



Recent Advances in the Neurobiology of Altered Motivation Following Bariatric Surgery

Julianna N. Brutman¹ · Sunil Sirohi² · Jon F. Davis¹

Published online: 9 November 2019
© Springer Science+Business Media, LLC, part of Springer Nature 2019

Abstract

Purpose of Review There is compelling evidence in the clinical population that long-term weight loss secondary to bariatric surgery is mitigated by the reemergence of maladaptive feeding behaviors and in some cases new onset substance abuse.

Recent Findings A review of the current literature suggests that physical restructuring of the GI tract during WLS alters secretion of feeding peptides and nutrient-sensing mechanisms that directly target the brain's endogenous reward system, the mesolimbic dopamine system.

Summary Post-surgical changes in GI physiology augment activation of the mesolimbic system. In some patients, this process may contribute to a reduced appetite for palatable food whereas in others it may support maladaptive motivated behavior for food and chemical drugs. It is concluded that future studies are required to detail the timing and duration of surgical-induced changes in GI-mesolimbic communication to more fully understand this phenomenon.

Keywords RYGB · VSG · Motivation · Reward · SUD · Dopamine

Introduction

Obesity is a significant unmet health concern that is strongly associated with other comorbidities such as type-2 diabetes, cancer, cardiovascular disease, and hypertension [1–7]. Roughly 6.6% of the population in the USA is considered morbidly obese, classified as having a body mass index of 40 or higher [6]. Presently, we understand obesity as a multitietiological disease with primarily genetic, psychological, and environmental roots [8–13]. A large body of work suggests that exposure to highly palatable food augments key

neuroendocrine signals, which remodel the brain's reward circuitry to reinforce pathological feeding behaviors [14–17]. However, given our limited understanding of the exact etiology of obesity, our ability to treat the disease is quite limited. Presently, weight loss surgery (WLS) is the most effective way to treat morbid obesity [5, 7, 18–25]. The two most common weight loss procedures are Roux-en-Y gastric bypass (RYGB) and vertical sleeve gastrectomy (VSG), with RYGB taking precedence over VSG due to greater effectiveness for long-term, sustained weight loss (Fig. 1) [7, 18, 26–28].

The goal of weight loss surgery is straightforward: restructure the gastrointestinal tract to reduce appetite and metabolic complications, and promote body weight loss [7, 18, 26–28]. An intended consequence of this procedure involves attenuation of food-reinforced behavior, an event that derives from adaptation of brain reward circuits [7, 18, 26–28]. Currently, our understanding of how WLS induces neuroadaptations in the brain's endogenous reward circuit, the mesolimbic dopamine system, is unclear. The goal of this review is to summarize current (2016–present) findings regarding changes in motivation for food and substances of abuse following WLS, as well as the potential mesolimbic and neuroendocrine alterations that underlie these behaviors.

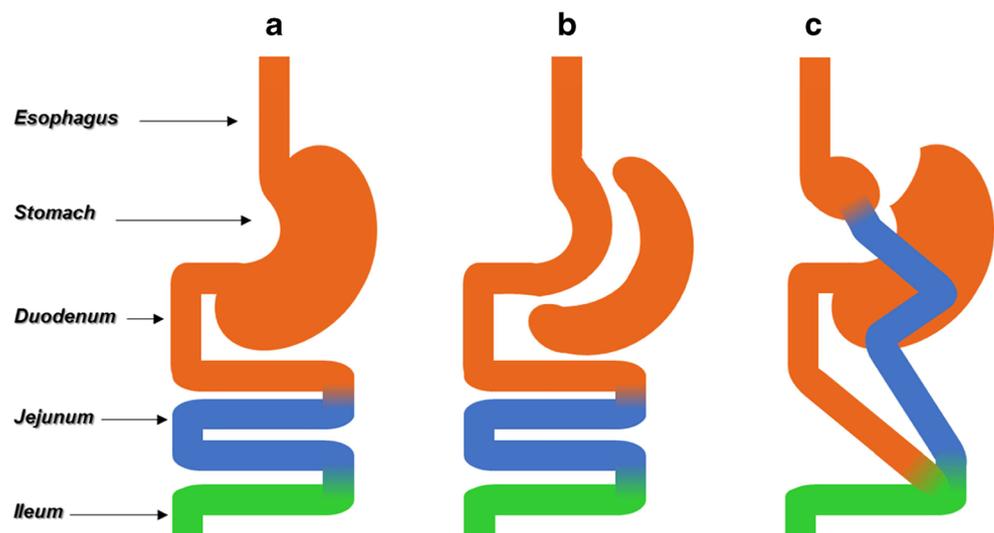
This article is part of the Topical Collection on *Eating Disorders*.

✉ Jon F. Davis
jon.davis@wsu.edu

¹ Department of Integrative Physiology and Neuroscience, College of Veterinary Medicine, Washington State University, 1815 Ferdinand's Lane, Pullman, WA 99164, USA

² Laboratory of Endocrine and Neuropsychiatric Disorders, Division of Basic Pharmaceutical Sciences, College of Pharmacy, Xavier University of Louisiana, New Orleans, LA, USA

Fig. 1 Basic schematic illustration of the gastrointestinal track before (a) and after VSG (b) and RYGB (c) surgical procedures



WLS-Induced Regulation of Food- and Drug-Reinforced Behavior

Following WLS, the reinforcing properties of food, alcohol, and drugs appear to be reset [19–25]. In the clinical population, hedonic feeding behaviors decrease 2 years following surgery, identified as a decreased preference for sweet foods and increased food restriction patterns in patients that report pathological feeding behaviors prior to surgery [19, 29]. However, it has been observed that these behavioral feeding changes are not permanent in some RYGB patients [29–32]. Specifically, after the 2-year mark, there is a greater potential for recidivism of unhealthy feeding in some patients, which often reverses the weight loss effects obtained by the procedure [29–34]. Thus, understanding how food reinforcement mechanisms adapt to RYGB has become a primary focus, with an emphasis on understanding post-surgical changes in sensory detection of food and altered neuroendocrine signaling.

In this context, work from Geary et al. examined increases in sweet preferences for an artificially sweetened diet (ASD) in RYGB female rats [19]. The goals of this study were to determine (1) if the preference for a low-energy ASD changed in female rats following RYGB and (2) how the presence of estrogen modulated dietary changes and weight loss [19]. These questions pose importance because RYGB and other bariatric procedures are most commonly performed on female patients in the clinical population, yet most of the preclinical work has been completed in male rodents [2, 35]. The results from this study suggest that RYGB increases preference and intake of a low-energy ASD relative to an Ensure maintenance diet 6 months following surgery [19]. When access to the ASD is given prior to surgery, RYGB still increases the amount of ASD intake following surgery, suggesting an innate change in taste preference after RYGB [19]. Interestingly,

RYGB rats treated with estrogen (E2) lost the most weight while consuming the most ASD, suggesting the importance of circulating E2 for RYGB success [19]. Notably, exogenous administration of the adipocyte hormone leptin only slightly attenuated the anticipation for ASD in RYGB rats, supporting the notion that hypoleptinemia may contribute to observed differences in diet preference [19]. Collectively, the results establish adaptations in sucrose preference following RYGB and suggest that pre-menopausal women may be better candidates for the surgery compared with women with low circulating estrogen.

