIFTP in order to avoid overtreatment of these highly indolent tumors.

Keywords Encapsulated follicular variant · Papillary thyroid carcinoma · Noninvasive follicular thyroid neoplasm with papillary-like nuclear features (NIFTP) · *RAS* · Oncocytic

These authors contributed equally: Bin Xu, Ed Reznik

These authors jointly supervised this work: Ronald Ghossein, Ian Ganly

Supplementary information The online version of this article (<https://doi.org/10.1007/s12020-019-01848-6>) contains supplementary material, which is available to authorized users.

✉ Ronald Ghossein
ghosseir@mskcc.org

✉ Ian Ganly
ganlyi@mskcc.org

Extended author information available on the last page of the article

Introduction

The incidence of thyroid carcinoma has increased more than any other cancer in the United States, with an annual increase of 3.6% per year and 56,430 new cases diagnosed annually [1, 2]. The increase is in part attributed to a rise in the prevalence of the follicular variant of papillary thyroid carcinoma (FVPTC), a diagnosis that is rendered with a certain degree of inter-observer subjectivity [3]. For example, the percentage of FVPTC among all papillary thyroid carcinomas (PTC) regardless of tumor size has nearly tripled from 18 to 57% over the past four decades, becoming the most common architectural patterns encountered in PTC [3].

Histologically, PTC can be subtyped using architectural patterns or cytologic features. FVPTC is a variant of PTC that is characterized by an exclusively follicular growth pattern, while the oncocytic variant is used to describe PTC with prominent oncocytic cytomorphology characterized by abundant eosinophilic granular cytoplasm [4, 5]. Recently, compelling clinical outcome data and molecular evidence, including The Cancer Genomic Atlas (TCGA) of PTC, have demonstrated that noninvasive FVPTC follows a highly indolent clinical course and has a molecular signature resembling follicular adenoma/follicular carcinoma with activating *RAS* mutations as the most frequently encountered genetic alteration [5–9]. In 2016, a group of 28 endocrine experts critically reexamined this entity and advocated for a nomenclature revision to non-invasive follicular thyroid neoplasm with papillary-like nuclear features (NIFTP), in an effort to reduce overtreatment of this highly indolent tumor by eliminating the term “carcinoma” [5]. Such nomenclature revision was subsequently recognized and adopted by the World Health Organization (WHO) [4].

As the cohort of 109 patients with NIFTP studied by the consensus conference [5] and in a previous report [8] did not explicitly address tumors with oncocytic features, i.e., oncocytic noninvasive encapsulated follicular variant of papillary thyroid carcinoma (O-NI-EFVPTC), there is currently no study with long-term follow up specifically designated to investigate the clinical behavior and outcome of O-NI-EFVPTC. As a consequence, such lesions are labeled and staged by many pathologists as O-NI-EFVPTC rather than NIFTP. Indeed, the United Kingdom endocrine pathology society currently considers the presence of oncocytic features as an exclusion criterion for the use of the NIFTP diagnostic term [10].

In 2013, Ganly et al. studied a group of Hurthle cell carcinomas (HCC), i.e., carcinoma with invasion, oncocytic cytoplasm, absence of nuclear features of PTC and solid and/or follicular growth patterns [11]. In this study, HCC showed a distinct mutational, transcriptional and copy number profile different from those of PTC with a low frequency (being 11%) of *RAS* mutations and no *BRAF* mutations [11]. However, the molecular profile of oncocytic FVPTC has not yet been studied to date.

In the current retrospective multi-institutional study, we gathered and analyzed the clinical outcome of 61 patients with unifocal O-NI-EFVPTC from three tertiary hospitals who were not treated with post-operative RAI. A subgroup of 15 O-NI-EFVPTC were also subjected to targeted next generation sequencing to explore the molecular profile of these lesions and compare it to that of Hurthle cell adenoma (HCA), PTC, FVPTC, and HCC.

Methods

Study cohort and histopathologic review

After obtaining approval from the various institutional review boards, the pathology database of three tertiary hospitals, namely Memorial Sloan Kettering Cancer center, (MSKCC), New York, NY, USA, Sunnybrook Health Sciences Centre (SHSC), Toronto, ON, Canada, and Ospedale Maggiore, Bologna, Italy, were searched for candidate cases of unifocal O-NI-EFVPTC. All cases were reviewed independently by three endocrine pathologists (BX, GT, and RG) to confirm the diagnosis using the criteria proposed by Nikiforov et al. [5]. In brief, PTC NI-EFVPTC was diagnosed when a PTC fulfilled all of the following criteria: (1) encapsulation or clear demarcation; (2) exclusive/predominant follicular growth pattern lacking psammoma bodies and with <1% papillae and <30% solid growth pattern; (3) nuclear atypia in the form of nuclear enlargement, nuclear membrane irregularity and/or chromatin clearing with a nuclear score of 2–3; (4) absence of invasion (vascular or capsular); (5) no tumor necrosis; and (6) mitotic index <3 per 10 high power fields (400×). A tumor was considered as oncocytic only when at least 75% of the lesional cells exhibited unequivocal Hurthle cell/oncocytic phenotype as defined by the Armed Forces Institute of Pathology (AFIP) fascicle [12]. The tumor cells must show abundant eosinophilic granular cytoplasm (i.e., oncocytic/Hurthle cell phenotype, Fig. 1). As Hurthle cell lesions commonly have large nuclei with slight nuclear membrane irregularity, convincing diagnostic nuclear features of papillary thyroid carcinoma, e.g., marked nuclear enlargement, membrane irregularity and/or chromatin clearing, had to be present for a lesion to be considered as O-NI-EFVPTC. Additionally, the tumor cells typically lacked prominent central nucleoli as those seen in Hurthle cell neoplasm, but rather had peripherally located nucleoli. Patients whose tumor was less than 1 cm, who had separate foci of carcinoma, who had no follow up, or who received post-operative RAI were excluded from the current study. A total of 61 cases operated between 1984 and 2017 fulfilled the above inclusion criteria.

Clinical review

The patients' charts were reviewed to record the following clinical parameters: age at diagnosis, sex, surgery type (total thyroidectomy vs. lobectomy/hemithyroidectomy), duration of clinical follow up (FU) and clinical outcome. Recurrence was defined as structural recurrence confirmed by imaging and/or histopathologic examination.

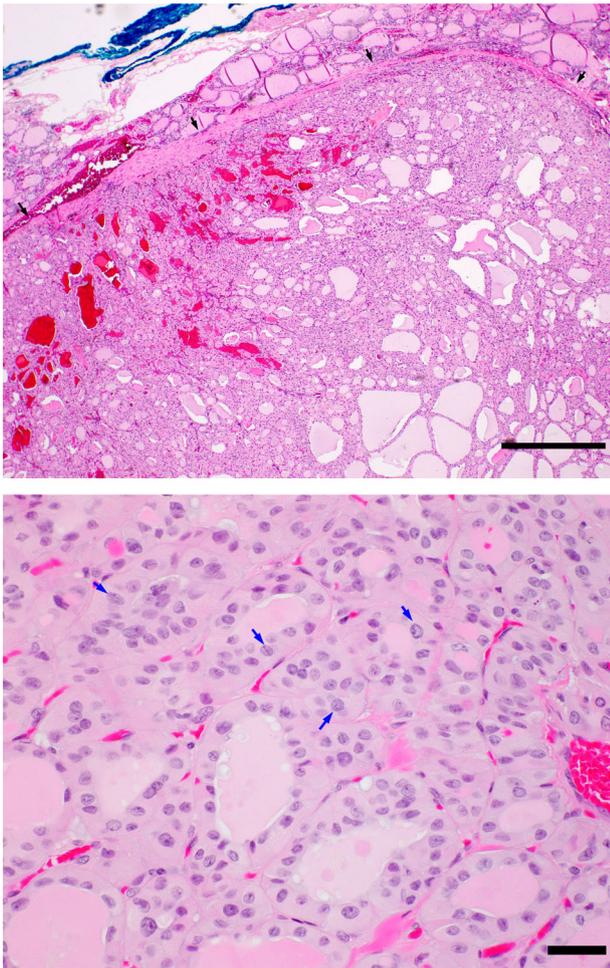


Fig. 1 Upper panel: Oncocyctic noninvasive encapsulated follicular variant of papillary thyroid carcinoma (O-NI-EFVPTC) is completely encapsulated (black arrows) without evidence of capsular or vascular invasion. Lower panel: The lesional cells show oncocytic features with abundant eosinophilic cytoplasm and nuclear features of papillary thyroid carcinoma with chromatin clearing and frequent nuclear grooves (blue arrows). Scale bars: 200 microns in upper panel, and 30 microns in lower panel

DNA extraction and targeted next generation sequencing

Identification of somatic DNA mutations

Fifteen O-NI-EFVPTC from the MSKCC cohort with tissue available for DNA extraction were subjected to the MSK-IMPACT™ (Memorial Sloan Kettering-Integrated Mutation Profiling of Actionable Cancer Targets) platform, an FDA (Food and Drug Administration)-approved deep-coverage, targeted next-generation sequencing assay as previously described [13, 14]. In brief, the MSK-IMPACT assay detects single nucleotide variants (SNVs), small

insertions/deletion (indels), copy number variants (CNVs) and fusion/structural variants in 468 oncogenes, using custom DNA probes designed for targeted sequencing of all exons and selected introns, including canonical and selected non-canonical transcripts. Genomic DNA from tumor and patient matched normal samples were extracted from formalin-fixed paraffin-embedded (FFPE) tissue and subjected to sequence library preparation (Kapa Biosystems) and exon capture (NimbleGen). Pooled libraries containing captured DNA fragments were subsequently sequenced on the Illumina HiSeq 2500 system. Paired-sample variant calling was performed on tumor samples and their respective matched normals to identify point mutations, SNVs and indels. MuTect (version 1.1.4) was used for SNV calling and SomaticIndelDetector, a tool in GATKv3.3.0, was used for detecting indel events. Variants were subsequently annotated using Annovar, and annotations relative to the canonical transcript for each gene (derived from a list of known canonical transcripts obtained from the UCSC genome browser) were reported. Annotated SNV and indel calls were subjected to a series of filtering steps to ensure only high-confidence calls were admitted to the final step of manual review [15].

Identification of copy number alterations

The FACETS analysis [16] was performed for copy number alteration. The FACETS algorithm is an open-source allele-specific copy number analysis tool, which can enhance the sensitivity to identify amplifications or deletions in tumors by joint modeling of total and allele-specific patterns, allowing reliable determination of total and allele-specific copy number call [16]. A minimum of 1.5-fold change was required to be considered as amplification or deletion.

Identification of mitochondrial DNA (mtDNA) mutations

Somatic mtDNA mutation analysis was performed using a custom informatics pipeline for mtDNA as previously described [17]. Briefly, a pileup file for paired tumor and normal tissue sample was generated using samtools mpileup with minimal mapping quality 10 and base alignment quality 13 [18]. Mutation annotation format files for mtDNA variants were then generated using vcf2maf, and further annotated with calls from MitImpact (including the APOGEE score and MitoTIP). Putative variants were filtered if they fell in regions 302–315, 514–525, or 3106–3110 of the mitochondrial rCRS. Variants were retained if they contained at least 5 reads in support of the variant, with at least 2 reads in both the forward and reverse direction. Variants were prioritized as putatively loss-of-function if they were (1) associated with disease in MITOMAP or (2) either a nonsense or frameshift variant.

Results

Clinico-pathologic findings in O-NI-EFVPTC

Sixty-one patients with unifocal O-NI-EFVPTC who did not receive RAI were included in this study. The number of included cases according to the institutions was as follows: MSKCC $n = 34$; Bologna-Ospedale Maggiore $n = 22$; and SHSC $n = 5$. All cases from MSKCC and SHSC (39 of 61, 64%) had an initial diagnosis of encapsulated FVPTC while the cases from Ospedale Maggiore were diagnosed as Hurthle cell adenomas 22 (36%) of 61.

The clinico-pathologic characteristics are summarized in Table 1. O-NI-EFVPTC predominantly affected female patients with a female to male ratio of 2.2:1. The median age of diagnosis was 56 (range: 8–82). The median tumor size was 2.5 cm (range 1.0–5.0 cm). Twenty-eight patients (46%) underwent lobectomy, while the remaining 33 (54%) had total thyroidectomy. In 37 (61%) cases, the entire tumor or tumor capsule was submitted for histologic examination, while in the remaining 24 patients (39%), the tumors were sampled representatively. In cases where the tumor and its capsule were representatively sampled, an average of 5 tumor sections (median = 4, range: 2–12) and 2 tumor sections per centimeter of tumor (median = 2, range: 1–4) were examined per case. All tumors were confined to the thyroid and were resected completely with negative surgical margins. Lymph node(s) were sampled in 12 (20%) patients. No lymph node metastases were detected at diagnosis clinically and/or pathologically. Benign conditions such as chronic lymphocytic thyroiditis and nodular hyperplasia, were observed in 9 (15%) and 28 (46%) patients, respectively.

Clinical outcome of O-NI-EFVPTC

The median follow up in our cohort was 5.3 years (range:0.1–20.5). Among them, 55 (87%) had at least 1-year follow up with a median follow up of 6.7 years, 52 (85%) had at least 2-year follow up with a median follow up of 6.9 years, 33 (54%) had at least 5-year follow up with a median follow up of 10.2 years, and 17 (28%) had at least 10-year follow up. No structural recurrence or disease specific death was observed in the entire cohort.

Molecular profile of O-NI-EFVPTC

The molecular profile of 15 cases of O-NI-EFVPTC is listed in Table 2.

Somatic mutations

The nuclear DNA mutation profile of O-NI-EFVPTC is illustrated in Fig. 2. Mutations in the *RAS* gene family

appear to be the main driver with alterations detected in 5 (33%) cases, with a particular preference for mutations to the homologous Q61 position. This included two mutations to the Q61 hotspot in *NRAS* (Q61R and Q61K, 13% of samples), two mutations to *KRAS* (13% of samples), including the hotspot mutation G12D, as well as a less common Q61R variant homologous to the *NRAS* Q61 variant. An additional mutation to *HRAS* at position Q61 was also observed and one tumor harbored a relatively rare *BRAF* K601E mutation. Additional mutations were detected in other cancer-associated genes including *TSHR* (thyroid stimulating hormone receptor), *EZH1* (enhancer of zest 1 polycomb repressive Complex 2 Subunit), *NFE2L2* (nuclear factor erythroid 2 like 2), *TET2* (TET methylcytosine dioxygenase 2), *MCL1*, *CDKN2C* (cyclin dependent kinase inhibitor 2C), *AURKA* (aurora kinase A) and *GLI* (glioma-associated oncogene family zinc finger 1): each was detected in one case (7%). Aggressive molecular signatures, e.g., *TP53* and *TERT* promoter mutations, were not seen in our cohort. No mutations were detected in 6 (40%) cases.