Additional work from Mathes and colleagues examined similar changes in dietary preference following RYGB in male rats [20]. Specifically, rats received pre- and post-surgery access to a cafeteria diet paradigm (beans, low-fat/low-sugar; yogurt, low-fat/high-sugar; peanut butter, high-fat/low-sugar; and sugar-fat whip, high-fat/high-sugar; comparable amounts of protein) and changes in macronutrient preference were tracked [20]. RYGB rats decreased fat intake but increased carbohydrate and protein intake. RYGB rats also decreased carbohydrate consumption from sugar, but interestingly, most calories were still obtained from the high-fat/high-sugar option [20]. These studies lend insight into the complexities of macronutrient preference following RYGB. In this regard, new work has established an increase in neuronal activity in response to a high-fat diet (HFD) exposure following RYGB surgery [21]. Interestingly, this change in neuronal activity is region- and time-specific. More specifically, at day 10 post-surgery, but not day 40, eating a voluntary meal induces exaggerated c-Fos expression in the lateral parabrachial nucleus, nucleus tractus solitarius, area postrema, and the central nucleus of the amygdala [21]. These data suggest that brainstem anorexic pathway activation is important in the early stages of RYGB, but the effects may wane as time continues [21]. Future studies that antagonize brain anorectic pathways in

conjunction with assessments of feeding behavior will further elucidate the importance of these signaling mechanisms as regulators of appetite after RYGB.

Separate from the feeding behavior, some RYGB patients are more prone to developing substance abuse disorders (SUD), namely alcohol and opioid abuse [23–25, 36–41]. For example, a body of preclinical work from Davis et al. showed that alcohol use increased following RYGB [36, 37]. In these studies, the authors focused on how the mesolimbic reward pathway adapted to feeding and alcohol intake after RYGB [36, 37]. In recent studies from Sirohi et al., RYGB rats decreased hedonic intake of a highly palatable HFD intake post-surgery, while both SHAM and RYGB rats increased alcohol intake, and preferred lower concentrations of alcohol relative to non-operated pair-fed (PF) controls [23]. In addition, only RYGB rats escalated alcohol intake following acute withdrawal from a liquid alcohol diet that elevated blood alcohol concentration [23].

Similar to increased alcohol intake, roughly 8% of RYGB patients develop chronic opioid use, compared with 3% of the general population [25, 42]. Specifically, recent work from Biegler and colleagues examined intravenous morphine self-administration in RYGB rats using a self-administration task where rats were trained on fixed-ratio lick mode of reinforcement to obtain morphine injection [25]. The results of this study indicate that RYGB rats significantly increased morphine-seeking behavior following training and that RYGB rats consumed significantly more morphine relative to control rats in a time-dependent fashion [25]. Collectively, these results suggest that reinforcement and consumption of morphine increase in RYGB rats. In combination, these studies illustrate the ability of the RYGB procedure to alter preference and reinforcement for palatable food and chemical drugs. Future studies that address the molecular underpinnings of the observed behavioral adaptations will undoubtedly provide more mechanistic insight into this phenomenon. It is worth mentioning that in contrast to RYGB, reduced alcohol has (at least at lower concentrations) been reported following VSG in two recent preclinical studies [24, 43]. In addition to reduced alcohol drinking behavior, hedonic feeding was also decreased and alterations in the ghrelin signaling and/or hypothalamic orexigenic transcripts could mediate these effects [24, 43]. These data suggest that bariatric procedures differentially modulate alcohol intake behavior and particular gut manipulations may carry a reduced risk of developing alcohol use disorders. Studies from Sirohi et al. suggest that reduced mRNA expression of orexigenic peptides including orexin and ghrelin receptor-1a in the hypothalamus may contribute to decreased food intake after VSG [43]; however, further studies are needed to evaluate mechanistic changes that alter alcohol intake.

Presently, the clearest evidence of convergence between clinical and preclinical models of bariatric surgery derives

from behavioral outcomes of alcohol intake after RYGB. For example, RYGB stimulates alcohol intake in both patients [122] and rodents [37,41] highlighting the translational validity of this phenomenon. In terms of weight loss, the effect of RYGB on body weight loss is not universally consistent in patients or rodents. The reasons for failed RYGB in patients are multifactorial and involve psychological processes [127] but not post-surgical changes in gastrointestinal signals that control energy balance [123]. In rodents, weight regain after RYGB can derive from genetic deletion of the satiety hormone leptin [125]. Weight regain has also been observed in 25% of obese rats receiving RYGB. In those studies, increased caloric intake coupled with decreased fecal output was concluded to contribute to body weight regain [124]. When viewed collectively, both clinical and preclinical studies have isolated new onset alcohol intake and body weight regain as negative consequences associated with the RYGB procedure. Points of convergence for these outcomes following VSG, especially alcohol-related endpoints, require more data to discern definite trends.

WLS Induces Functional Changes in CNS Circuits That Regulate Food and Drug Reinforcement

Brain reward circuitry plays a critical role in regulating food and drug reward, and alteration in this circuitry could contribute to pathological food and drug intake [49]. Importantly, disorganization in brain reward circuitry has been documented in obesity which may impair decision making and trigger food/drug-reinforced behavior [50–55]. In this context, bariatric surgeries (e.g., RYGB and VSG) have been shown to normalize obesity-induced alterations in the brain reward regions and overall brain functional connectivity, possibly contributing to beneficial or detrimental effects post-surgery [56–65]. For example, a longitudinal study evaluated behavioral, hormonal, and whole-brain fMRI changes 12 months following VSG and compared these data to pre-surgery baseline in 18 human subjects [60]. This study reported improvement in the maladaptive eating behavior and reduction in the feeding peptides (e.g., ghrelin, leptin, insulin) [60]. In addition, baseline brain activity in response to palatable food anticipation in the nucleus accumbens (NAc) and hypothalamus was significantly predictive of 12-month weight loss. Furthermore, a robust improvement in brain activity in the brain reward regions was observed 12-months post-surgery [60]. Another recent fMRI study reported similar VSG-induced reversal of altered brain activity and improved functional connectivity in brain regions (e.g., orbitofrontal cortex, frontal gyrus) implicated in reward, cognitive, and executive functioning [62]. Interestingly, similar normalization in brain activity has also been documented following RYGB. For example, a prospective fMRI study

evaluated brain activity in response to high-calorie foods before and 6 months after VSG and RYGB and compared with weight-stable controls. A positive correlation between fasting ghrelin and ventral tegmental area (VTA) brain activity was observed in both RYGB and VSG groups. In addition, VTA brain activity in response to high-calorie food significantly declined in the RYGB but not in VSG compared with controls [66]. A bold fMRI study assessed neural inhibitory control (go/no-go task) in response to high- and low-calorie food pictures following RYGB and found increased activation of the prefrontal cortex, cingulate cortex, and right inferior frontal operculum during response inhibition to high caloric food [67]. The same group in another bold fMRI study examined food preference and neural activity in response to food images and order of food image presentation before and 2 months after RYGB in human subjects. It was found that RYGB-induced alterations in the frontoparietal brain region (implication in top-down executive control) correlated with the shift in food preference whereas no such correlation was found regarding plasma ghrelin and endocannabinoid concentrations [62].

In addition to functional activity, obesity has been shown to induce structural brain damage. For example, reduction in gray- and white-matter brain volume which was negatively associated with body fat percentage has been documented in obesity [69]. Zang et al. reported for the first-time recouping of structural neuroplasticity in various brain regions (i.e., orbital frontal cortex, corpus callosum, caudate, insula, inferior frontal gyrus, hippocampus, insula, and external capsule) following laparoscopic VSG [64]. A recent study reported similar structural changes in human subjects 1 month following VSG [65] but no such changes were evident approximately 4 months later [70]. These early structural and functional changes caused by bariatric surgeries could pave the foundation for improved brain connectivity, which may rescue impaired hedonic and cognitive response to palatable food as observed in obesity. Overall, these data suggest that normalization in the dysregulated activity of various brain regions implicated in reward processing and cognitive/executive control following bariatric surgeries may mediate post-surgical changes in the food intake and weight loss.

Separate from structural and functional changes, obesity has been linked to alterations in various neurotransmitter systems in brain reward circuitry which may contribute to impairment in food/drug reward under pathological conditions, and possible alterations in these neurotransmitter systems could mediate effects of bariatric surgery [71–73]. The midbrain reward circuitry which primarily involves excitation of VTA dopaminergic neurons causing dopamine (DA) release in the NAc is a critical structure implicated in mediating food/drug-reinforced behavior [74, 75]. Neuroadaptations in the VTA and striatal dopaminergic neurotransmission have been reported following prolonged exposure to palatable food, diet-

induced obesity, and alcohol intake [76–80]. For example, obese rats show reduced striatal dopamine D2 receptor (D2Rs) density [80]. Similarly, lower D2/3 receptors availability has been documented in human subjects under obese conditions [81]. Recent studies further provide evidence of hypodopaminergic state—associated with obesity [82, 83]. Whether bariatric surgery can reverse this hypodopaminergic state is less clear. Karlsson et al. did not find alterations in the DR receptors availability following 6-months of bariatric surgery [56]. However, another study reported an increase in striatal dopamine (D2/3R) receptors binding following 2 years of RYGB surgery [84]. Moreover, RYGB surgery has been shown to reduce markers of dopaminergic neurotransmission (tyrosine hydroxylase and DAT) transcripts in the VTA [85]. It is important to note that earlier studies assessing D2 receptor availability at shorter time points have reported increased or no changes 6 weeks following RYGB surgery [86, 87]. A relatively recent preclinical study examined D1-, D2-like receptor and dopamine transporter (DAT) binding in diet-induced (9 weeks on diets) obese rats and found reduced D1-like binding in ventrolateral striatum and NAc core and shell, reduced D2-like binding in all striatal areas, and reduced DAT binding dorsomedial striatum which was reversed following RYGB surgery [88••]. These differences across preclinical studies could be attributed to variation in the surgical procedures and/or relative time of observed outcomes [89].

The opioidergic system is another critical and extensively studied brain neurotransmitter system implicated in food/drug reward [90–93]. Interestingly, a significant decrease in mu-opioid receptors (MOR) availability in the brain reward regions has been documented in morbid obesity [94, 95]. Considering the role of MOR in regulating motivation and reward, decreased MOR availability could stimulate increased hedonic feeding in obesity. Since bariatric surgeries reduce hedonic feeding [23, 96], it is interesting to see how bariatric surgeries impact the central opioidergic system. Using positron emission tomography, Karlsson et al. quantified brain D2R and MOR in the 16 women with obesity before and 6 months after weight loss surgery and also compared the data with 14 lean human subjects. While D2R availability was not altered in any brain region in this study, MORs availability was ~23% higher post-surgery in the brain regions also implicated in the reward [56]. Considering the role of MOR in regulating motivation and reward, decreased MOR availability could stimulate increased hedonic feeding in obesity. Therefore, restored MORs signaling nicely aligns with reduced hedonic feeding behavior observed following bariatric surgery [23, 96]. However, a rodent study examining fat preference, MOR availability, and MOR protein levels in the diet-induced obese rats reported a reduction in fat intake, MOR availability, and protein levels in the brain reward circuitry [97]. It is important to note that the MOR availability assessments in this study were carried out 15–17 weeks post-

operatively which is equivalent to ~9 years of human life. Therefore, the observed discrepancy in the clinical and pre-clinical studies could be related to the sampling time following surgery, species differences in brain opioid receptor system, and/or surgical procedures [89, 98].

WLS Alters Feeding Peptides and Nutrient Sensing Mechanisms That Regulate Mesolimbic Function

Feeding peptides produced by the gastrointestinal (GI) tract and nutrient-sensing mechanisms communicate extensively with the central nervous system (CNS) to regulate energy balance [14–17]. A key feature of the RYGB procedure is the restructuring of the GI tract to reroute the small intestines to a volume-reduced stomach [6, 7]. Importantly, this process alters the secretion of feeding peptides that target the CNS to regulate mesolimbic dopamine release [99]. This adaptation in GI-CNS communication has been hypothesized to underlie the dramatic changes in feeding- and alcohol-reinforced behaviors that accompany the RYGB procedure [100]. Perhaps the most dramatic example of GI adaptation involves RYGB-induced increases in glucagon-like peptide-1 (GLP-1) secretion. GLP-1 is produced in the small intestines and once released functions to lower blood glucose and reduce food intake [101]. Following RYGB, plasma GLP-1 levels are significantly increased in response to a mixed meal in patients [102] and rodents [103]. In addition, GLP-1 secretion is stimulated by alcohol ingestion in rats [36]. The GLP-1 receptor is expressed in the VTA and NAc key mesolimbic regions that regulate reinforced behaviors [104]. A central application of GLP-1 agonists reduces psychostimulant-induced phasic dopamine release in the NAc [105]. In addition, GLP-1 application in the VTA decreases psychostimulant self-administration in rodents [106]. In combination, these examples suggest that excursions in GLP-1 secretion following RYGB may target the mesolimbic system directly to direct food and drug intake. To our knowledge, data supporting the direct involvement of mesolimbic GLP-1 signaling after RYGB has not been reported and remains an interesting area of inquiry.

Separate from GLP-1, RYGB also dramatically regulates GI ghrelin secretion. Ghrelin is a 28 amino acid protein secreted primarily by the stomach that acts at the ghrelin-1a receptor (GHSR-1a) in the CNS to control feeding behavior [14]. Multiple observations in patients [107] and rodents [23, 37] suggest that the RYGB procedure results in decreased circulating ghrelin levels, a process that is seemingly dynamic. For example, rodent studies indicate that ghrelin production and secretion is elevated 30 days after surgery but that this effect wanes at more distal time points leading to an overall decrease in plasma ghrelin at 90 days after surgery [37]. This becomes important when considering that GHSR-1a surface expression

is decreased in conditions of elevated plasma ghrelin [108]. Therefore, acute surges in ghrelin after surgery could have long-lasting impacts on GHSR-1a function. GHSR-1a is expressed in VTA neurons, where it acts to stimulate dopamine release and food- and alcohol-reinforced behaviors [109–112]. Recent work from Sirohi et al. was one of the first studies to link changes in feeding peptides with mesolimbic function and behavior following RYGB [23]. In this study, the authors discovered that GHSR-1a activity controls tonic dopamine release from VTA neurons in male Long Evans rats [23••]. The group further reported that the ability of GHSR-1a antagonism to reduce tonic dopamine was ineffective in RYGB rats that consumed more alcohol relative to controls [23••]. It is important to note here that GHSR-1a displays constitutive activity without the ghrelin ligand present [113] and that this process is functional to control feeding behavior [114]. Thus, one interpretation of the Sirohi data is that GHSR-1a activity in the VTA is increased after RYGB and that this process serves to promote excess alcohol consumption after surgery. It is currently unclear if plasma levels of ghrelin contribute to altered GHSR-1a control of VTA dopamine firing or if a separate process accounts for this observation. Moreover, additional studies that determine the timing and nature of altered GHSR-1a activity in the VTA are required to determine if this phenomenon contributes to adaptations in food and alcohol intake after RYGB.