Fusion genes

None of the 15 tested O-NI-EFVPTC contained fusions that were reported in thyroid carcinoma [6], e.g., *PAX8-PPAR γ* , as well as those involving *RET*, *BRAF*, *NTRK3*, *ALK*, *THADA*, *FGFR1*, *MET*, and *LTK* loci. In addition, none of the fusion genes recently reported in HCC [17] were identified.

Copy number alterations

The FACETS plots for all 15 cases are shown in Supplementary Fig. 1. Seven cases (47%) had a copy number quiet profile with no detectable copy number gains or losses. Five cases (cases 3, 4, 5, 7, and 11) had focal chromosomal alterations in chromosomes 2, 4, 9, 21, and 22. Three cases showed quite different copy number alterations (cases 9, 10, and 12) in which there was whole chromosome duplication of chromosome 7 (Fig. 3). These tumors did not show the widespread uniparental disomy typical of HCC [17].

Mitochondrial DNA mutations

Nonsilent mitochondrial DNA mutations were detected in ten (67%) O-NI-EFVPTC, including one case (7%) which harbored two nonsilent mutations (Fig. 4, case 1 of Table 2). The 6 mtDNA encoding complex I subunits were enriched for mutations, being detected in 6 cases (40%). Of the 10 cases with mtDNA mutations, four (27% of all cases) were frameshift or nonsense mutations, which may lead to potential inactivation of mitochondrial respiration in these tumors and subsequent activation of mitochondrial biogenesis [19].

Table 1 Clinico-pathologic characteristics of patients with unifocal oncocytic noninvasive encapsulated follicular variant of papillary thyroid carcinoma (O-NI-EFVPTC) who did not receive post-operative radioactive iodine treatment

All patients (<i>n</i> = 61)			
Site	MSKCC, US	34	56%
	Bologna, Italy	22	36%
	SHSC, Canada	5	8%
Sex	Female	42	69%
	Male	19	31%
Age: mean, median (range)			52, 56 (8–82)
Surgical procedure	Lobectomy	28	46%
Total/subtotal thyroidectomy	33		54%
Tumor sampling	Entirely or entire capsule	37	61%
Representative	24		39%
Number of sections sampled per tumor in cases that were representatively sampled: mean, median (range)			5, 4 (2–12)
Number of sections per cm of tumor in cases that were representatively sampled: mean, median (range)			2,2 (1–4)
Tumor size(cm): mean, median (range)			2.5, 2.5 (1.0–5.0)
Background thyroid	Chronic lymphocytic thyroiditis	9	15%
Nodular hyperplasia	28		46%
Other/none	25		41%
Sampling and status of lymph nodes	Not sampled	49	80%
	Benign lymph node(s)	12	20%
Post-operative radioactive iodine	None	61	100%
Follow up (FU) duration (years): mean, median (range)			7.2, 5.3 (0.1–20.5)
Disease status at last FU	No evidence of disease (NED)	61	100%
Patients with at least 1-year FU (<i>n</i> = 55)			
Surgical procedure	Lobectomy	26	47%
	Total/subtotal thyroidectomy	29	53%
FU duration (years): mean, median (range)			7.9, 6.7 (1–20.5)
Disease status at last FU	NED	55	100%
Patients with at least 2-year FU (<i>n</i> = 52)			
Surgical procedure	Lobectomy	23	44%
	Total/subtotal thyroidectomy	29	56%
FU duration (years): mean, median (range)			8.3, 6.9 (2.7 -20.5)
Disease status at last FU	NED	52	100%
Patients with at least 5-year FU (<i>n</i> = 33)			
Surgical procedure	Lobectomy	17	52%
	Total/subtotal thyroidectomy	16	48%
FU duration(years):mean, median (range)			10.9, 10.2 (5.1–20.5)
Disease status at last FU	NED	33	100%

Discussion

In the current study, we examined the clinical outcome and molecular profile of a large cohort of oncocytic NI-

EFVPTC with predominant oncocytic cytology but otherwise fulfilling the initial diagnostic criteria of NIFTP in order to determine the clinical behaviors and molecular signatures of these lesions. Only unifocal cases of

Table 2 Molecular profile of O-NI-EFVPTC

Case	RAS and genotyped <i>BRAF</i>	Other mutations	Copy number alteration	Mitochondrial DNA mutation
1	<i>NRAS</i> p. Q61K	<i>GLI</i> p.E316Q	–	MT-CO1 nonsense mutation G6930 AMT-ND5 missense mutation T12569C
2	<i>NRAS</i> p. Q61R	–	–	–
3	<i>KRAS</i> p. G12D	<i>AURKA</i> p.L159fs	Loss in part of 9	MT-TS2 3' flank T12228C
4	<i>KRAS</i> p. Q61R	–	Loss in 21	MT-TL1 5' flank A3243G
5	<i>HRAS</i> p. Q61R	<i>CDKN2C</i> p.D67N	Loss of 22	MY-CYB missense mutation T15729C
6	<i>BRAF</i> p. K601E	<i>TET2</i> p. V1718LM <i>CLL1</i> p. E41A	–	MT-ND1 missense mutation c.G3901A
7	–	<i>NFE2L2</i> p.29_32del	Gain of 9p	MT-RNR1 5' flank G709A
8	–	<i>EZH1</i> p.Q571R	–	MT-ND4 frame shift deletion A11038-
9	–	<i>TSHR</i> p.M453T	Gain of 5, 10, 12, 17, 18, 20, 21, X; WCD of 7; Copy neutral LOH in 9	–
10	–	–	Gain of 5, 9, 12, 16, 20; WCD of 7; Copy neutral LOH in X	–
11	–	–	Loss of 2p; balanced gain in 4	–
12	–	–	Gain of 4, 12, 18, 21; WCD of 7	MT-ND2 missense mutation T5412C
13	–	–	–	MT-ND4 nonsense mutation G11390A
14	–	–	–	MT-ND4 frame shift insertion -10952C
15	–	–	–	–

Note: Frameshift or nonsense mutations in bold characters

WCD whole chromosome duplication, LOH loss of heterozygosity

O-NI-EFVPTC without separate carcinoma foci were included in this study to avoid the confounding effect of multifocal disease. Additionally, we only included patients treated with surgery alone (lobectomy or total thyroidectomy) without post-operative RAI, as recommended by the ATA guidelines [20, 21]. This enabled us to follow the natural history of resected O-NI-EFVPTC. With regard to follow up, we relied on structural rather than biochemical recurrence to assess patients' disease status. This was partly due to the fact that some cases were old and did not have adequate serum thyroglobulin data. Although a long follow up time was not available in all cases, this study comprised 33 patients, each followed for at least 5 years, treated by surgery without RAI therapy who did not recur with a median follow up of 10.2 years. Seventeen patients had at least 10-year follow up. As the literature has shown that most differentiated thyroid carcinomas recur during the first decade [22, 23], the outcome data from the present study suggest that O-NI-EFVPTC may follow a very indolent behavior similar to their non-oncocyctic counterparts [5, 24].

Future studies with larger cohort size and longer follow up may be required to confirm the indolent nature of these tumors. The definition of NIFTP is in constant evolution. The most recent suggested criteria requires a total lack of true papillary formations [9]. Despite the fact that we used the less stringent initial criterion of less than 1% papillae, none of the patients developed nodal metastasis. The lack of nodal disease in our cohort was also consistent with the fact that O-NI-EFVPTC behaves similarly to other encapsulated follicular neoplasms [25], in contrast to *BRAFV600E*-mutated tumors that are typically prone to develop local lymph nodes metastases [12, 26]. Additional studies with longer follow up may be required to specifically address the long-term outcome of the patients with these tumors.

Interestingly, the molecular profile of O-NI-EFVPTC straddles between FVPTC/NIFTP and HCC, a thyroid carcinoma that shares the eosinophilic cytoplasmic features of O-NI-EFVPTC but lacks the nuclear features of PTC (Table 3) [5–7, 11]. The mutation landscape of O-NI-EFVPTC largely resembles those of FVPTC and NIFTP

Fig. 2 Somatic mutations in O-NI-EFVPTC. Oncoplot shows that O-NI-EFVPTC is enriched with *RAS* mutations. The only *BRAF* mutation found is *BRAF* K601E



and differs significantly from Hurthle cell neoplasms. *NRAS*, *KRAS*, and *HRAS* hotspot mutations are detected in a significant proportion (33%) of O-NI-EFVPTC, comparable with the 37% frequency found in the FVPTC of the TCGA PTC study [6, 27], 30% in the NIFTP consensus cohort [5] and 63% of NIFTP reported by Johnson et al. [7]. However, the *NRAS*, *KRAS*, and *HRAS* mutations in O-NI-EFVPTC are significantly higher than in Hurthle cell neoplasms, being 9–11% in HCC and 0% in Hurthle cell adenoma [11, 17, 28]. *BRAF* V600E, a driver mutation commonly seen in classical variant and tall cell variant of PTC, is absent in O-NI-EFVPTC (current study), as well as in NIFTP, Hurthle cell adenoma, and HCC as shown in previous studies [5–7, 11, 17]. A single case of O-NI-EFVPTC harbored *BRAF* K601E, which was also described in 1 to 3% of FVPTC and NIFTP [5–7].

Additional molecular alterations observed in O-NI-EFVPTC include *GLI*, *AURKA*, *CDKN2C*, *TET2*, *MCL1*, *NFE2L2*, *EZH1*, and *TSHR* mutations. *CDKN2C*, *EZH1* and *TSHR* mutations were also detected in four PTCs in the TCGA cohort: including three cases each with

CDKN2C, *EZH1* or *TSHR* mutation and one case with both *TSHR* and *EZH1* mutation. Interestingly, all four cases are follicular variant, including two O-NI-EFVPTC, as shown in cBioPortal [27, 29]. *CDKN2C* encodes a protein that belongs to the INK4 family of cyclin-dependent kinase inhibitors, which functions as a cell growth regulator by controlling cell cycle G1 progression. *EZH1* is the catalytic subunit of PRC2 complex which mediates methylation of histone H3 lys27 (H3K27), leading to transcription repression of the targeted genes. *TSHR* encodes thyroid stimulating hormone receptor. The contribution of these gene mutations to the pathogenesis of O-NI-EFVPTC remains to be investigated.

No gene fusions were detected in this cohort of O-NI-EFVPTC. In comparison, *PPAR γ* and *THADA* rearrangement has been reported in 22% of NIFTP [5] and 1 to 3% of FVPTC in the TCGA cohort [6, 27]. We do not know the reason behind this lack of gene fusions in O-NI-EFVPTC.

Oncocytic change in thyroid neoplasms is the result of an aberrant increase in mitochondrial mass [30], and has been described in a spectrum of thyroid follicular-cell derived

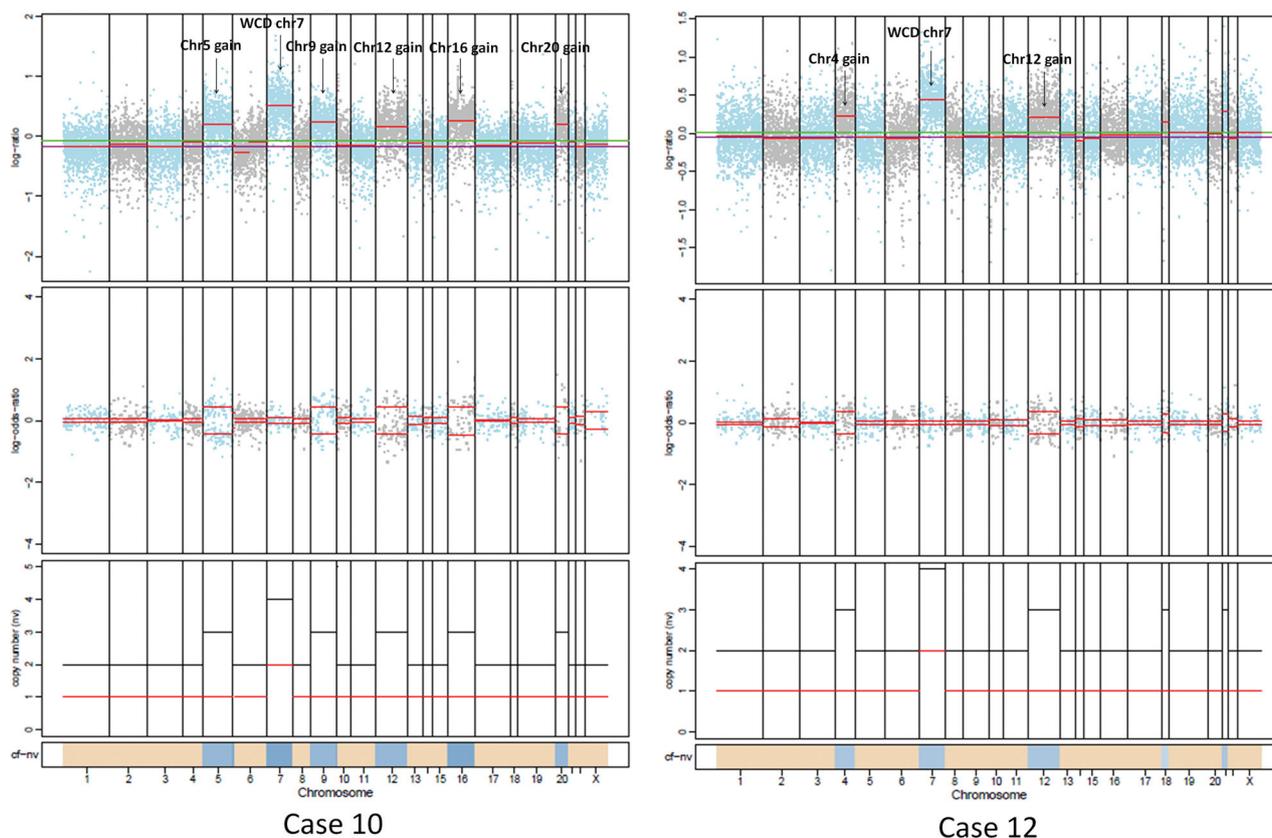


Fig. 3 Allele specific copy number alterations in case 10 and 12. In case 10, the tumor has whole chromosome duplication of chromosome 7 in addition to gain of Chromosomes 5, 9, 12, 16 and 20. In case 12,