In terms of nutrient sensing, a recent study indicates that altered lipid sensing may play a significant role in adaptations in mesolimbic function and behavior following RYGB. Oleoylethanolamide (OEA), a lipid species produced in the GI tract, binds the proliferator-associated receptor- α (PPAR- α) to exert biological action [115]. Notably, it has been previously demonstrated that OEA-PPAR- α signaling regulates striatal dopamine release in obese rodents [116]. In a recent study, Hankir et al. reported that OEA is upregulated following RYGB in diet-induced obese rats [117]. In addition, the authors discovered that striatal dopamine levels following an oral meal were increased following RYGB relative to obese sham controls and that this response required intact vagal afferent signaling. Additionally, expression of striatal dopamine-1 receptor (D1R) levels was increased by RYGB in this study. It is well accepted that RYGB shifts preference from high- to low-fat diets in patients [99, 100, 118, 119] and rodent models [20, 23, 119]. A particularly striking discovery from Hankir et al. was the observation that OEA-PPAR- α signaling regulated reductions high-fat preference following RYGB [117••]. Using a series of behavioral pharmacologic manipulations, the authors discovered that attenuation of OEA-PPAR- α signaling in the GI tract or D1R signaling in the dorsal striatum attenuated the decrease in high-fat diet preference in rats receiving the RYGB procedure [117]. The combined results of this study were the first to indicate that adaptations in lipid sensing are transmitted to the mesolimbic

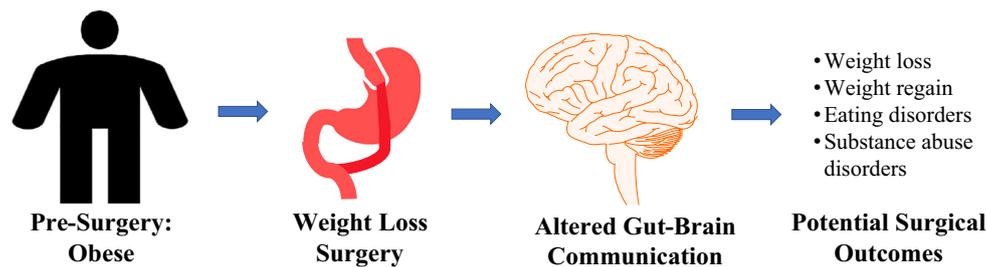


Fig. 2 Proposed adaptation following WLS. WLS is completed on morbidly obese patients facing life-threatening health complications. However, WLS functionally alters the ability of the GI tract to secrete key neuroendocrine hormones, altering neuronal activity in the mesolimbic dopamine system. Recent research suggests that, in some instances, this physiological adaptation rewires the brain to promote maladaptive behaviors such as seeking substances of abuse, including

dopamine system to control feeding behavior and that OEA-PPAR- α signaling in particular is a positive adaptation that regulates shifts in diet preference to low-fat foods following RYGB.

Efforts to design therapeutic options that target gastrointestinal peptide signaling to treat obesity are an active area of investigation. As new treatments emerge, it will be important to determine if these drugs can enhance post-surgical outcomes following bariatric interventions. A great example of this concept is found in preclinical studies that examined the blood glucose-lowering response of RYGB surgery. Habegger and colleagues were able to demonstrate that obese rats sensitive to the glucose-lowering effects of GLP-1 experienced greater reductions in blood glucose following the RYGB procedure [126]. This is one example of many that leverage knowledge of gut-brain communication to refine surgical interventions aimed at reducing metabolic complications. It is our contention that future approaches of this type will greatly refine the efficacy of bariatric surgery on brain and behavioral function.

Conclusions

In conclusion, it is apparent that restructuring the nature of WLS is capable of altering how the GI tract communicates with mesolimbic circuitry and that this process participates in the control of appetitive and consummatory aspects of ingestive behavior. A significant portion of the recent literature highlights neuroendocrine signals, such as GLP-1, ghrelin, and OEA, as potential signaling mechanisms that facilitate GI communication with the mesolimbic dopamine system. The nature of these modifications for patient success is unclear. In this regard, adaptations within the mesolimbic circuit may aid in reduced appetite and/or bodyweight loss. However, these changes could also provoke harmful behavior such as new-onset alcohol intake or opioid use (Fig. 2). Understanding how the brain adapts to WLS continues to be

chemical drugs, alcohol, and food. These maladaptive changes may contribute to the maintenance of long-term body weight after WLS surgery. We propose that the surgical restructuring of the gut alters the brain, resulting in behavioral changes that may be of detriment to clinical populations; thus, further preclinical and clinical research is needed to understand these phenomena

an important area of inquiry for obesity researchers. In this context, an unexplored hypothesis in the field involves understanding if new set points for appetite and body weight contribute to weight regain and/or substance disorders. Future preclinical research that capitalizes on techniques that enable defined neurocircuit manipulations paired with in vivo imaging will fill critical gaps in knowledge regarding how the brain adapts to a restructured GI tract.

Compliance with Ethical Standards

Conflict of Interest Julianna N. Brutman, Sunil Sirohi, and Jon F. Davis each declare no potential conflicts of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

References

Papers of particular interest, published recently, have been highlighted as: •• Of major importance

1. Adult Obesity Facts | Overweight & Obesity | CDC. (2019, January 31). Retrieved May 29, 2019, from <https://www.cdc.gov/obesity/data/adult.html>
2. Flegal KM, Carroll MD, Kit BK, Ogden CL. Prevalence of obesity and trends in the distribution of body mass index among US adults, 1999-2010. *JAMA*. 2012;307(5):491–7. <https://doi.org/10.1001/jama.2012.39>.
3. Arterburn DE, Maciejewski ML, Tsevat J. Impact of morbid obesity on medical expenditures in adults. *Int J Obes*. 2005;29(3):334–9. <https://doi.org/10.1038/sj.ijo.0802896>.
4. Drenick EJ, Bale GS, Seltzer F, Johnson DG. Excessive mortality and causes of death in morbidly obese men. *JAMA*. 1980;243(5):443–5. <https://doi.org/10.1001/jama.1980.03300310031018>.
5. Lutz TA, Bueter M. The physiology underlying Roux-en-Y gastric bypass: a status report. *Am J Physiol Regul Integr Comp Physiol*. 2014;307(11):R1275–91. <https://doi.org/10.1152/ajpregu.00185.2014>.