the tumor has whole chromosome duplication of chromosome 7 in addition to gain of chromosome 4 and 12

neoplasms, including Hurthle cell adenoma, HCC, and various variants of papillary thyroid carcinoma [4]. This accumulation of mitochondria has been linked to mutations in the genes coding for some of the subunits of the five multimeric complexes of the oxidative phosphorylation (OXPHOS) system localized to the inner mitochondrial membrane [31, 32]. If mutations of these subunits render them missing/defective, the entire multimeric complex does not assemble properly, OXPHOS is impaired, and there is a compensatory accumulation of mitochondria, with phenotypic (i.e., accumulation of mitochondria), biochemical and metabolic defects [31, 32]. In the past decade a high frequency of somatic mitochondrial DNA variants has been reported in oncocyctic thyroid neoplasms [17, 30, 32, 33], and much less frequently in nuclear coded mitochondrial genes [17, 30, 33]. As high as 71% of HCC harbors mtDNA mutations, with 37% of tumors demonstrating detrimental frameshift or nonsense variants [17].

Among the 15 O-NI-EFVPTC tested, a very similar mtDNA mutation profile (compared to HCC) was identified with 10 cases (67%) showing nonsilent mtDNA mutations,

including three cases with frameshift or nonsense mutations. This high rate of mtDNA mutations is consistent with the oncocyctic phenotype of O-NI-EFVPTC and further confirms the importance of mtDNA in the development of oncocyctic thyroid neoplasms. The similar rate of mtDNA mutations in the extremely indolent O-NI-EFVPTC and the much more aggressive HCC suggests that mitochondrial DNA mutations are probably not sufficient to cause an aggressive behavior in oncocyctic thyroid tumors.

Multiple previous studies, including our own, have reported a plethora of chromosomal aberrations in Hurthle cell neoplasms using array comparative genomic hybridization and next generation sequencing techniques [11, 17, 28, 30, 33, 34]. Ganly et al. recently reported the unique chromosomal alterations seen in 56 cases of HCC (18). In this study, the more aggressive widely invasive HCC had whole chromosome duplication of chromosome 7 and 5 with global uniparental disomy (UPD) of the remaining chromosomes. This global UPD results in widespread loss of heterozygosity (Table 3). Numerous chromosomal gains and losses involving chromosome 1, 2, 3, 4, 5, 6, 7, 8, 10,

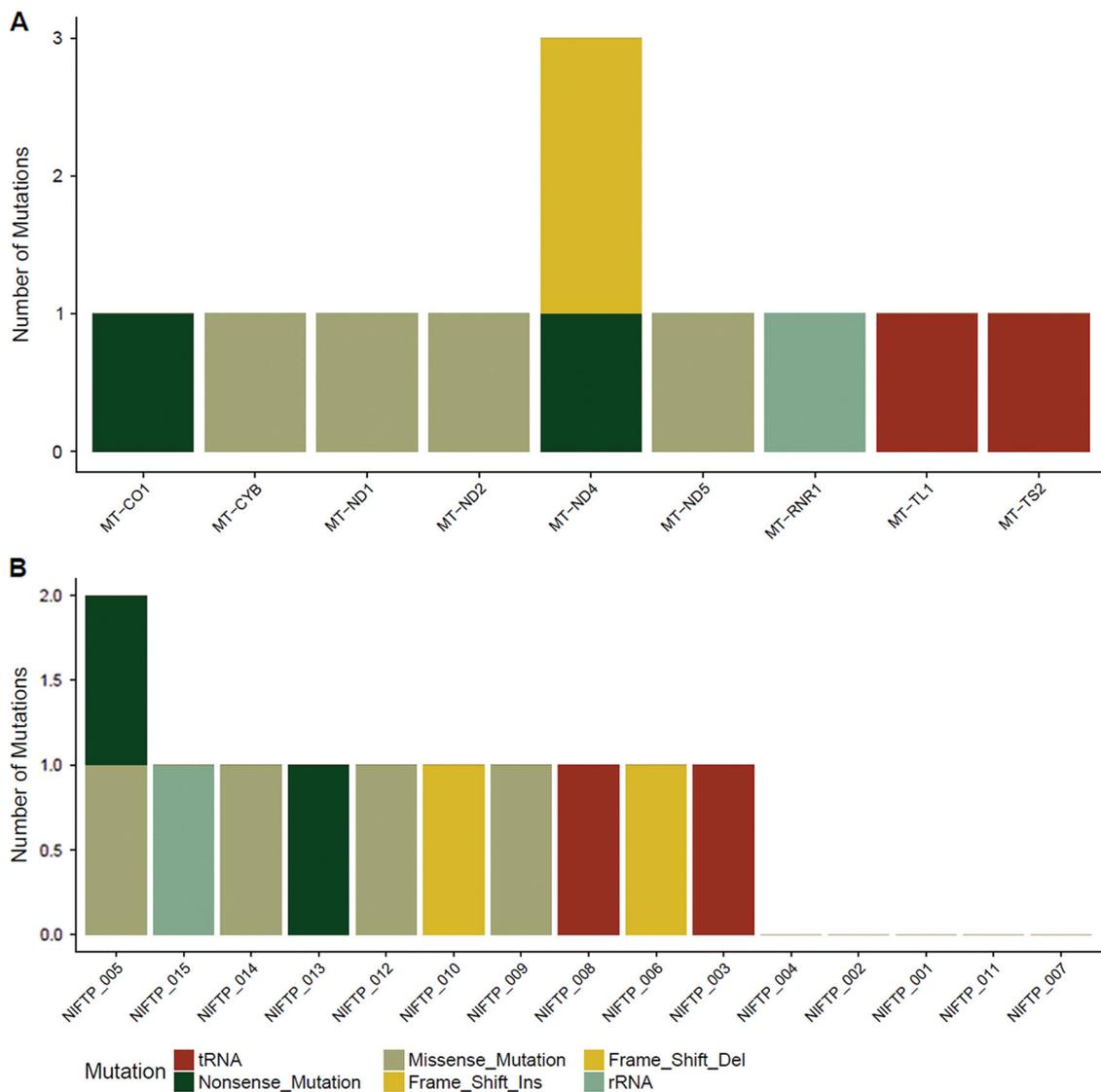


Fig. 4 Mitochondrial DNA (mtDNA) mutations in O-NI-EFVPTC. **a** Distribution of non-synonymous, tRNA, and rRNA mutations across mtDNA-encoded genes in 15 cases of O-NI-EFVPTC. **b** mtDNA mutation(s) in each tested O-NI-EFVPTC sample

12, 16, 17, 19, 20, and 22 have been described previously in HCC (Table 3). Among O-NI-EFVPTC, several tumors lacked any chromosome alteration and 5 showed single chromosome alterations. Three tumors showed whole chromosome duplication of chromosome 7 as seen in HCC. However, these tumors did not show the widespread UPD that is seen in HCC [17].