6. Sturm R, Hattori A. Morbid obesity rates continue to rise rapidly in the US. *Int J Obes* (2005). 2013;37(6):889–91. <https://doi.org/10.1038/ijo.2012.159>.
7. Wang Y, Song Y, Chen J, Zhao R, Xia L, Cui Y, et al. Roux-en-Y gastric bypass versus sleeve gastrectomy for super super obese and super obese: systematic review and meta-analysis of weight results, comorbidity resolution. *Obes Surg*. 2019;29(6):1954–64. <https://doi.org/10.1007/s11695-019-03817-4>.
8. Bouret S, Levin BE, Ozanne SE. Gene-environment interactions controlling energy and glucose homeostasis and the developmental origins of obesity. *Physiol Rev*. 2015;95(1):47–82. <https://doi.org/10.1152/physrev.00007.2014>.
9. Casazza K, Brown A, Astrup A, Bertz F, Baum C, Brown MB, et al. Weighing the evidence of common beliefs in obesity research. *Crit Rev Food Sci Nutr*. 2015;55(14):2014–53. <https://doi.org/10.1080/10408398.2014.922044>.
10. Cooksey-Stowers K, Schwartz MB, Brownell KD. Food swamps predict obesity rates better than food deserts in the United States. *Int J Environ Res Public Health*. 2017;14(11). <https://doi.org/10.3390/ijerph14111366>.
11. Godfrey KM, Reynolds RM, Prescott SL, Nyirenda M, Jaddoe VWV, Eriksson JG, et al. Influence of maternal obesity on the long-term health of offspring. *Lancet Diabetes Endocrinol*. 2017;5(1):53–64. [https://doi.org/10.1016/S2213-8587\(16\)30107-3](https://doi.org/10.1016/S2213-8587(16)30107-3).
12. Hruby A, Hu FB. The epidemiology of obesity: A big picture. *Pharmacoeconomics*. 2015;33(7):673–89. <https://doi.org/10.1007/s40273-014-0243-x>.
13. Kim TJ, von dem Knesebeck O. Income and obesity: what is the direction of the relationship? A systematic review and meta-analysis. *BMJ Open*. 2018;8(1). <https://doi.org/10.1136/bmjopen-2017-019862>.
14. Müller TD, Nogueiras R, Andermann ML, Andrews ZB, Anker SD, Argente J, et al. Ghrelin. *Mol Metabolism*. 2015;4(6):437–60. <https://doi.org/10.1016/j.molmet.2015.03.005>.
15. Bliss ES, Whiteside E. The gut-brain axis, the human gut microbiota and their integration in the development of obesity. *Front Physiol*. 2018;9. <https://doi.org/10.3389/fphys.2018.00900>.
16. de Lartigue G, Barbier de la Serre C, Espero E, Lee J, Raybould HE. Diet-induced obesity leads to the development of leptin resistance in vagal afferent neurons. *Am J Physiol Endocrinol Metab*. 2011;301(1):E187–95. <https://doi.org/10.1152/ajpendo.00056.2011>.
17. Sanmiguel C, Gupta A, Mayer EA. Gut microbiome and obesity: a plausible explanation for obesity. *Curr Obes Rep*. 2015;4(2):250–61. <https://doi.org/10.1007/s13679-015-0152-0>.
18. Kang JH, Le QA. Effectiveness of bariatric surgical procedures. *Medicine*. 2017;96(46):e8632. <https://doi.org/10.1097/MD.0000000000008632>.
19. Geary N, Bächler T, Whiting L, Lutz TA, Asarian L. RYGB progressively increases avidity for a low-energy, artificially sweetened diet in female rats. *Appetite*. 2016;98:133–41. <https://doi.org/10.1016/j.appet.2015.11.029>.
20. Mathes CM, Letourneau C, Blonde GD, le Roux CW, Spector AC. Roux-en-Y gastric bypass in rats progressively decreases the proportion of fat calories selected from a palatable cafeteria diet. *Am J Physiol Regul Integr Comp Physiol*. 2016;310(10):R952–9. <https://doi.org/10.1152/ajpregu.00444.2015>.
21. Mumphrey MB, Hao Z, Townsend RL, Patterson LM, Münzberg H, Morrison CD, et al. Eating in mice with gastric bypass surgery causes exaggerated activation of brainstem anorexia circuit. *Int J Obes* (2005). 2016;40(6):921–8. <https://doi.org/10.1038/ijo.2016.38>.
22. Washington MC, Mhalhal TR, Johnson-Rouse T, Berger J, Heath J, Seeley R, et al. Roux-en-Y gastric bypass augments the feeding responses evoked by gastrin-releasing peptides. *J Surg Res*. 2016;206(2):517–24. <https://doi.org/10.1016/j.jss.2016.08.057>.
23. Sirohi S, Richardson BD, Lugo JM, Rossi DJ, Davis JF. Impact of Roux-en-Y gastric bypass surgery on appetite, alcohol intake behaviors, and midbrain ghrelin signaling in the rat. *Obesity*. 2017;25(7):1228–36. <https://doi.org/10.1002/oby.21839>. **Discovered that GHSR-1a signaling is altered in mesolimbic dopamine neurons in rats behaviorally characterized for increased alcohol intake and reduced hedonic food intake.**
24. Orellana ER, Jamis C, Horvath N, Hajnal A. Effect of vertical sleeve gastrectomy on alcohol consumption and preferences in dietary obese rats and mice: a plausible role for altered ghrelin signaling. *Brain Res Bull*. 2018;138:26–36. <https://doi.org/10.1016/j.brainresbull.2017.08.004>.
25. Biegler JM, Freet CS, Horvath N, Rogers AM, Hajnal A. Increased intravenous morphine self-administration following Roux-en-Y gastric bypass in dietary obese rats. *Brain Res Bull*. 2016;123:47–52. <https://doi.org/10.1016/j.brainresbull.2015.08.003>.
26. Pories WJ. Bariatric surgery: risks and rewards. *J Clin Endocrinol Metab*. 2008;93(11 Suppl 1):S89–96. <https://doi.org/10.1210/jc.2008-1641>.
27. Stefater MA, Wilson-Pérez HE, Chambers AP, Sandoval DA, Seeley RJ. All bariatric surgeries are not created equal: insights from mechanistic comparisons. *Endocr Rev*. 2012;33(4):595–622. <https://doi.org/10.1210/er.2011-1044>.
28. Wolfe BM, Kvach E, Eckel RH. Treatment of obesity: weight loss and bariatric surgery. *Circ Res*. 2016;118(11):1844–55. <https://doi.org/10.1161/CIRCRESAHA.116.307591>.
29. Pepino MY, Bradley D, Eagon JC, Sullivan S, Abumrad NA, Klein S. Changes in taste perception and eating behavior after bariatric surgery-induced weight loss in women. *Obesity (Silver Spring, Md.)*. 2014;22(5):E13–20. <https://doi.org/10.1002/oby.20649>.
30. Magro DO, Geloneze B, Delfini R, Pareja BC, Callejas F, Pareja JC. Long-term weight regain after gastric bypass: a 5-year prospective study. *Obes Surg*. 2008;18(6):648–51. <https://doi.org/10.1007/s11695-007-9265-1>.
31. Karmali S, Brar B, Shi X, Sharma AM, de Gara C, Birch DW. Weight recidivism post-bariatric surgery: a systematic review. *Obes Surg*. 2013;23(11):1922–33. <https://doi.org/10.1007/s11695-013-1070-4>.
32. Still CD, Wood GC, Chu X, Manney C, Strodel W, Petrick A, et al. Clinical factors associated with weight loss outcomes after Roux-en-Y gastric bypass surgery. *Obesity (Silver Spring, Md.)*. 2014;22(3):888–94. <https://doi.org/10.1002/oby.20529>.
33. Ullrich J, Ernst B, Wilms B, Thurnheer M, Schultes B. Roux-en-Y gastric bypass surgery reduces hedonic hunger and improves dietary habits in severely obese subjects. *Obes Surg*. 2013;23(1):50–5. <https://doi.org/10.1007/s11695-012-0754-5>.
34. Brethauer SA, Aminian A, Romero-Talamás H, Batayyah E, Mackey J, Kennedy L, et al. Can diabetes be surgically cured? Long-term metabolic effects of bariatric surgery in obese patients with type 2 diabetes mellitus. *Ann Surg*. 2013;258(4):628–36; discussion 636–637. <https://doi.org/10.1097/SLA.0b013e3182a5034b>.
35. DeMaria EJ, Pate V, Warthen M, Winegar DA. Baseline data from American Society for Metabolic and Bariatric Surgery-designated Bariatric Surgery Centers of Excellence using the Bariatric Outcomes Longitudinal Database. *Surg Obes Relat Dis Off J Am Soc Bariatric Surg*. 2010;6(4):347–55. <https://doi.org/10.1016/j.soard.2009.11.015>.
36. Davis JF, Schurdak JD, Magrisso IJ, Mul JD, Grayson BE, Pfluger PT, et al. Gastric bypass surgery attenuates ethanol consumption in ethanol-preferring rats. *Biol Psychiatry*. 2012;72:354–60. <https://doi.org/10.1016/j.biopsych.2012.01.035>.