In conclusion, the above data strongly suggest that O-NI-EFVPTC are similar to NIFTP at the molecular level in regard to their MAPK alterations (enrichment in RAS-like mutations) and at the outcome level having an indolent clinical behavior with negligible risk of metastasis and recurrence, even when treated conservatively without post-operative RAI. These findings further reinforce the fact that invasion rather than nuclear or cytoplasmic features drives

outcome in encapsulated follicular patterned tumors [35, 36]. The presence of mitochondrial DNA mutations confirms that O-NI-EFVPTC are part of the spectrum of oncocytic neoplasms, and further supports the role of mitochondrial DNA mutations in the acquisition of the oncocytic phenotype. Therefore, given the molecular and behavioral similarity, it is worth considering rebranding oncocytic noninvasive encapsulated follicular variant of papillary thyroid carcinoma as NIFTP just as its non-oncocytic counterpart. A note in the pathology report should mention the presence of oncocytic features. Including O-NI-EFVPTC in the NIFTP category will help reduce unnecessary treatments with its side effects and will prevent the adverse psychosocial and financial consequences of a cancer diagnosis.

Table 3 Molecular profile in follicular variant of papillary thyroid carcinoma (FVPTC, The Cancer Genomic Atlas TCGA cohort, data retrieved from cBioPortal(1)), noninvasive follicular thyroid neoplasm with papillary-like nuclear features (NIFTP), Hurthle cell carcinoma (HCC), Hurthle cell adenoma (HCA) and O-NI-EFVPTC

	FVPTC (<i>n</i> = 99, TCGA) (1, 2)	NIFTP (<i>n</i> = 27, Nikiforov) (3)	NIFTP (<i>n</i> = 32, Johnson) (4)	HCC (<i>n</i> = 27, Ganly) (5)	HCC (<i>n</i> = 56, Ganly) (6)	HCA (<i>n</i> = 8) (5)	O-NI-EFVPTC Current study (<i>n</i> = 15)
Mutations							
<i>NRAS</i>	25%	19%	34%	11%	9%	0	13%
<i>HRAS</i>	10%	7%	19%	0	0	0	7%
<i>KRAS</i>	2%	4%	9%	0	0	0	13%
<i>BRAF</i> K601E	2%	3%	3%	0	0	0	7%
Total <i>RAS</i> / <i>BRAF</i> K601E	39%	33%	66%	11%	9%	0	40%
Fusions							
<i>RET</i> (4%)		<i>PPARγ</i> (22%)	NA	<i>PPARγ</i> (0)	<i>PPARγ</i> (0)	None	None
<i>THADA</i> (3%)		<i>THADA</i> (22%)			<i>THADA</i> (0)		
<i>BRAF</i> (3%)					<i>CHCHD10-VPREB3</i> (13%)		
<i>PPARγ</i> (1%)					<i>HEPHL1-PANX1</i> (9%)		
<i>MET</i> (1%)					<i>TMEM233-PRKAB1</i> (9%)		
<i>NTRK3</i> (1%)							
Copy number alterations							
NA for FVPTC	NA	NA	NA	Gain: 4p, 5p, 6p, 7p, 8p, 10p, 12p, 16q	Gain: 1q, 2q, 3q, 5p, 7p, 10q, 12q, 14q, 20p, Xp. WCD of chr 5, 7, 12 in widely invasive HCC	NA	Gain: 4, 5, 9, 10, 12, 16, 17, 18, 20, 21. WCD chr 7. Loss: 2p, 9, 21, 22
PTC: 1q gain (15%) 22q loss (10%) High frequency of focal gains/losses (2.4%) Copy number quiet (73%)				Loss: 4q, 6p, 7p, 9q, 12q, 16q	Loss: 1q, 2q, 3q, 4q, 7q, 9q, 10q, 11p, 15q, 16q, 17q. Global UPD in widely invasive HCC.		
Mitochondria DNA							
NA	NA	NA	NA	NA	71% with nonsilent mutations, including 37% of frameshift or nonsense mutations	NA	67% with nonsilent mutations, including 27% of frameshift or nonsense mutations

NA not available, UPD uniparental disomy, WCD whole chromosome duplication

Funding Research reported in this publication was supported in part by the Cancer Center Support Grant of the National Institutes of Health/National Cancer Institute under award number P30CA008748. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. Research reported in this publication was also supported in part by an Italian Government-Ministero della Salute Grant No. RF-2011-02350857 (to G.T.)

Compliance with ethical standards

Conflict of interest All authors declare that they have no conflict of interest.

Ethical approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. This article does not contain any studies with animals performed by any of the authors.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