37. Davis JF, Tracy AL, Schurdak JD, Magrisso IJ, Grayson BE, Seeley RJ, et al. Roux en Y gastric bypass increases ethanol intake in the rat. *Obes Surg*. 2013;23(7):920–30. <https://doi.org/10.1007/s11695-013-0884-4>.
38. Dutta S, Morton J, Shepard E, Peebles R, Farrales-Nguyen S, Hammer LD, et al. Methamphetamine use following bariatric surgery in an adolescent. *Obes Surg*. 2006;16(6):780–2. <https://doi.org/10.1381/09608920677346646>.
39. Ertelt TW, Mitchell JE, Lancaster K, Crosby RD, Steffen KJ, Marino JM. Alcohol abuse and dependence before and after bariatric surgery: a review of the literature and report of a new data set. *Surg Obes Relat Dis*. 2008;4(5):647–50. <https://doi.org/10.1016/j.soard.2008.01.004>.
40. Hajnal A, Zharikov A, Polston JE, Fields MR, Tomasko J, Rogers AM, et al. Alcohol reward is increased after Roux-en-Y gastric bypass in dietary obese rats with differential effects following ghrelin antagonism. *PLoS One*. 2012;7(11):e49121. <https://doi.org/10.1371/journal.pone.0049121>.
41. Thanos PK, Subrizi M, Delis F, Cooney RN, Culnan D, Sun M, et al. Gastric bypass increases ethanol and water consumption in diet-induced obese rats. *Obes Surg*. 2012;22(12):1884–92. <https://doi.org/10.1007/s11695-012-0749-2>.
42. Boudreau D, Von Korff M, Rutter CM, Saunders K, Ray GT, Sullivan MD, et al. Trends in long-term opioid therapy for chronic non-cancer pain. *Pharmacoepidemiol Drug Saf*. 2009;18(12):1166–75. <https://doi.org/10.1002/pds.1833>.
43. Sirohi S, Skripnikova E, Davis JF. Vertical sleeve gastrectomy attenuates hedonic feeding without impacting alcohol drinking in rats. *Obesity (Silver Spring, Md.)*. 2019;27(4):603–11. <https://doi.org/10.1002/oby.22415>.
44. King WC, Chen J-Y, Mitchell JE, Kalarchian M, Steffen KJ, Engel SG, et al. Prevalence of alcohol use disorders before and after bariatric surgery. *JAMA*. 2012;307:2516–25. <https://doi.org/10.1001/jama.2012.6147>.
45. Himes SM, Grothe KB, Clark MM, Swain JM, Collazo-Clavell ML, Sarr MG. Stop regain: a pilot psychological intervention for bariatric patients experiencing weight regain. *Obes Surg*. 2015;25(5):922–7. <https://doi.org/10.1007/s11695-015-1611-0>.
46. Tamboli RA, Breitman I, Marks-Shulman PA, Jabbour K, Melvin W, Williams B, et al. Early weight regain after gastric bypass does not affect insulin sensitivity but is associated with elevated ghrelin. *Obesity*. 2014;22(7):1617–22. <https://doi.org/10.1002/oby.2077>.
47. Hao Z, Münzberg H, Rezai-Zadeh K, Keenan M, Coulon D, Lu H, et al. Leptin deficient ob/ob mice and diet-induced obese mice responded differently to Roux-en-Y bypass surgery. *Int J Obes*. 2015;39(5):798–805. <https://doi.org/10.1038/ijo.2014.189>.
48. Guijarro A, Suzuki S, Chen C, Kirchner H, Middleton FA, Nadtochiy S, et al. Characterization of weight loss and weight regain mechanisms after Roux-en-Y gastric bypass in rats. *Am J Phys Regul Integr Comp Phys*. 2007;293(4):R1474–89. <https://doi.org/10.1152/ajpregu.00171.2007>.
49. Volkow ND, Baler RD. NOW vs LATER brain circuits: implications for obesity and addiction. *Trends Neurosci*. 2015;38:345–52.
50. Marqués-Iturria I, Scholtens LH, Garolera M, Pueyo R, García-García I, González-Tartiere P, et al. Affected connectivity organization of the reward system structure in obesity. *Neuroimage*. 2015;111:100–6.
51. Tuominen L, Tuulari J, Karlsson H, Hirvonen J, Helin S, Salminen P, et al. Aberrant mesolimbic dopamine-opiate interaction in obesity. *Neuroimage*. 2015;122:80–6.
52. Geha P, Cecchi G, Todd Constable R, Abdallah C, Small DM. Reorganization of brain connectivity in obesity. *Hum Brain Mapp*. 2017;38:1403–20.
53. Avery JA, Powell JN, Breslin FJ, Lepping RJ, Martin LE, Patrician TM, et al. Obesity is associated with altered mid-insula functional connectivity to limbic regions underlying appetitive responses to foods. *J Psychopharmacol (Oxford)*. 2017;31:1475–84.
54. Ho M-C, Chen VC-H, Chao S-H, Fang C-T, Liu Y-C, Weng J-C. Neural correlates of executive functions in patients with obesity. *PeerJ*. 2018;6:e5002.
55. Chen VC-H, Liu Y-C, Chao S-H, McIntyre RS, Cha DS, Lee Y, et al. Brain structural networks and connectomes: the brain-obesity interface and its impact on mental health. *Neuropsychiatr Dis Treat*. 2018;14:3199–208.
56. Karlsson HK, Tuulari JJ, Tuominen L, Hirvonen J, Honka H, Parkkola R, et al. Weight loss after bariatric surgery normalizes brain opioid receptors in morbid obesity. *Mol Psychiatry*. 2016;21:1057–62.
57. Thanos PK, Michaelides M, Subrizi M, Miller ML, Bellezza R, Cooney RN, et al. Roux-en-Y gastric bypass alters brain activity in regions that underlie reward and taste perception. *PLoS One*. 2015;10:e0125570.
58. Wiemerslage L, Zhou W, Olivo G, Stark J, Hogenkamp PS, Larsson EM, et al. A resting-state fMRI study of obese females between pre- and postprandial states before and after bariatric surgery. *Eur J Neurosci*. 2017;45:333–41.
59. Olivo G, Zhou W, Sundbom M, Zhukovsky C, Hogenkamp P, Nikontovic L, et al. Resting-state brain connectivity changes in obese women after Roux-en-Y gastric bypass surgery: a longitudinal study. *Sci Rep*. 2017;7:6616.
60. Holsen LM, Davidson P, Cerit H, Hye T, Moondra P, Haimovici F, et al. Neural predictors of 12-month weight loss outcomes following bariatric surgery. *Int J Obes*. 2018;42:785–93.
61. Pearce AL, Mackey E, Cherry JBC, Olson A, You X, Magge SN, et al. Effect of adolescent bariatric surgery on the brain and cognition: a pilot study. *Obesity (Silver Spring)*. 2017;25:1852–60.
62. Li P, Shan H, Liang S, et al. Sleeve gastrectomy recovering disordered brain function in subjects with obesity: a longitudinal fMRI study. *Obes Surg*. 2018;28:2421–8.
63. Han W, Tellez LA, Niu J, Medina S, Ferreira TL, Zhang X, et al. Striatal dopamine links gastrointestinal rerouting to altered sweet appetite. *Cell Metab*. 2016;23:103–12.
64. Zhang Y, Ji G, Xu M, Cai W, Zhu Q, Qian L, et al. Recovery of brain structural abnormalities in morbidly obese patients after bariatric surgery. *Int J Obes*. 2016;40:1558–65.
65. Liu L, Ji G, Li G, et al. Structural changes in brain regions involved in executive-control and self-referential processing after sleeve gastrectomy in obese patients. *Brain Imaging Behav*. 2018.
66. Faulconbridge LF, Ruparel K, Loughead J, Allison KC, Hesson LA, Fabricatore AN, et al. Changes in neural responsivity to highly palatable foods following Roux-en-Y gastric bypass, sleeve gastrectomy, or weight stability: an fMRI study. *Obesity (Silver Spring)*. 2016;24:1054–60.
67. Zoon HFA, de Bruijn SEM, Jager G, Smeets PAM, de Graaf C, Janssen IMC, et al. Altered neural inhibition responses to food cues after Roux-en-Y gastric bypass. *Biol Psychol*. 2018;137:34–41.
68. Zoon HFA, de Bruijn SEM, Smeets PAM, de Graaf C, Janssen IMC, Schijns W, et al. Altered neural responsivity to food cues in relation to food preferences, but not appetite-related hormone concentrations after RYGB-surgery. *Behav Brain Res*. 2018;353:194–202.
69. Karlsson HK, Tuulari JJ, Hirvonen J, Lepomäki V, Parkkola R, Hiltunen J, et al. Obesity is associated with white matter atrophy: a combined diffusion tensor imaging and voxel-based morphometric study. *Obesity*. 2013;21:2530–7.
70. Bohon C, Geliebter A. Change in brain volume and cortical thickness after behavioral and surgical weight loss intervention. *Neuroimage Clin*. 2019;21:101640.