References

1. H. Lim, S.S. Devesa, J.A. Sosa, D. Check, C.M. Kitahara, Trends in thyroid cancer incidence and mortality in the United States, 1974–2013. *JAMA* **317**(13), 1338–1348 (2017). <https://doi.org/10.1001/jama.2017.2719>
2. R.L. Siegel, K.D. Miller, A. Jemal, Cancer statistics, 2018. *CA Cancer J. Clin.* **68**(1), 7–30 (2018). <https://doi.org/10.3322/caac.21442>
3. C.K. Jung, M.P. Little, J.H. Lubin, A.V. Brenner, S.A. Wells Jr., A.J. Sigurdson, Y.E. Nikiforov, The increase in thyroid cancer incidence during the last four decades is accompanied by a high frequency of BRAF mutations and a sharp increase in RAS mutations. *J. Clin. Endocrinol. Metab.* **99**(2), E276–E285 (2014). <https://doi.org/10.1210/jc.2013-2503>
4. R.V. Lloyd, R.Y. Osamura, G. Kloppel, J. Rosai. *WHO classification of tumours of endocrine organs*. (International Agency for Research on Cancer (IARC), Lyon), 2017)
5. Y.E. Nikiforov, R.A. Seethala, G. Tallini, Z.W. Baloch, F. Basolo, L.D. Thompson, J.A. Barletta, B.M. Wenig, A. Al Ghuzlan, K. Kakudo, T.J. Giordano, V.A. Alves, E. Khanafshar, S.L. Asa, A.K. El-Naggar, W.E. Gooding, S.P. Hodak, R.V. Lloyd, G. Maytal, O. Mete, M.N. Nikiforova, V. Nose, M. Papotti, D.N. Poller, P.M. Sadow, A.S. Tischler, R.M. Tuttle, K.B. Wall, V.A. LiVolsi, G.W. Randolph, R.A. Ghossein, Nomenclature revision for encapsulated follicular variant of papillary thyroid carcinoma: a paradigm shift to reduce overtreatment of indolent tumors. *JAMA Oncol.* **2**(8), 1023–1029 (2016). <https://doi.org/10.1001/jamaoncol.2016.0386>
6. Cancer Genome Atlas Research, N., Integrated genomic characterization of papillary thyroid carcinoma. *Cell* **159**(3), 676–690 (2014). <https://doi.org/10.1016/j.cell.2014.09.050>
7. D.N. Johnson, L.V. Furtado, B.C. Long, C.J. Zhen, M. Wurst, I. Mujacic, S. Kadri, J.P. Segal, T. Antic, N.A. Cipriani, Non-invasive follicular thyroid neoplasms with papillary-like nuclear features are genetically and biologically similar to adenomatous nodules and distinct from papillary thyroid carcinomas with extensive follicular growth. *Arch. Pathol. Lab. Med.* **142**(7), 838–850 (2018). <https://doi.org/10.5858/arpa.2017-0118-OA>
8. J. Liu, B. Singh, G. Tallini, D.L. Carlson, N. Katabi, A. Shaha, R.M. Tuttle, R.A. Ghossein, Follicular variant of papillary thyroid carcinoma: a clinicopathologic study of a problematic entity. *Cancer* **107**(6), 1255–1264 (2006). <https://doi.org/10.1002/cncr.22138>
9. R.V. Lloyd, S.L. Asa, V.A. LiVolsi, P.M. Sadow, A.S. Tischler, R.A. Ghossein, R.M. Tuttle, Y.E. Nikiforov, The evolving diagnosis of noninvasive follicular thyroid neoplasm with papillary-like nuclear features (NIFTP). *Hum. Pathol.* **74**, 1–4 (2018). <https://doi.org/10.1016/j.humpath.2017.12.027>
10. S.J. Johnson, T.J. Stephenson, D.N. Poller, NIFTP addendum to the RCPATH Dataset for thyroid cancer histopathology reports. <http://www.ukeps.com/docs/niftp.pdf> (2016).
11. I. Ganly, J. Ricarte Filho, S. Eng, R. Ghossein, L.G. Morris, Y. Liang, N. Socci, K. Kannan, Q. Mo, J.A. Fagin, T.A. Chan, Genomic dissection of Hurthle cell carcinoma reveals a unique class of thyroid malignancy. *J. Clin. Endocrinol. Metab.* **98**(5), E962–E972 (2013). <https://doi.org/10.1210/jc.2012-3539>
12. J. Rosai, R.A. DeLellis, M.L. Carcangiu, W.J. Frable, T. Giovanni. *Tumor of the thyroid and parathyroid gland (AFIP atlas of tumor pathology series 4)*. (American Registry of Pathology Press, Silver Spring, MD), 2015)
13. D.T. Cheng, T.N. Mitchell, A. Zehir, R.H. Shah, R. Benayed, A. Syed, R. Chandramohan, Z.Y. Liu, H.H. Won, S.N. Scott, A.R. Brannon, C. O'Reilly, J. Sadowska, J. Casanova, A. Yannes, J.F. Hechtman, J. Yao, W. Song, D.S. Ross, A. Oultache, S. Dogan, L. Borsu, M. Hameed, K. Nafa, M.E. Arcila, M. Ladanyi, M.F. Berger, Memorial Sloan Kettering-Integrated Mutation Profiling of Actionable Cancer Targets (MSK-IMPACT): a hybridization capture-based next-generation sequencing clinical assay for solid tumor molecular oncology. *J. Mol. Diagn.* **17**(3), 251–264 (2015). <https://doi.org/10.1016/j.jmoldx.2014.12.006>
14. L.G. Morris, R. Chandramohan, L. West, A. Zehir, D. Chakravarty, D.G. Pfister, R.J. Wong, N.Y. Lee, E.J. Sherman, S.S. Baxi, I. Ganly, B. Singh, J.P. Shah, A.R. Shaha, J.O. Boyle, S.G. Patel, B.R. Roman, C.A. Barker, S.M. McBride, T.A. Chan, S. Dogan, D.M. Hyman, M. F. Berger, D.B. Solit, N. Riaz, A.L. Ho. The molecular landscape of recurrent and metastatic head and neck cancers: insights from a precision oncology sequencing platform. *JAMA Oncol.* **3**(2), 244–255 (2017). <https://doi.org/10.1001/jamaoncol.2016.1790>
15. J.T. Robinson, H. Thorvaldsdottir, W. Winckler, M. Guttman, E.S. Lander, G. Getz, J.P. Mesirov, Integrative genomics viewer. *Nat. Biotechnol.* **29**(1), 24–26 (2011). <https://doi.org/10.1038/nbt.1754>
16. R. Shen, V.E. Seshan, FACETS: allele-specific copy number and clonal heterogeneity analysis tool for high-throughput DNA sequencing. *Nucleic Acids Res.* **44**(16), e131 (2016). <https://doi.org/10.1093/nar/gkw520>
17. I. Ganly, V. Makarov, S. Deraje, Y. Dong, E. Reznik, V. Seshan, G. Nanjangud, S. Eng, P. Bose, F. Kuo, L.G.T. Morris, I. Landa, P.B. Carrillo Alborno, N. Riaz, Y.E. Nikiforov, K. Patel, C. Umbricht, M. Zeiger, E. Kebebew, E. Sherman, R. Ghossein, J.A. Fagin, T.A. Chan, Integrated genomic analysis of hurthle cell cancer reveals oncogenic drivers, recurrent mitochondrial mutations, and unique chromosomal landscapes. *Cancer Cell.* **34**(2), 256–270.e255 (2018). <https://doi.org/10.1016/j.ccell.2018.07.002>
18. H. Li, B. Handsaker, A. Wysoker, T. Fennell, J. Ruan, N. Homer, G. Marth, G. Abecasis, R. Durbin, The Sequence Alignment/Map format and SAMtools. *Bioinformatics* **25**(16), 2078–2079 (2009). <https://doi.org/10.1093/bioinformatics/btp352>
19. E. Reznik, Q. Wang, K. La, N. Schultz, C. Sander, Mitochondrial respiratory gene expression is suppressed in many cancers. *eLife* **6**, PMID: 28099114 (2017). <https://doi.org/10.7554/eLife.21592>
20. B.R. Haugen, A.M. Sawka, E.K. Alexander, K.C. Bible, P. Caturegli, G.M. Doherty, S.J. Mandel, J.C. Morris, A. Nassar, F. Pacini, M. Schlumberger, K. Schuff, S.I. Sherman, H. Somersat, J. A. Sosa, D.L. Steward, L. Wartofsky, M.D. Williams, American Thyroid Association Guidelines on the Management of Thyroid Nodules and Differentiated Thyroid Cancer Task Force Review and Recommendation on the Proposed Renaming of Encapsulated

- Follicular Variant Papillary Thyroid Carcinoma Without Invasion to Noninvasive Follicular Thyroid Neoplasm with Papillary-Like Nuclear Features. *Thyroid*. **27**(4), 481–483 (2017). <https://doi.org/10.1089/thy.2016.0628>
21. B.R.M. Haugen, E.K. Alexander, K.C. Bible, G. Doherty, S.J. Mandel, Y.E. Nikiforov, F. Pacini, G. Randolph, A. Sawka, M. Schlumberger, K.G. Schuff, S.I. Sherman, J.A. Sosa, D. Steward, R.M.M. Tuttle, L. Wartofsky, 2015 American Thyroid Association Management Guidelines for Adult Patients with Thyroid Nodules and Differentiated Thyroid Cancer. *Thyroid*. **26**, 1–133 (2016). <https://doi.org/10.1089/thy.2015.0020>
 22. E.L. Mazzaferri, S.M. Jhiang, Long-term impact of initial surgical and medical therapy on papillary and follicular thyroid cancer. *Am. J. Med.* **97**(5), 418–428 (1994)
 23. J.F. Nwatsock, D. Taieb, F.D. Zok, O. Mundler, Late recurrences of thyroid carcinoma 24 years after a complete remission: when monitoring should be stopped? *World J. Nucl. Med.* **11**(1), 42–43 (2012). <https://doi.org/10.4103/1450-1147.98749>
 24. L.D. Thompson, Ninety-four cases of encapsulated follicular variant of papillary thyroid carcinoma: a name change to non-invasive follicular thyroid neoplasm with papillary-like nuclear features would help prevent overtreatment. *Mod. Pathol.* **29**(7), 698–707 (2016). <https://doi.org/10.1038/modpathol.2016.65>
 25. M. Rivera, J. Ricarte-Filho, J. Knauf, A. Shaha, M. Tuttle, J.A. Fagin, R.A. Ghossein, Molecular genotyping of papillary thyroid carcinoma follicular variant according to its histological subtypes (encapsulated vs infiltrative) reveals distinct BRAF and RAS mutation patterns. *Mod. Pathol.* **23**(9), 1191–1200 (2010). <https://doi.org/10.1038/modpathol.2010.112>
 26. M. Rivera, R.M. Tuttle, S. Patel, A. Shaha, J.P. Shah, R.A. Ghossein, Encapsulated papillary thyroid carcinoma: a clinicopathologic study of 106 cases with emphasis on its morphologic subtypes (histologic growth pattern). *Thyroid* **19**(2), 119–127 (2009). <https://doi.org/10.1089/thy.2008.0303>
 27. E. Cerami, J. Gao, U. Dogrusoz, B.E. Gross, S.O. Sumer, B.A. Aksoy, A. Jacobsen, C.J. Byrne, M.L. Heuer, E. Larsson, Y. Antipin, B. Reva, A.P. Goldberg, C. Sander, N. Schultz, The cBio cancer genomics portal: an open platform for exploring multi-dimensional cancer genomics data. *Cancer Discov.* **2**(5), 401–404 (2012). <https://doi.org/10.1158/2159-8290.cd-12-0095>
 28. G. Tallini, A. Hsueh, S. Liu, G. Garcia-Rostan, M.R. Speicher, D. C. Ward, Frequent chromosomal DNA unbalance in thyroid oncogenic (Hurthle cell) neoplasms detected by comparative genomic hybridization. *Lab. Invest.* **79**(5), 547–555 (1999)
 29. J. Gao, B.A. Aksoy, U. Dogrusoz, G. Dresdner, B. Gross, S.O. Sumer, Y. Sun, A. Jacobsen, R. Sinha, E. Larsson, E. Cerami, C. Sander, N. Schultz, Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci. Signal.* **6** (269), p11 (2013). <https://doi.org/10.1126/scisignal.2004088>
 30. G. Gasparre, E. Bonora, G. Tallini, G. Romeo, Molecular features of thyroid oncogenic tumors. *Mol. Cell. Endocrinol.* **321**(1), 67–76 (2010). <https://doi.org/10.1016/j.mce.2010.02.022>
 31. E. Bonora, A.M. Porcelli, G. Gasparre, A. Biondi, A. Ghelli, V. Carelli, A. Baracca, G. Tallini, A. Martinuzzi, G. Lenaz, M. Rugolo, G. Romeo, Defective oxidative phosphorylation in thyroid oncogenic carcinoma is associated with pathogenic mitochondrial DNA mutations affecting complexes I and III. *Cancer Res.* **66**(12), 6087–6096 (2006). <https://doi.org/10.1158/0008-5472.can-06-0171>
 32. G. Gasparre, A.M. Porcelli, E. Bonora, L.F. Pennisi, M. Toller, L. Iommarini, A. Ghelli, M. Moretti, C.M. Betts, G.N. Martinelli, A. R. Ceroni, F. Curcio, V. Carelli, M. Rugolo, G. Tallini, G. Romeo, Disruptive mitochondrial DNA mutations in complex I subunits are markers of oncogenic phenotype in thyroid tumors. *Proc. Natl Acad. Sci. USA* **104**(21), 9001–9006 (2007). <https://doi.org/10.1073/pnas.0703056104>
 33. V. Maximo, T. Botelho, J. Capela, P. Soares, J. Lima, A. Taveira, T. Amaro, A.P. Barbosa, A. Preto, H.R. Harach, D. Williams, M. Sobrinho-Simoes, Somatic and germline mutation in GRIM-19, a dual function gene involved in mitochondrial metabolism and cell death, is linked to mitochondrion-rich (Hurthle cell) tumours of the thyroid. *Br. J. Cancer* **92**(10), 1892–1898 (2005). <https://doi.org/10.1038/sj.bjc.6602547>
 34. N. Wada, Q.Y. Duh, D. Miura, L. Brunaud, M.G. Wong, O.H. Clark, Chromosomal aberrations by comparative genomic hybridization in hurthle cell thyroid carcinomas are associated with tumor recurrence. *J. Clin. Endocrinol. Metab.* **87**(10), 4595–4601 (2002). <https://doi.org/10.1210/jc.2002-020339>
 35. I. Ganly, L. Wang, R.M. Tuttle, N. Katabi, G.A. Ceballos, H.R. Harach, R. Ghossein, Invasion rather than nuclear features correlates with outcome in encapsulated follicular tumors: further evidence for the reclassification of the encapsulated papillary thyroid carcinoma follicular variant. *Hum. Pathol.* **46**(5), 657–664 (2015). <https://doi.org/10.1016/j.humpath.2015.01.010>
 36. B. Xu, G. Tallini, T. Scognamiglio, B.R. Roman, R.M. Tuttle, R. A. Ghossein, Outcome of large noninvasive follicular thyroid neoplasm with papillary-like nuclear features. *Thyroid*. **27**(4), 512–517 (2017). <https://doi.org/10.1089/thy.2016.0649>

Affiliations

Bin Xu¹ · Ed Reznik² · R. Michael Tuttle³ · Jeffrey Knauf³ · James A. Fagin³ · Nora Katabi⁴ · Snjezana Dogan⁴ · Nathaniel Aleynick⁴ · Venkatraman Seshan² · Sumit Middha⁴ · Danny Enepekides⁵ · Gian Piero Casadei⁶ · Erica Solaroli⁷ · Giovanni Tallini⁸ · Ronald Ghossein⁴ · Ian Ganly⁹

¹ Department of Laboratory Medicine and Molecular Diagnostics, Sunnybrook Health Sciences Centre, Toronto, ON, Canada

² Department of Epidemiology and Biostatistics, Memorial Sloan Kettering Cancer center, New York, NY, USA

³ Department of Medicine, Memorial Sloan Kettering Cancer center, New York, NY, USA

⁴ Department of Pathology, Memorial Sloan Kettering Cancer center, New York, NY, USA

⁵ Department of Otolaryngology, Sunnybrook Health Sciences

Centre, Toronto, ON, Canada

⁶ Anatomic Pathology Unit, Ospedale Maggiore, Bologna, Italy

⁷ Endocrinology Unit, Ospedale Maggiore, Bologna, Italy

⁸ Department of Experimental, Diagnostic and Specialty Medicine-Anatomic Pathology, University of Bologna School of Medicine, Bologna, Italy

⁹ Department of Surgery, Memorial Sloan Kettering Cancer center, New York, NY, USA