71. Volkow ND, Wang GJ, Fowler JS, Tomasi D, Baler R. Food and drug reward: overlapping circuits in human obesity and addiction. *Curr Top Behav Neurosci*. 2012;11:1–24.
72. Volkow ND, Wang G-J, Tomasi D, Baler RD. Obesity and addiction: neurobiological overlaps. *Obes Rev*. 2013;14:2–18.
73. Hankir MK, Seyfried F, Hintschich CA, Diep TA, Kleberg K, Kranz M, et al. Gastric bypass surgery recruits a gut PPAR- α -striatal D1R pathway to reduce fat appetite in obese rats. *Cell Metab*. 2017;25:335–44.
74. Wise RA. Role of brain dopamine in food reward and reinforcement. *Philos Trans R Soc Lond Ser B Biol Sci*. 2006;361:1149–58.
75. Rodd ZA, Melendez RI, Bell RL, Kuc KA, Zhang Y, Murphy JM, et al. Intracranial self-administration of ethanol within the ventral tegmental area of male Wistar rats: evidence for involvement of dopamine neurons. *JNeurosci*. 2004;24:1050–7.
76. Liu S, Globa AK, Mills F, Naef L, Qiao M, Bamji SX, et al. Consumption of palatable food primes food approach behavior by rapidly increasing synaptic density in the VTA. *Proc Natl Acad Sci U S A*. 2016;113:2520–5.
77. Diana M, Pistis M, Carboni S, Gessa GL, Rossetti ZL. Profound decrement of mesolimbic dopaminergic neuronal activity during ethanol withdrawal syndrome in rats: electrophysiological and biochemical evidence. *Proc Natl Acad Sci U S A*. 1993;90:7966–9.
78. Cook JB, Hendrickson LM, Garwood GM, Toungate KM, Nania CV, Morikawa H. Junk food diet-induced obesity increases D2 receptor autoinhibition in the ventral tegmental area and reduces ethanol drinking. *PLoS One*. 2017;12:e0183685.
79. Koyama S, Mori M, Kanamaru S, Sazawa T, Miyazaki A, Terai H, et al. Obesity attenuates D2 autoreceptor-mediated inhibition of putative ventral tegmental area dopaminergic neurons. *Physiol Rep*. 2014;2:e12004.
80. Johnson PM, Kenny PJ. Dopamine D2 receptors in addiction-like reward dysfunction and compulsive eating in obese rats. *Nat Neurosci*. 2010;13:635–41.
81. de Weijer BA, van de Giessen E, van Amelsvoort TA, Boot E, Braak B, Janssen IM, et al. Lower striatal dopamine D2/3 receptor availability in obese compared with non-obese subjects. *EJNMMI Res*. 2011;1:37.
82. Wu C, Garamszegi SP, Xie X, Mash DC. Altered dopamine synaptic markers in postmortem brain of obese subjects. *Front Hum Neurosci*. 2017;11:386.
83. Pak K, Kim S-J, Kim IJ. Obesity and brain positron emission tomography. *Nucl Med Mol Imaging*. 2018;52:16–23.
84. van der Zwaal EM, de Weijer BA, van de Giessen EM, Janssen I, Berends FJ, van de Laar A, et al. Striatal dopamine D2/3 receptor availability increases after long-term bariatric surgery-induced weight loss. *Eur Neuropsychopharmacol*. 2016;26:1190–200.
85. Blum K, Thanos PK, Wang G-J, Febo M, Demetrovics Z, Modestino EJ, et al. The food and drug addiction epidemic: targeting dopamine homeostasis. *Curr Pharm Des*. 2018;23:6050–61.
86. de Weijer BA, van de Giessen E, Janssen I, Berends FJ, van de Laar A, Ackermans MT, et al. Striatal dopamine receptor binding in morbidly obese women before and after gastric bypass surgery and its relationship with insulin sensitivity. *Diabetologia*. 2014;57:1078–80.
87. Steele KE, Prokopowicz GP, Schweitzer MA, Magunson TH, Lidor AO, Kuwabawa H, et al. Alterations of central dopamine receptors before and after gastric bypass surgery. *Obes Surg*. 2010;20:369–74.
88. Hamilton J, Swenson S, Hajnal A, Thanos PK. Roux-en-Y gastric bypass surgery normalizes dopamine D1, D2, and DAT levels. *Synapse*. 2018;72:e22058. **Found restoration of D2-receptor binding in striatum and hence mesolimbic dopamine function in RYGB rats relative to obese controls.**
89. Doumouras AG, Saleh F, Anvari S, Gmora S, Anvari M, Hong D. Mastery in bariatric surgery: the long-term surgeon learning curve of Roux-en-Y gastric bypass. *Ann Surg*. 2018;267:489–94.
90. Merrer JL, Becker JAJ, Befort K, Kieffer BL. Reward processing by the opioid system in the brain. *Physiol Rev*. 2009;89:1379–412.
91. Hyttia P. Involvement of mu-opioid receptors in alcohol drinking by alcohol-preferring AA rats. *Pharmacol Biochem Behav*. 1993;45:697–701.
92. Bazov I, Kononenko O, Watanabe H, et al. The endogenous opioid system in human alcoholics: molecular adaptations in brain areas involved in cognitive control of addiction. *AddictBiol*. 2011.
93. Nogueiras R, Romero-Picó A, Vazquez MJ, Novelle MG, López M, Diéguez C. The opioid system and food intake: homeostatic and hedonic mechanisms. *Obes Facts*. 2012;5:196–207.
94. Joutsa J, Karlsson HK, Majuri J, et al. Binge eating disorder and morbid obesity are associated with lowered mu-opioid receptor availability in the brain. *Psychiatry Res Neuroimaging*. 2018;276:41–5.
95. Karlsson HK, Tuominen L, Tuulari JJ, Hirvonen J, Parkkola R, Helin S, et al. Obesity is associated with decreased μ -opioid but unaltered dopamine D2 receptor availability in the brain. *J Neurosci*. 2015;35:3959–65.
96. Sirohi S, Skripnikova E, Davis JF. Vertical sleeve gastrectomy attenuates hedonic feeding without impacting alcohol drinking in rats. *Obesity (Silver Spring)*. 2019;27:603–11.
97. Hankir MK, Patt M, Patt JTW, Becker GA, Rullmann M, Kranz M, et al. Suppressed fat appetite after Roux-en-Y gastric bypass surgery associates with reduced brain μ -opioid receptor availability in diet-induced obese male rats. *Front Neurosci*. 2017;10.
98. Pasternak GW, Pan Y-X. Mu opioids and their receptors: evolution of a concept. *Pharmacol Rev*. 2013;65:1257–317.
99. le Roux CW, Welbourn R, Werling M, Osborne A, Kokkinos A, Laurenus A, et al. Gut hormones as mediators of appetite and weight loss after Roux-en-Y gastric bypass. *Ann Surg*. 2007;246(5):780–5. <https://doi.org/10.1097/SLA.0b013e3180caa3e3>.
100. Ivezaj V, Stoeckel LE, Avena NM, Benoit SC, Conason A, Davis JF, et al. Obesity and addiction: can a complication of surgery help us understand the connection? *Obes Rev*. 2017;18:765–75. <https://doi.org/10.1111/obr.12542>.
101. Drucker DJ. The biology of incretin hormones. *Cell Metab*. 2006;3:153–65. Retrieved from. <https://doi.org/10.1016/j.cmet.2006.01.004n>.
102. Dar MS, Chapman WH, Pender JR, Drake AJ, O'Brien K, Tanenberg RJ, et al. GLP-1 response to a mixed meal: what happens 10 years after Roux-en-Y gastric bypass (RYGB)? *Obes Surg*. 2012;22(7):1077–83. <https://doi.org/10.1007/s11695-012-0624-1>.
103. Chambers AP, Jessen L, Ryan KK, Sisley S, Wilsonpérez HE, Stefater MA, et al. Weight-independent changes in blood glucose homeostasis after gastric bypass or vertical sleeve gastrectomy in rats. *Gastroenterology*. 2011;141(3):950–8. <https://doi.org/10.1053/j.gastro.2011.05.050>.
104. Alhadeff AL, Rupprecht LE, Hayes MR. GLP-1 neurons in the nucleus of the solitary tract project directly to the ventral tegmental area and nucleus accumbens to control for food intake. *Endocrinology*. 2012;153(2):647–58. <https://doi.org/10.1210/en.2011-1443>.
105. Fortin SM, Roitman MF. Central GLP-1 receptor activation modulates cocaine-evoked phasic dopamine signaling in the nucleus accumbens core. *Physiol Behav*. 2017;176:17–25. <https://doi.org/10.1016/j.physbeh.2017.03.019>.

106. Schmidt HD, Mietlicki-Baase EG, Ige KY, Maurer JJ, Reiner DJ, Zimmer DJ, et al. Glucagon-like peptide-1 receptor activation in the ventral tegmental area decreases the reinforcing efficacy of cocaine. *Neuropsychopharmacology*. 2016;41(7):1917–28. <https://doi.org/10.1038/npp.2015.362>.
107. Cummings DE, Weigle DS, Frayo RS, Breen P, Ma MK, Dellinger EP, et al. Plasma ghrelin levels after diet-induced weight loss or gastric bypass surgery. *N Engl J Med*. 2002;346(21):1623–30. <https://doi.org/10.1056/NEJMoa012908>.
108. Camiña JP, Carreira MC, El Messari S, Llorens-Cortes C, Smith RG, Casanueva FF. Desensitization and endocytosis mechanisms of ghrelin-activated growth hormone secretagogue receptor 1a. *Endocrinology*. 2004;145(2):930–40. <https://doi.org/10.1210/en.2003-0974>.
109. Jerlhag E, Egecioglu E, Landgren S, Salomé N, Heilig M, Moechars D, et al. Requirement of central ghrelin signaling for alcohol reward. *Proc Natl Acad Sci U S A*. 2009;106:11318–23. <https://doi.org/10.1073/pnas.0812809106>.
110. Abizaid A, Liu ZW, Andrews ZB, Shanabrough M, Borok E, Elsworth JD, et al. Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. *J Clin Invest*. 2006;116(12):3229–39. <https://doi.org/10.1172/JCI29867>.
111. Zigman JM, Nakano Y, Coppari R, Balthasar N, Marcus JN, Lee CE, et al. Mice lacking ghrelin receptors resist the development of diet-induced obesity. *J Clin Invest*. 2005;115(12):3564–72. <https://doi.org/10.1172/JCI26002>.
112. Holst B, Schwartz TW. Constitutive ghrelin receptor activity as a signaling set-point in appetite regulation. *Trends Pharmacol Sci*. 2004;25:113–7. <https://doi.org/10.1016/j.tips.2004.01.010>.
113. Petersen PS, Woldbye DPD, Madsen AN, Egerod KL, Jin C, Lang M, et al. In vivo characterization of high basal signaling from the ghrelin receptor. *Endocrinology*. 2009;150(11):4920–30. <https://doi.org/10.1210/en.2008-1638>.
114. Fu J, Gaetani S, Oveisi F, Lo Verme J, Serrano A, Rodríguez De Fonseca F, ... Piomelli D. Oleoylethanolamide regulates feeding and body weight through activation of the nuclear receptor PPAR- α . *Nature*. 2003;425(6953). Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/12955147>.
115. Fu J, Gaetani S, Oveisi F, Verme JL, Serrano A, Rodríguez De Fonseca F, et al. Oleoylethanolamide regulates feeding and body weight through activation of the nuclear receptor PPAR- α . *Nature*. 2003;425(6953):90–3. <https://doi.org/10.1038/nature01921>.
116. Tellez LA, Medina S, Han W, Ferreira JG, Licon-Limón P, Ren X, et al. A gut lipid messenger links excess dietary fat to dopamine deficiency. *Science*. 2013;341(6147):800–2. <https://doi.org/10.1126/science.1239275>.
117. Hankir MK, Seyfried F, Hintschich CA, Diep TA, Kleberg K, Kranz M, et al. Gastric bypass surgery recruits a gut PPAR- α -striatal D1R pathway to reduce fat appetite in obese rats. *Cell Metabol*. 2017;25(2):335–44. <https://doi.org/10.1016/j.cmet.2016.12.006>. **Discovered that OEA-PPAR- α signaling increase mesolimbic dopamine secretion.**
118. Bottin JH, Thomas EL, Balogun B, Bech PR, Ghatei MA, Moorthy K, et al. Changes in appetite, food intake, and appetite regulating hormones during acute weight loss induced by Roux-en-Y gastric bypass and low-calorie diet. *Obes Facts*. 2015;8:66–72. <https://doi.org/10.1159/000382140>.
119. Shin AC, Zheng H, Pistell PJ, Berthoud HR. Roux-en-Y gastric bypass surgery changes food reward in rats. *Int J Obes*. 2011;35(5):642–51. <https://doi.org/10.1038/ijo.2010.174>.
120. Habegger KM, Heppner KM, Amburgy SE, Ottaway N, Holland J, Raver C, et al. GLP-1R responsiveness predicts individual gastric bypass efficacy on glucose tolerance in rats. *Diabetes*. 2014;63(2):505–13. <https://doi.org/10.2337/db13-0511>.
121. Hayes MR, Schmidt HD. GLP-1 influences food and drug reward. *Curr Opin Behav Sci*. 2016;9:66–70.
122. Menzies JRW, Skibicka KP, Leng G, Dickson SL. Ghrelin, reward and motivation. *Endocr Dev*. 2013;25:101–11.
123. Abizaid A, Liu Z-W, Andrews ZB, Shanabrough M, Borok E, Elsworth JD, et al. Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. *J Clin Invest*. 2006;116:3229–39.
124. Skibicka KP, Hansson C, Alvarez-Crespo M, Friberg PA, Dickson SL. Ghrelin directly targets the ventral tegmental area to increase food motivation. *Neuroscience*. 2011;180:129–37.
125. Jerlhag E, Egecioglu E, Landgren S, Salome N, Heilig M, Moechars D, et al. Requirement of central ghrelin signaling for alcohol reward. *Proc Natl Acad Sci U S A*. 2009;106:11318–23.
126. Barkholt P, Pedersen PJ, Hay-Schmidt A, Jelsing J, Hansen HH, Vrang N. Alterations in hypothalamic gene expression following Roux-en-Y gastric bypass. *Mol Metab*. 2016;5:296–304.
127. Blum K, Bailey J, Gonzalez AM, et al. Neuro-genetics of reward deficiency syndrome (RDS) as the root cause of “addiction transfer”: a new phenomenon common after bariatric surgery. *J Genet Syndr Gene Ther*. 2011;2012.
128. Backman O, Stockeld D, Rasmussen F, Näslund E, Marsk R. Alcohol and substance abuse, depression and suicide attempts after Roux-en-Y gastric bypass surgery. *Br J Surg*. 2016;103:1336–42.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.